



Hands-on lab on structural variants



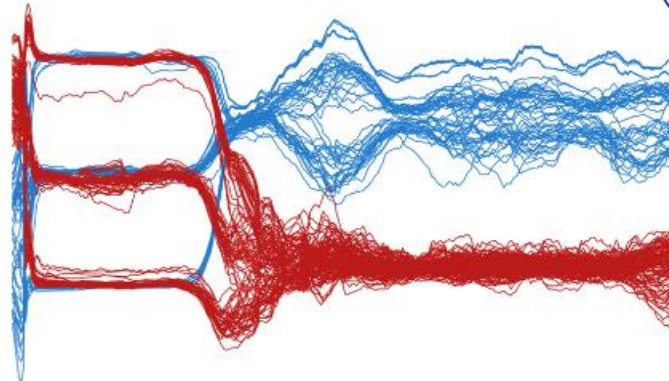
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EVOMICS 2026



**Many thanks to
Guy & the TAs,
Raja, Milos,
Flavia and
Rhianah!!!**



Tutorial

https://github.com/claimeerot/Tutorial_SV

IMPORTANT:

-> clone this repository in your working directory

-> open the folder Tutorial_SV

-> run everything from this location (and all paths should be ok!)

The screenshot shows the GitHub interface for the repository 'Tutorial_SV' by user 'claimeerot'. The repository is public and has 0 forks, 0 stars, and 0 watches. The main branch is 'main'. The repository contains several folders: '00_ressources', '01_pca_haploblocks', '02_assemblies', '03_LR', '04_SR', and '05_graphs'. The most recent commit is 'Update README.md' by 'claimeerot' 47 minutes ago. The repository description is 'A 1-day training about SV detection'. The repository also has a 'Readme' file, an 'Apache-2.0 license', and an 'Activity' tab. The 'Releases' section shows 'No releases published' and a link to 'Create a new release'.

claimeerot / Tutorial_SV

Type / to search

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Tutorial_SV Public

Pin Watch 0 Fork 0 Star 0

main 1 Branch 0 Tags

Go to file Add file Code

claimeerot Update README.md d8e5373 · 47 minutes ago 55 Commits

00_ressources	Add files via upload	3 days ago
01_pca_haploblocks	Update README.md	2 days ago
02_assemblies	Update README.md	4 days ago
03_LR	Update README.md	2 days ago
04_SR	Update README.md	1 hour ago
05_graphs	Update README.md	47 minutes ago

About

A 1-day training about SV detection

- Readme
- Apache-2.0 license
- Activity
- 0 stars
- 0 watching
- 0 forks

Releases

No releases published

[Create a new release](#)

Outline

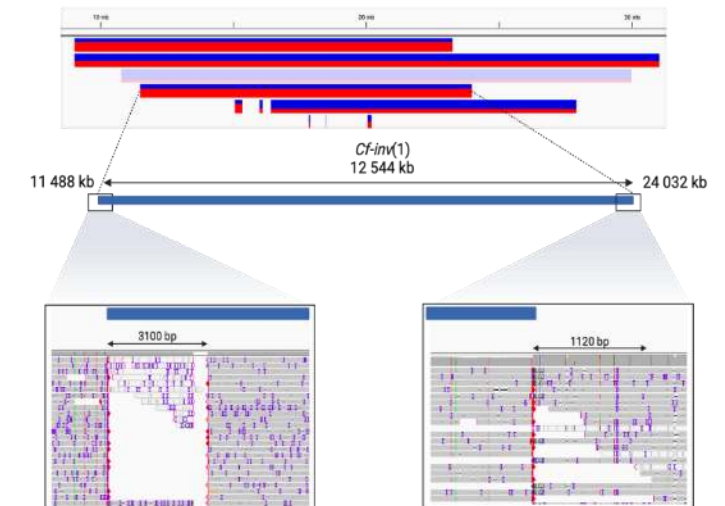
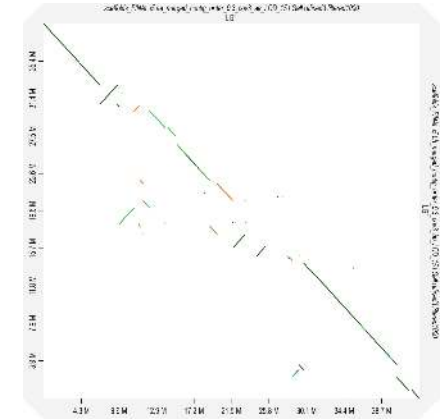
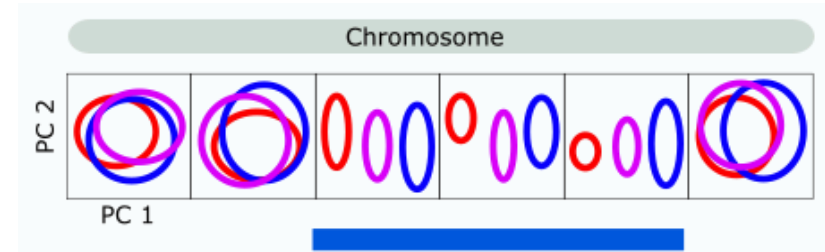
01 - Using SNPs & local PCA to detect haploblocks putatively representing large rearrangements

02 - Comparing assemblies for large rearrangements

03 - Assessing breakpoints and detecting other SVs with long-reads

04 - Using population dataset of short-reads to detect SVs

05 - Towards graph-based analysis



Data



Seaweed flies genomes
(*Coelopa frigida*)

Assemblies

-> 2 genomes
(ref.fasta)
(alt.fasta)

Subset of 10Mb

Long-reads

-> 1 sample (fri59)
ONT – Oxford
Nanopore
~20X

Short-reads

-> 22 samples (illumina PE 150bp)
~10X

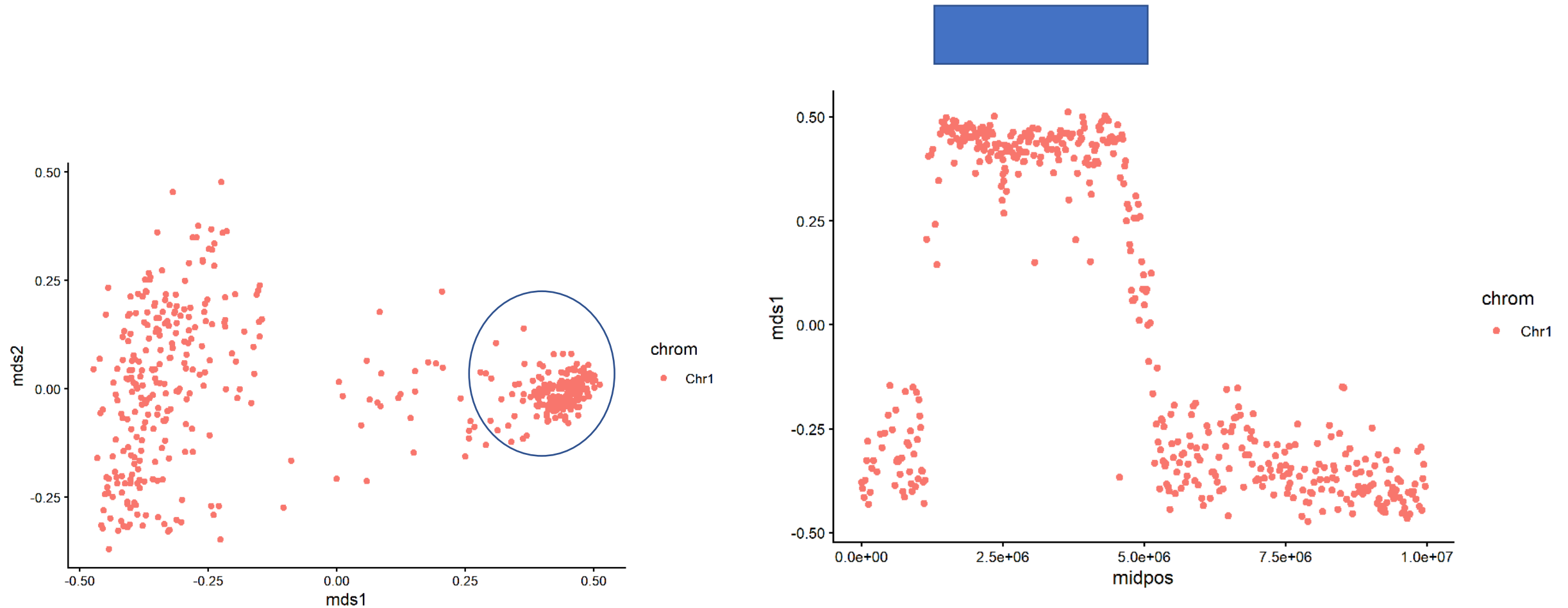
1 pair of fastq reads (A_R1, B_R2)

4 bamfiles (A, B, C, D)

1 vcf of SNPs from the 22 samples
(A to V)

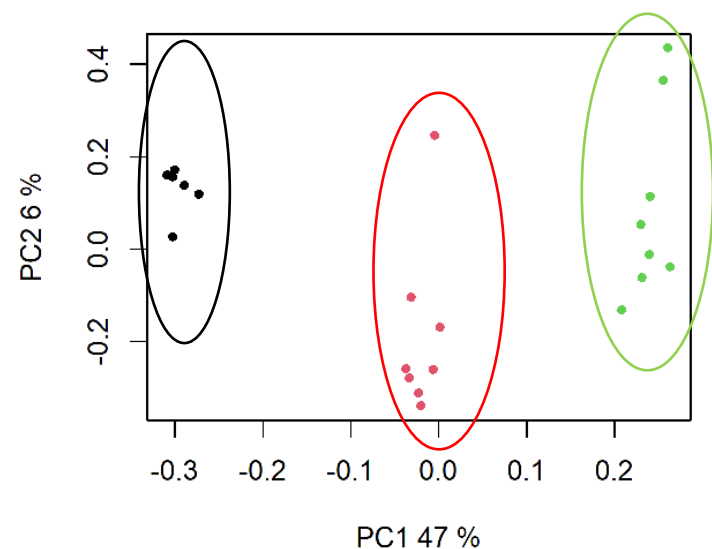
I - PCAs

What do you think? Do we have a region where we could suspect non-recombining blocks revealing a putative inversion? How could we do to genotype our individuals for this putative variant?



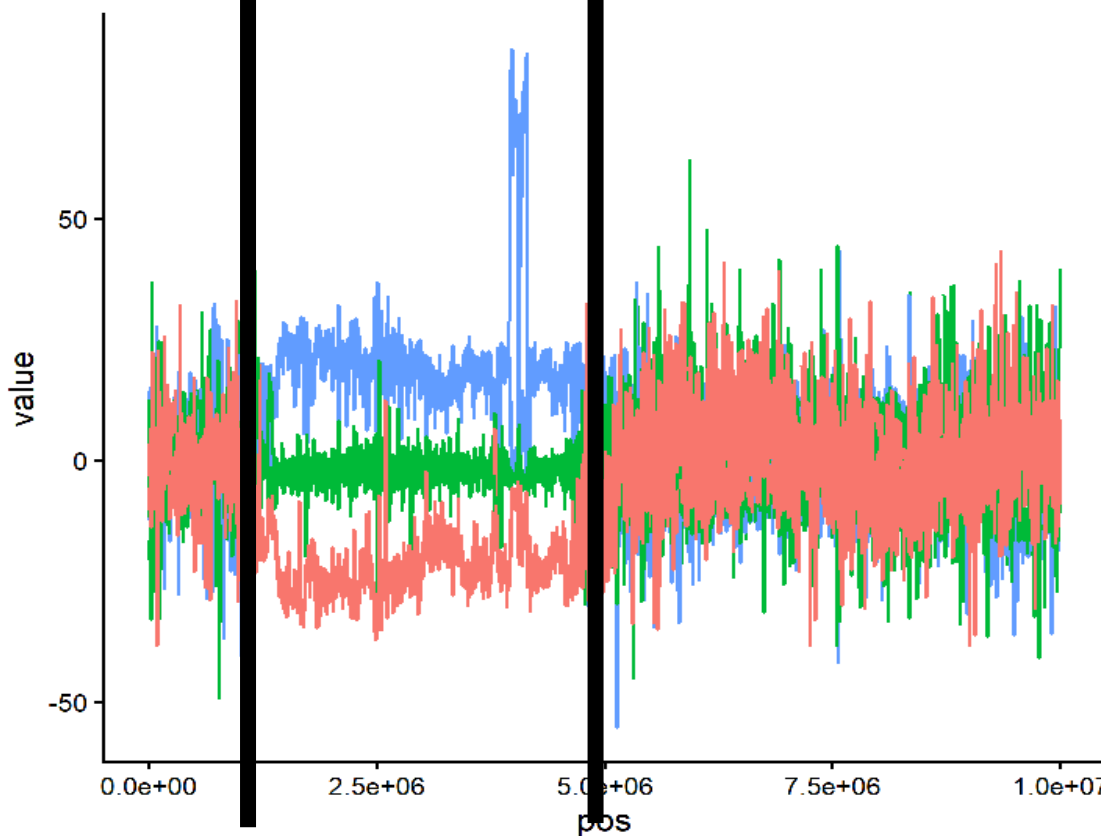
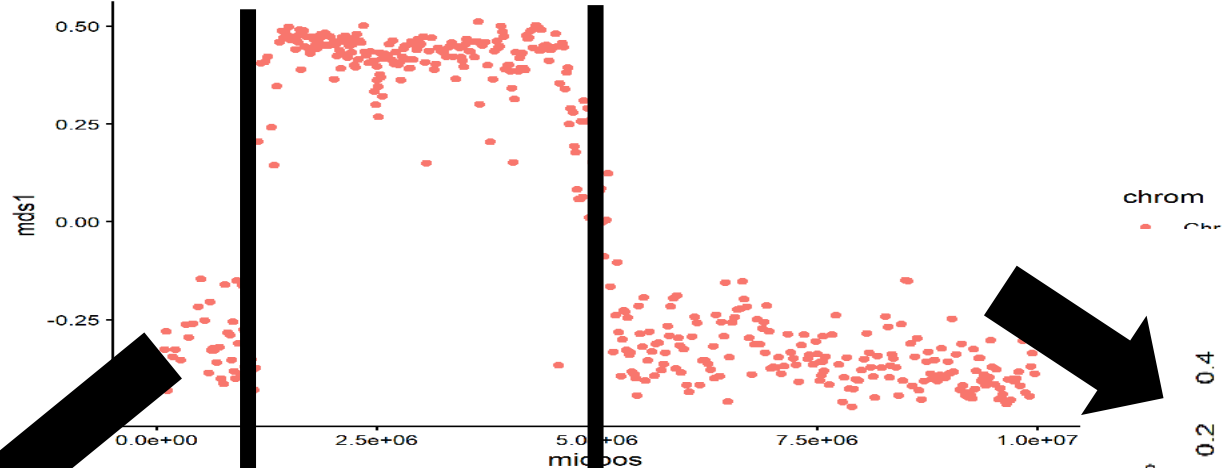
I - PCAs

Chr1_2144736

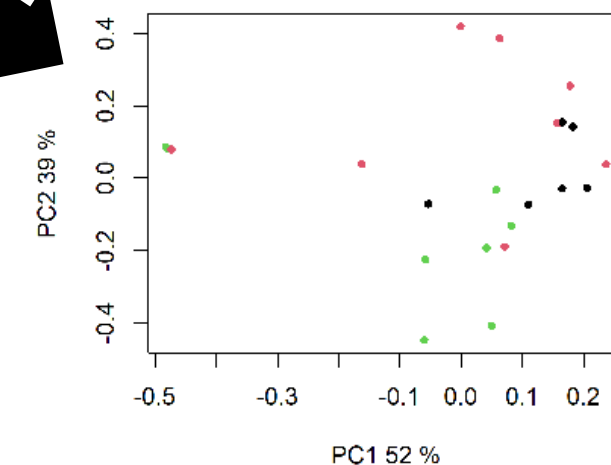


Using either information from lostruct or winpca, may you identify approximative coordinates for the suspected rearrangement?

from 1Mb to 4,5Mb



Chr1_7238135



factor(haplogroup)

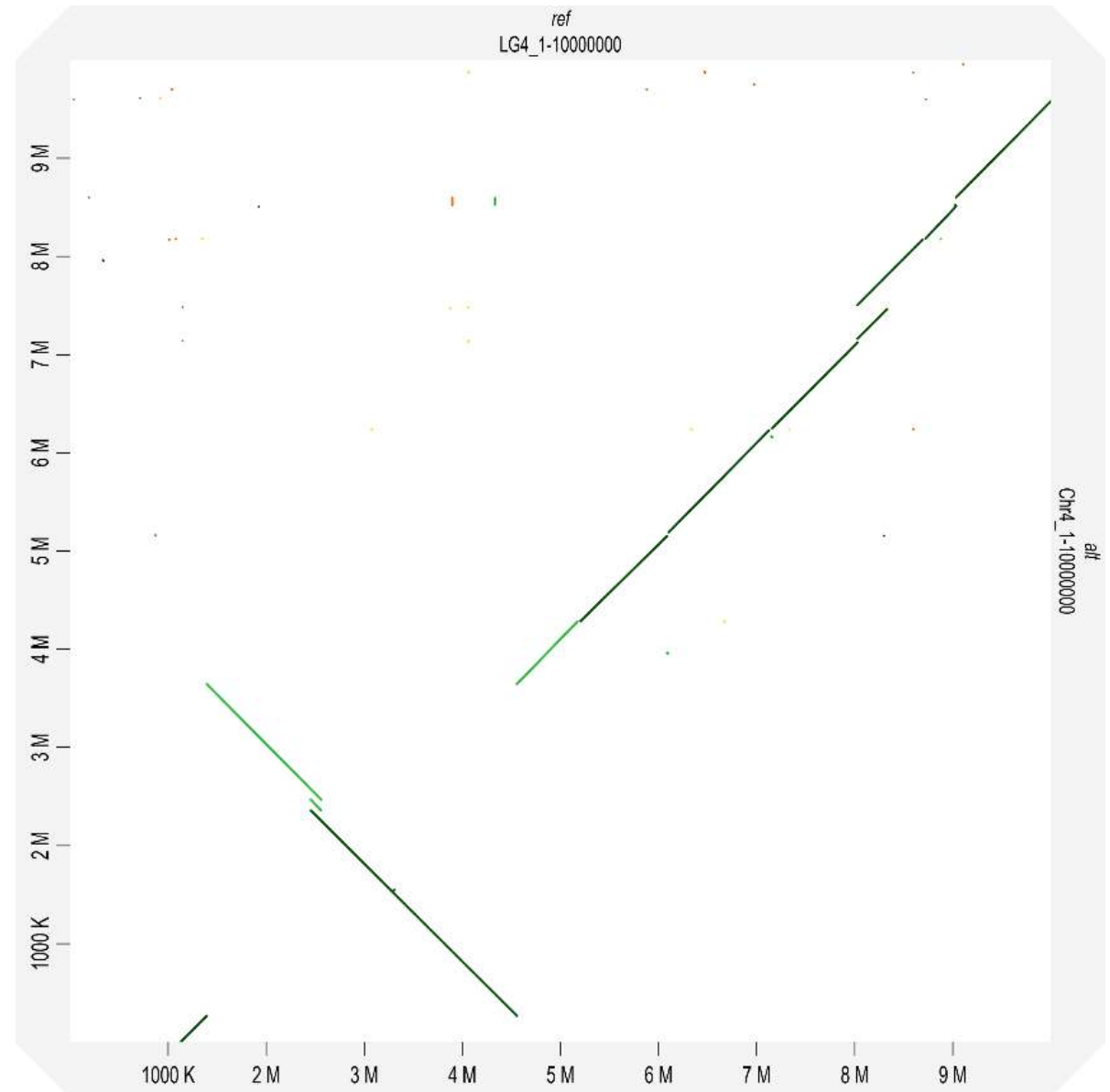
AA
AB
BB

2 – Assemblies comparison

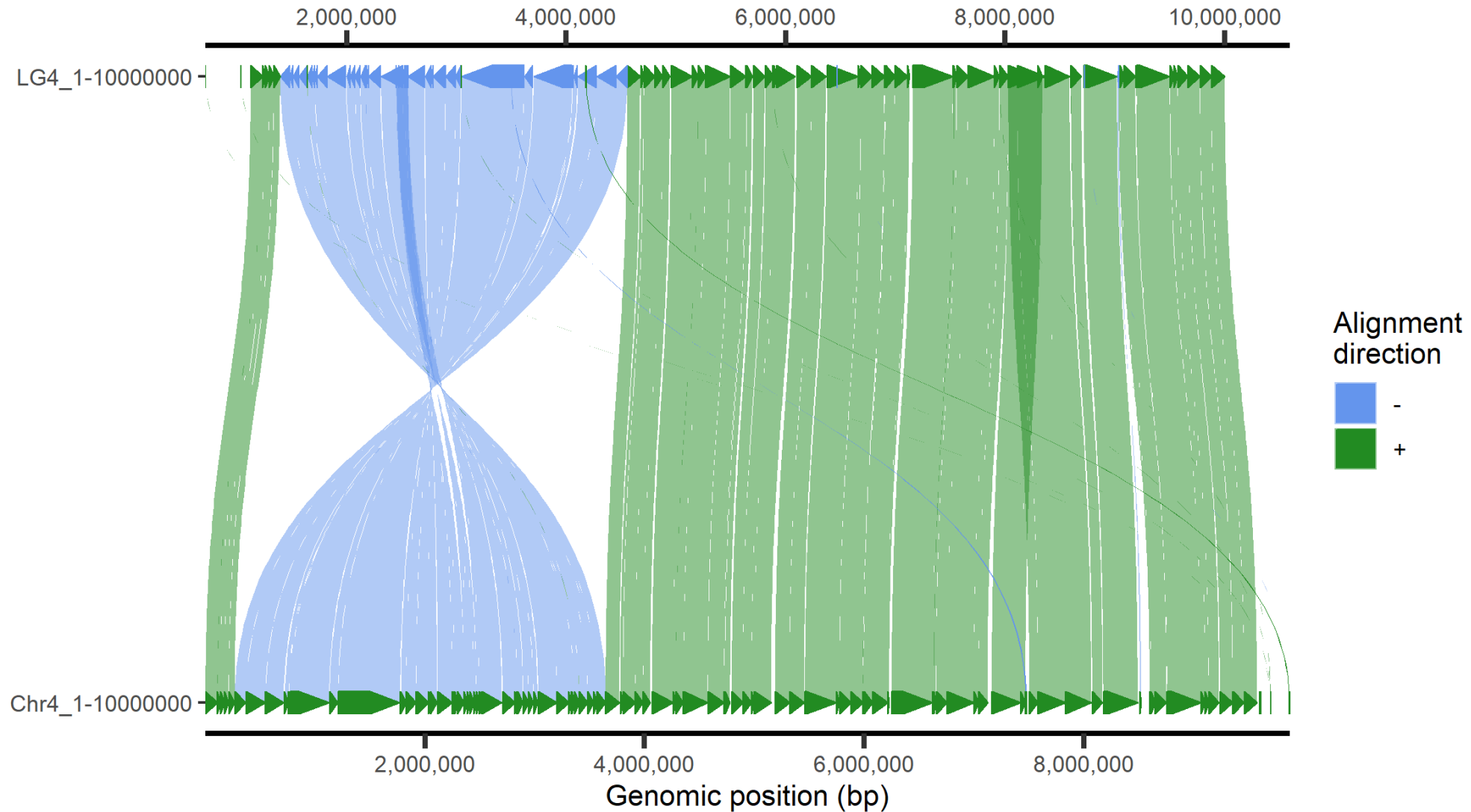
https://dgenies.toulouse.inra.fr/result/7EvbZ_2026011921013_1

What do you think? Do you visualise some rearrangements?
Does it fit what you suspected from the PCA analysis?

Yes, a large inversion from 1Mb to 4,5Mb



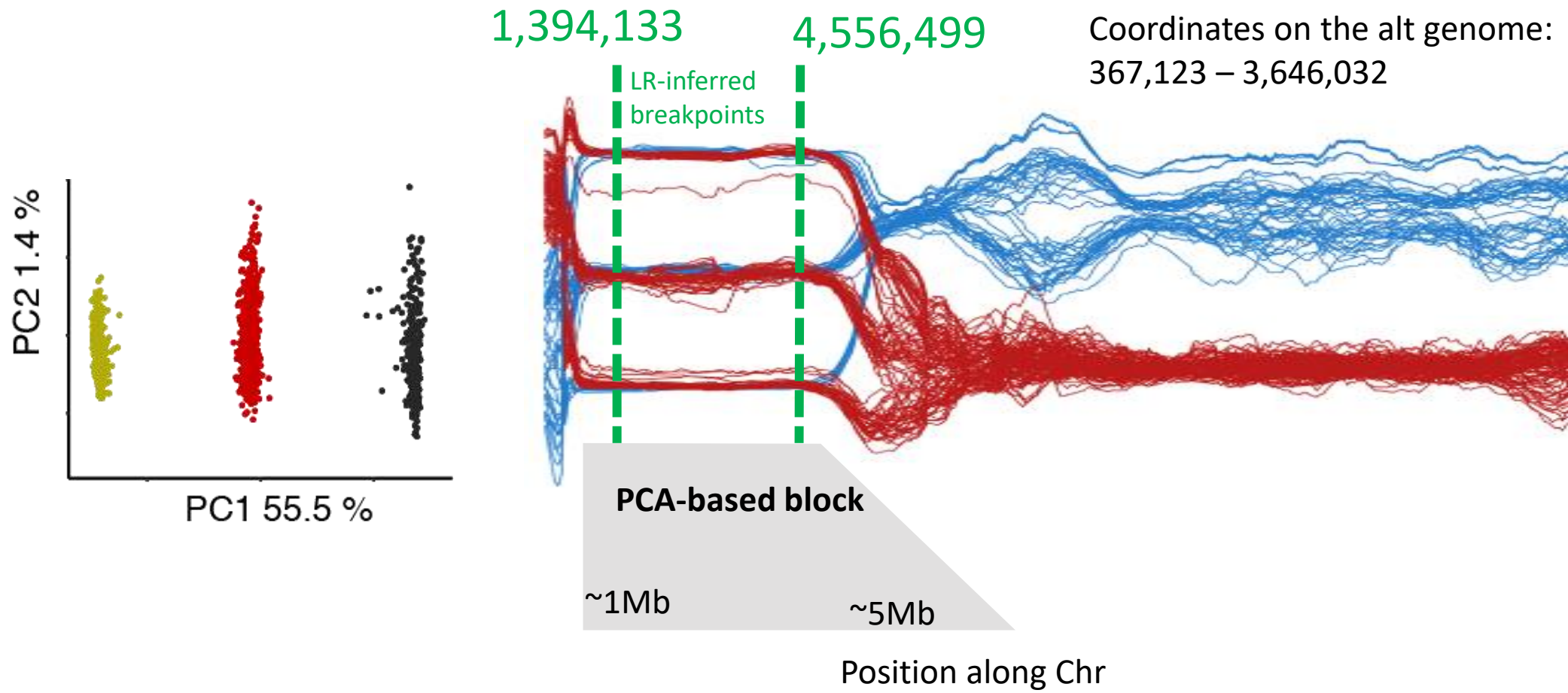
2 – Assemblies comparison



2 – Assemblies comparison

Using the paf file and/or the R package, can you extract the breakpoints of the suspected rearrangement?

How does it compare to the putative breakpoints identified by indirect PCA methods based on recombination suppression?



3 –Using long-reads

1845

- > How many SVs were detected?
- > What are the different pieces of information available for each variant?
- > How does it differ from a vcf of SNPs?
- > What kind of filters or classifications may you want to implement for subsequent analysis?

```
##FILTER=<ID=MOSAIC_VAF,Description="Mosaic variant allele fraction filter">
##FILTER=<ID=NOT_MOSAIC_VAF,Description="Variant allele fraction filter for non-mosaic">
##FILTER=<ID=ALN_NM,Description="Length adjusted mismatch filter">
##FILTER=<ID=STRAND_BND,Description="Strand support filter for BNDs">
##FILTER=<ID=STRAND,Description="Strand support filter for germline SVs">
##FILTER=<ID=STRAND_MOSAIC,Description="Strand support filter for mosaic SVs">
##FILTER=<ID=SVLEN_MIN,Description="SV length filter">
##FILTER=<ID=SVLEN_MIN_MOSAIC,Description="SV length filter for mosaic SVs">
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##INFO=<ID=IMPRECISE,Number=0,Type=Flag,Description="Structural variation with imprecise breakpoints">
##INFO=<ID=MOSAIC,Number=0,Type=Flag,Description="Structural variation classified as putative mosaic">
##INFO=<ID=SVLEN,Number=1,Type=Integer,Description="Length of structural variation">
##INFO=<ID=SVTYPE,Number=1,Type=String,Description="Type of structural variation">
##INFO=<ID=CHR2,Number=1,Type=String,Description="Mate chromosome for BND SVs">
##INFO=<ID=SUPPORT,Number=1,Type=Integer,Description="Number of reads supporting the structural variation">
##INFO=<ID=SUPPORT_INLINE,Number=1,Type=Integer,Description="Number of reads supporting an INS/DEL SV (non-split events only)">
##INFO=<ID=SUPPORT_LONG,Number=1,Type=Integer,Description="Number of soft-clipped reads putatively supporting the long insertion SV">
##INFO=<ID=END,Number=1,Type=Integer,Description="End position of structural variation">
##INFO=<ID=STDEV_POS,Number=1,Type=Float,Description="Standard deviation of structural variation start position">
##INFO=<ID=STDEV_LEN,Number=1,Type=Float,Description="Standard deviation of structural variation length">
##INFO=<ID=COVERAGE,Number=.,Type=Float,Description="Coverages near upstream, start, center, end, downstream of structural variation">
##INFO=<ID=STRAND,Number=1,Type=String,Description="Strands of supporting reads for structural variant">
##INFO=<ID=AC,Number=.,Type=Integer,Description="Allele count, summed up over all samples">
##INFO=<ID=SUPP_VEC,Number=1,Type=String,Description="List of read support for all samples">
##INFO=<ID=CONSENSUS_SUPPORT,Number=1,Type=Integer,Description="Number of reads that support the generated insertion (INS) consensus sequence">
##INFO=<ID=RNAME,Number=.,Type=String,Description="Names of supporting reads (if enabled with --output-rnames)">
##INFO=<ID=VAF,Number=1,Type=Float,Description="Variant Allele Fraction">
##INFO=<ID=NM,Number=.,Type=Float,Description="Mean number of query alignment length adjusted mismatches of supporting reads">
##INFO=<ID=PHASE,Number=.,Type=String,Description="Phasing information derived from supporting reads, represented as list of: HAPLOTYPE,PHASESET,HAPLOTYPE_SUPPORT,PHASESET_SUPPORT,HAPLOTYPE_FILTER,PHASESET_FILTER">
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT SAMPLE
Chr1 3564 Sniffles2.DEL.11A7S0 ACTATAATTTCTTTCAATAGTTATTTCAATTTATTTCAATATTAAATATATATAAAATACATTTTTTTAATTTTTGTTTATTATTAATAATATGTAATAAG A
60 PASS PRECISE;SVTYPE=DEL;SVLEN=-103;END=3666;SUPPORT=27;COVERAGE=44,43,43,43,42;STRAND=+-;STDEV_LEN=0.000;STDEV_POS=0.000;VAF=0.628 GT:GQ
:DR:DV 0/1:60:16:27
Chr1 5716 Sniffles2.DEL.11A8S0 TACACATTTTCTAAAAGCTGATTAATTTCTCTTTAAATTGATATATATATGTCATATTATATATAGAATGAA T 60 PASS PRECISE;SVTYPE
E=DEL;SVLEN=-72;END=5787;SUPPORT=16;COVERAGE=40,37,37,40,38;STRAND=+-;STDEV_LEN=0.000;STDEV_POS=0.421 GT:GQ:DR:DV 0/1:60:22:16
Chr1 13302 Sniffles2.INS.5S0 C CTTTAGATCAATTATTTGAGAATTGCAATTATTTGAGAAATTATTTTGAAGCTAAAAA 60 PASS PRECISE;SVTYPE=INS;SVLEN=57;E
ND=13302;SUPPORT=20;COVERAGE=35,36,36,37,37;STRAND=+-;STDEV_LEN=0.000;STDEV_POS=0.000;SUPPORT_LONG=0;VAF=0.556 GT:GQ:DR:DV 0/1:60:16:20
Chr1 36928 Sniffles2.INS.FS0 A ATTTAAGGTTGTTGAAAATTTGCTTGAATTGTTGAAATTTTGTAGAGGTCGTCACGAATTAACGTCTTTTTTCGCGAAATATCAGGTATTTAGATTTCCTAAAAAATTG
CCTGTATTATGAAATTTTCGAAAATTTAAATGTCGTCACGAATTAACGTTCTTTTCGCGAAATAACCGCTATTACGCGAGATATGAACGATATAAGATTTTCGAAAATTTTCGGAGCTGTAATTTAAAAAAAATTTAAGGGCGCTCAGCAATTAA
CGTATTT 60 PASS IMPRECISE;SVTYPE=INS;SVLEN=279;END=36928;SUPPORT=37;COVERAGE=39,39,39,40,38;STRAND=+-;STDEV_LEN=45.889;STDEV_POS=0.000;SUPPORT_LONG=0
;VAF=0.949 GT:GQ:DR:DV 1/1:60:2:37
Chr1 53053 Sniffles2.INS.13S0 T TTTTCGATATTTTGTCAATTATTTTTTTTGTTCATTTTTTTTATAAAAATTTTC 60 PASS PRECISE;SVTYPE=INS;SVLEN=56;E
ND=53053;SUPPORT=22;COVERAGE=48,46,46,46,45;STRAND=+-;STDEV_LEN=0.900;STDEV_POS=16.908;SUPPORT_LONG=0;VAF=0.478 GT:GQ:DR:DV 0/1:60:24:22
Chr1 55215 Sniffles2.INS.16S0 T TAAAAATGTTTATGATTCCTTAAAAACCTCTGTCGCTGAGATTCCCTAAGACATCGTTTGACCCCG 60 PASS PRECISE;SVTYPE=INS;SV
:
```

3 –Using long-reads

1845

- > How many SVs were detected?
- > What are the different pieces of information available for each variant?
- > How does it differ from a vcf of SNPs?
- > What kind of filters or classifications may you want to implement for subsequent analysis?

```
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT SAMPLE
Chr1 3564 Sniffles2.DEL.11A7S0 ACTATAATTTTCTTTCAATAGTTATTCAATTTATTTCAATATTAAATTATATAAAAAATACATTTTTTTTAATTTTTTGTTTATTTAAAAAATATGTAAATAAG A
60 PASS PRECISE;SVTYPE=DEL;SVLEN=-103;END=3666;SUPPORT=27;COVERAGE=44,43,43,43,42;STRAND=+-;STDEV_LEN=0.000;STDEV_POS=0.000;VAF=0.628 GT:GQ
:DR:DV 0/1:60:16:27
Chr1 5716 Sniffles2.DEL.11A8S0 TACACATTTTCTAAAAGCTGATTAAATTCCTTTAAATTGATATATATATATGTCATATTTATATATAGAAATGAA T 60 PASS PRECISE;SVTYPE=DEL;SVLEN=-72;END=5787;SUPPORT=16;COVERAGE=40,37,37,40,38;STRAND=+-;STDEV_LEN=0.000;STDEV_POS=0.000;VAF=0.421 GT:GQ:DR:DV 0/1:60:22:16
Chr1 13302 Sniffles2.INS.5S0 C CTTTAGATCAATTATTTGAGAATTTGATTATTTGAGAAATTATTATTTGAACTAAAAA 60 PASS PRECISE;SVTYPE=INS;SVLEN=57;END=13302;SUPPORT=20;COVERAGE=35,36,36,37,37;STRAND=+-;STDEV_LEN=0.000;STDEV_POS=0.000;SUPPORT_LONG=0;VAF=0.556 GT:GQ:DR:DV 0/1:60:16:20
Chr1 36928 Sniffles2.INS.FS0 A ATTTAAGGTTGTTGAAAATTTGCTTGAATTGTGAAATTTTTGTAGAGGTCGTCACGAATTAACGTCTTTTTTCGCGAAATATCACGTATTTAGATTTCTAAAAAATTG
CCTGTATTATGAAATTTTCGAAAATTTAAATGTCGTCACGAATTAACGTCTTTTTTCGCGAAATTAACGCGTATTTACGGAGATATGAACGATATAAGATTTGAAAAATTTCTGGAGCTGTAAATTTAAAAAAAATTTAAGGGCGTCACGAATTAA
CGTATTT 60 PASS IMPRECISE;SVTYPE=INS;SVLEN=279;END=36928;SUPPORT=37;COVERAGE=39,39,39,40,38;STRAND=+-;STDEV_LEN=45.889;STDEV_POS=0.000;SUPPORT_LONG=0
;VAF=0.949 GT:GQ:DR:DV 1/1:60:2:37
Chr1 53053 Sniffles2.INS.13S0 T TTTTCGATATTTTGTCAATTATTTTTTTTGTTTTCATTTTTTTTATAAAAATTTTC 60 PASS PRECISE;SVTYPE=INS;SVLEN=56;END=53053;SUPPORT=22;COVERAGE=48,46,46,46,45;STRAND=+-;STDEV_LEN=0.900;STDEV_POS=16.908;SUPPORT_LONG=0;VAF=0.478 GT:GQ:DR:DV 0/1:60:24:22
Chr1 55215 Sniffles2.INS.16S0 T TAAAAATGTTTATGATTCCCTAAAAAACCTCTTGTCGGTGAGATTCCCTAAGACATCGTTTGAACCCCG 60 PASS PRECISE;SVTYPE=INS;SVLEN=56;END=55215;SUPPORT=22;COVERAGE=48,46,46,46,45;STRAND=+-;STDEV_LEN=0.900;STDEV_POS=16.908;SUPPORT_LONG=0;VAF=0.478 GT:GQ:DR:DV 0/1:60:24:22
```

- > select SV based on length range?
- > filter by type
- > remove SVs that do not « PASS »
- > filter based on read support
- > check IMPRECISE breakpoints....

3 –Using long-reads

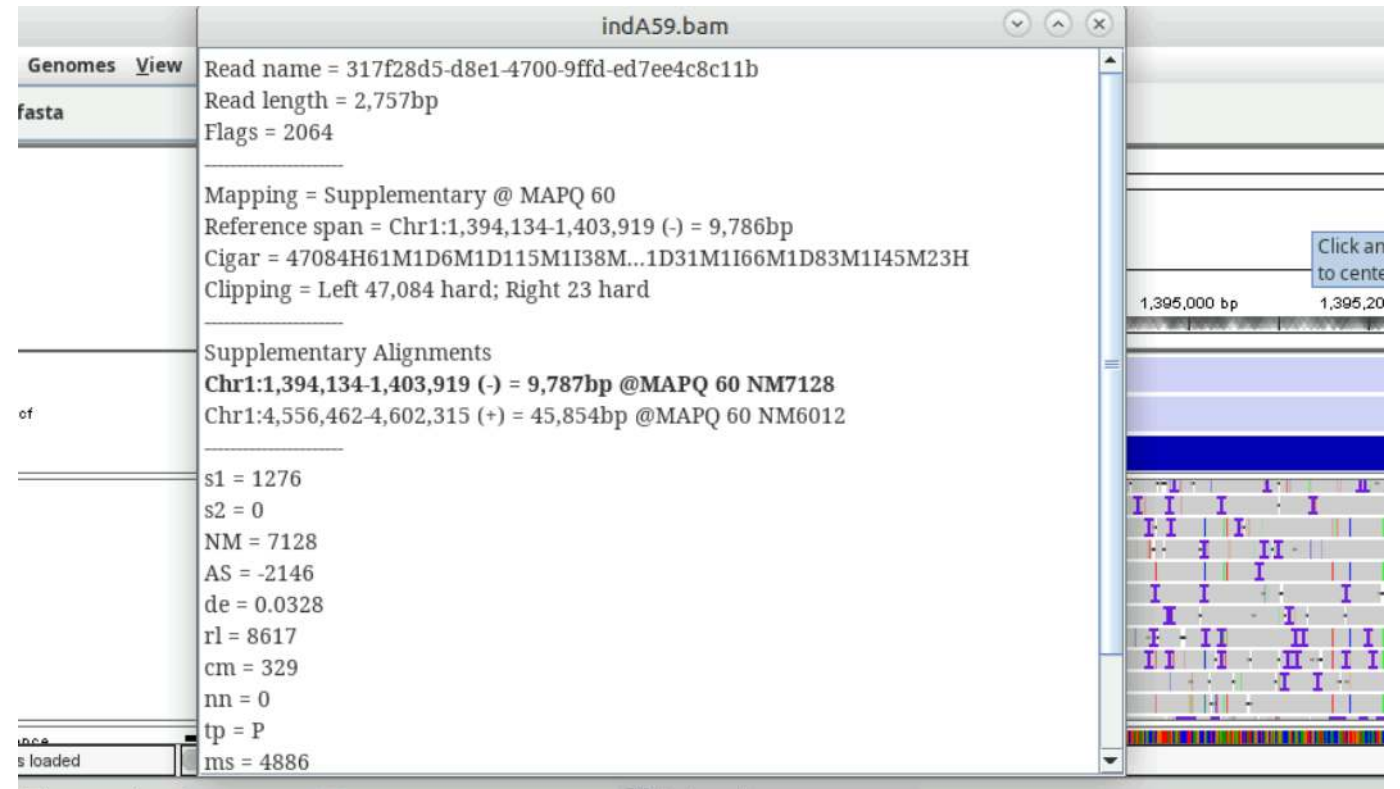


3 –Using long-reads

You can also zoom on the breakpoints previously identified with the assembly comparison for the very large rearrangement.

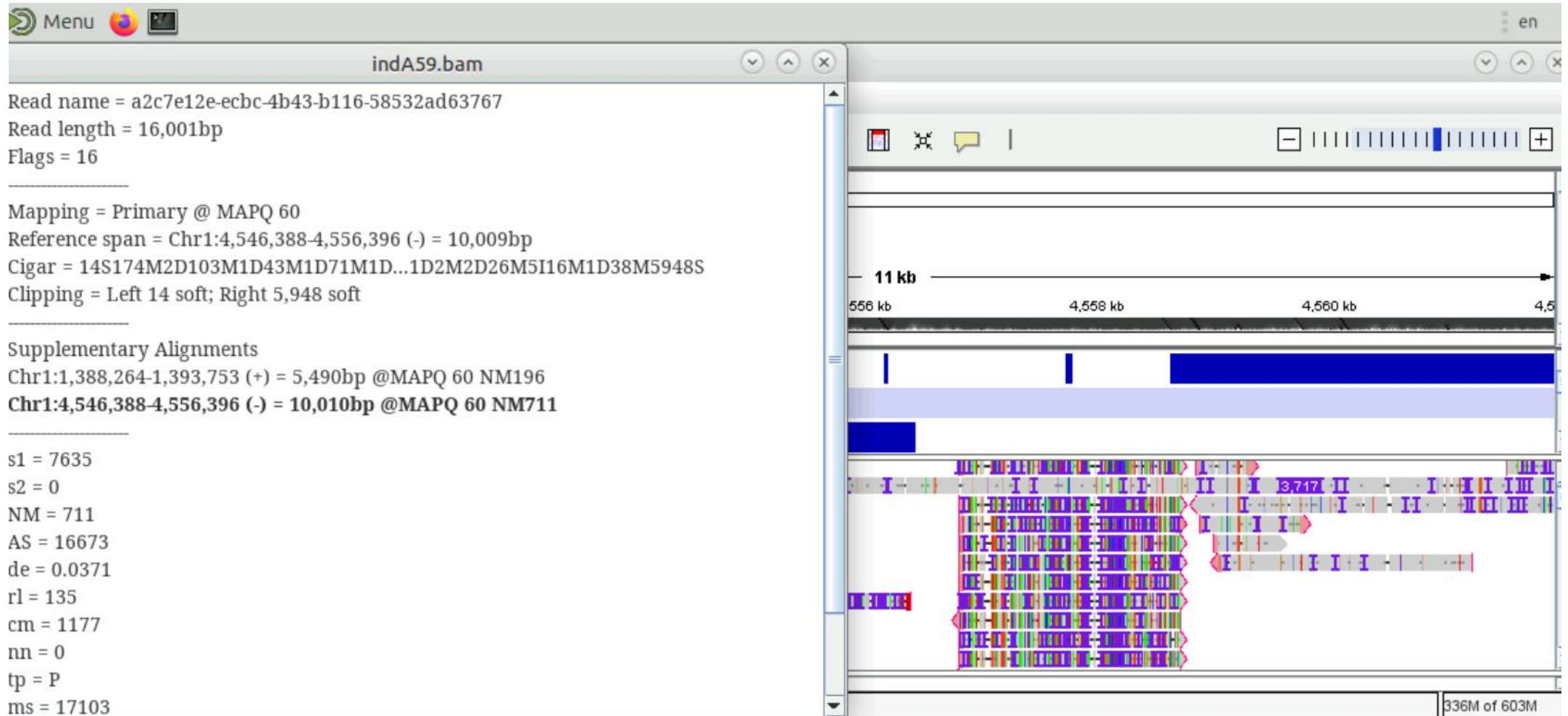
-> What do they look like here?

-> Are they supported by long-reads?



3 –Using long-reads

-> Which genotype is this individual "fri59" for our big inversion?



3 –Using long-reads

-> Which genotype is this individual "fri59" for our big inversion?

heterozygote

```
grep 'INV.22BAS0' 03_LR/fri59.vcf
```

```
Chr1 1394133 Sniffles2.INV.22BAS0 A <INV> 60 PASS  
PRECISE;SVTYPE=INV;SVLEN=3162327;END=4556460;SUPPORT=15;COVERAGE=34,34,40,24,39;STRAND=+;STDEV_  
LEN=0.000;STDEV_POS=0.000;VAF=0.341 GT:GQ:DR:DV 0/1:60:29:15
```

Also visible with the reads in IGV (some go over the break-points and some are split-reads)

4 – SV call with short-reads

-> What do you think about the information about SVs?
What differs and does not differ with the vcf produced by Sniffles?

```
##bcftools viewCommand=view 04_SR/allSV_SR.bcf; Date=Fri Jan 16 09:35:56 2026
#CHROM POS ID REF ALT QUAL FILTER INFO FORMAT B
Chr1 519 DEL00000000 GCTTAACACAGTTCTCTTTCTTAATGTTTCATTT G 480 PASS PRECISE;SVTYPE=DEL;SVMETHOD=EMBL.DELLYv1.7.2;END=552;PE=0;MAP
Q=0;CT=3to5;CIPOS=-4,4;CIEND=-4,4;SRMAPQ=60;INSLEN=0;HOMLEN=4;SR=8;SRQ=0.965347;CONSENSUS=TATTTTTTTTCATTTCCCGACCTTCCGTCATTATTATATTACAATCTCTGCTTGTTCCCGAGCAGCAT
TTTCTGGACGATAGTGAAAACCTTCCTAAGGAATCAATCCTTTAATCTGCTTTTTCAAATATTTTAATTTTGAAAATTGTCTTCTTTTTTATTATTTTCTCACAGTTTTTATCATTCTTTATTCTTTTCTTGCAA;CE=1.78796;CONSBP=116
GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-136.067,0,-26.8585:10000:PASS:626:862:1443:1:0:0:14:27
Chr1 958 INS000000001 A ATCTCGCTCTATCTCGCTCTATCTCGCTCTCTCTC 240 PASS PRECISE;SVTYPE=INS;SVMETHOD=EMBL.DELLYv1.7.2;END=958;SVLEN=34
;PE=0;MAPQ=0;CT=NtoN;CIPOS=-9,9;CIEND=-9,9;SRMAPQ=60;INSLEN=34;HOMLEN=12;SR=4;SRQ=0.978723;CONSENSUS=AAC TGGGAAATTGTGAGATATTTAAATGCGACTATTAATACGTTTAAACGCTCTAT
CTCGCTCTATCTCGCTCTATCTCGCTCTCTCTCTCTCTCTCTCTCACTCTTCACTCATTATTTTTTTATATCTTTTGTTTATTTCATTGGGGAGAGTTTTTGAATTTTCATTGCTTCTTG;CE=1.83651;CONSBP=55 GT:GL:GQ:FT:R
CL:RC:RCR:RDCN:DR:DV:RR:RV 1/1:-248.998,-14.748,0:147:PASS:23298:47469:24171:2:0:0:0:49
Chr1 5356 DUP00000002 G <DUP> 95 LowQual IMPRECISE;SVTYPE=DUP;SVMETHOD=EMBL.DELLYv1.7.2;END=180472;PE=2;MAPQ=59;CT=5to3;CIPOS=-529,529
;CIEND=-529,529 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/0:0,-10.0595,-348.193:101:PASS:2476:81500:46802:3:63:2:0:0
Chr1 5356 DUP00000003 G <DUP> 520 PASS IMPRECISE;SVTYPE=DUP;SVMETHOD=EMBL.DELLYv1.7.2;END=826732;PE=18;MAPQ=28;CT=5to3;CIPOS=-428,42
8;CIEND=-428,428 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-40.5413,0,-188.637:10000:PASS:2476:691588:632470:2:35:22:0:0
Chr1 5382 INV00000004 G <INV> 267 PASS IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=1021738;PE=10;MAPQ=29;CT=5to5;CIPOS=-375,3
75;CIEND=-375,375 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-31.3596,0,-146.018:10000:PASS:2479:1063929:385871:5:29:24:0:0
Chr1 5383 INV00000005 A <INV> 1251 PASS IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=616271;PE=38;MAPQ=30;CT=5to5;CIPOS=-311,31
1;CIEND=-311,311 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-90.5894,0,-495.263:10000:PASS:2479:415898:463178:2:129:51:0:0
Chr1 5383 DUP00000006 A <DUP> 1657 PASS IMPRECISE;SVTYPE=DUP;SVMETHOD=EMBL.DELLYv1.7.2;END=616734;PE=55;MAPQ=28;CT=5to3;CIPOS=-168,16
8;CIEND=-168,168 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-173.782,0,-156.21:10000:PASS:2479:420550:459580:2:32:83:0:0
Chr1 5383 DUP00000007 A <DUP> 373 PASS IMPRECISE;SVTYPE=DUP;SVMETHOD=EMBL.DELLYv1.7.2;END=617527;PE=15;MAPQ=24;CT=5to3;CIPOS=-362,36
2;CIEND=-362,362 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-29.2598,0,-120.299:10000:PASS:2479:422663:458989:2:22:17:0:0
Chr1 5383 INV00000008 A <INV> 181 PASS IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=1068736;PE=7;MAPQ=26;CT=5to5;CIPOS=-642,64
2;CIEND=-642,642 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/0:0,-6.88546,-437.908:69:PASS:2479:1152758:338466:7:76:7:0:0
Chr1 5383 INV00000009 A <INV> 120 LowQual IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=3052463;PE=2;MAPQ=60;CT=5to5;CIPOS=-601,60
1;CIEND=-601,601 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-6.57117,0,-135.602:66:PASS:2479:2103493:567688:7:24:4:0:0
Chr1 5387 INV00000010 A <INV> 157 PASS IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=4283625;PE=3;MAPQ=53;CT=5to5;CIPOS=-616,61
6;CIEND=-616,616 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-9.37838,0,-101.678:94:PASS:2479:2552441:827275:6:18:3:0:0
Chr1 5414 INV00000011 T <INV> 223 PASS IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=871200;PE=8;MAPQ=29;CT=5to5;CIPOS=-167,167
;CIEND=-167,167 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-21.6656,0,-151.094:10000:PASS:2516:769323:576244:3:29:18:0:0
Chr1 5436 DUP00000012 A <DUP> 70 LowQual IMPRECISE;SVTYPE=DUP;SVMETHOD=EMBL.DELLYv1.7.2;END=204296;PE=2;MAPQ=35;CT=5to3;CIPOS=-616,616
;CIEND=-616,616 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/0:0,-2.61808,-162.885:26:PASS:2603:102684:32982:6:30:2:0:0
Chr1 5494 INV00000013 T <INV> 51 LowQual IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=1010012;PE=2;MAPQ=29;CT=3to3;CIPOS=-563,56
3;CIEND=-563,563 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/0:0,-17.778,-419.003:10000:PASS:2994:1020499:419643:5:74:2:0:0
Chr1 5497 DEL00000014 A <DEL> 138 PASS IMPRECISE;SVTYPE=DEL;SVMETHOD=EMBL.DELLYv1.7.2;END=5278163;PE=4;MAPQ=34;CT=3to5;CIPOS=-542,54
2;CIEND=-542,542 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-5.08503,0,-129.371:51:PASS:3007:2954992:863025:7:25:4:0:0
```

Weird co-located SVs?

Lack of alt sequence for most variants

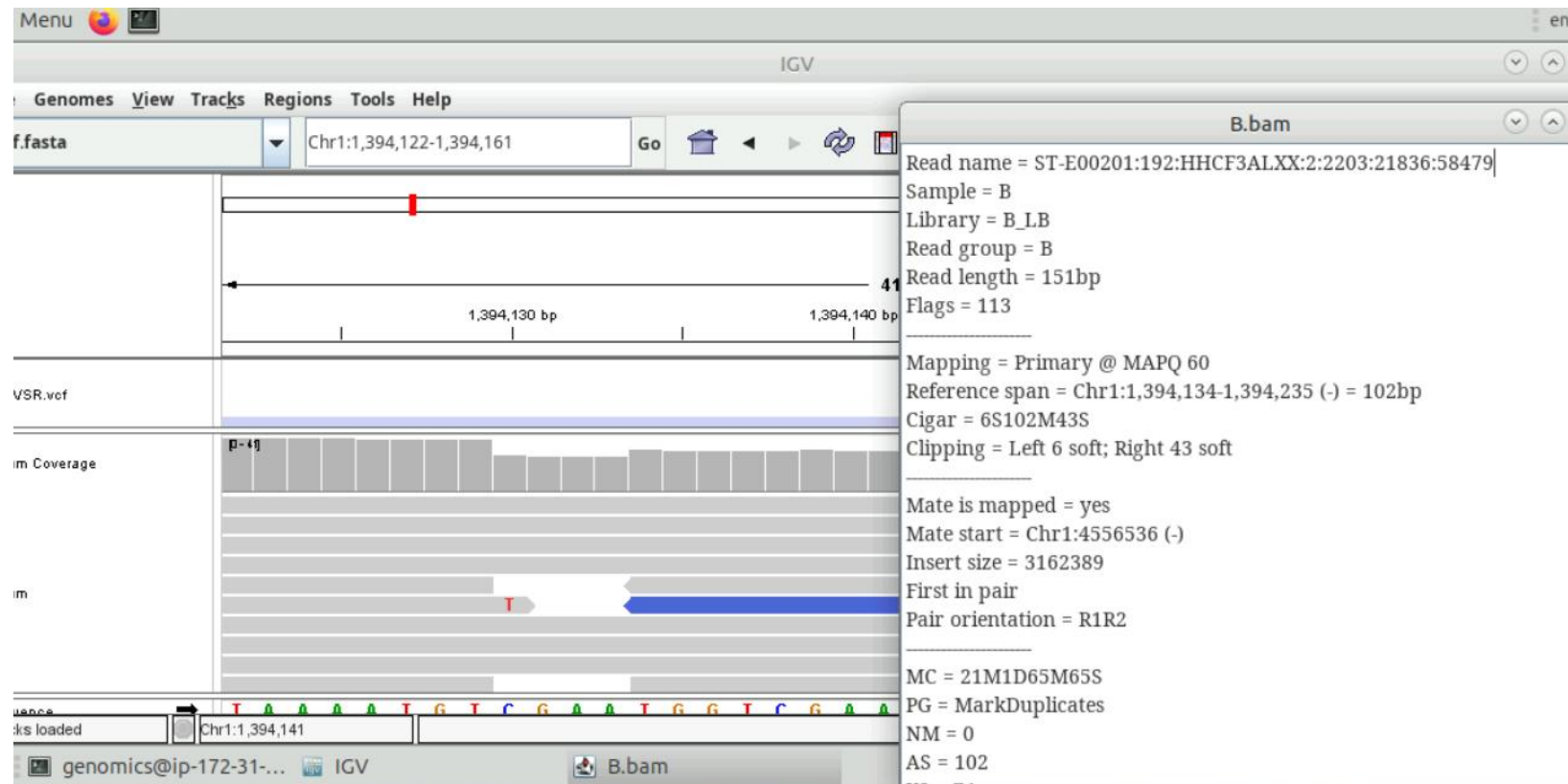
No SVLEN

4 – SV call with short-reads

-> What do you think about our large inversion? Was it detected by Delly?

```
Chr1 1393753 INV00011549 A <INV> 2120 PASS IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=4556341;PE=39;MAPQ=60;
CT=5to5;CIPOS=-50,50;CIEND=-50,50 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 1/1:-98.1971,-5.41565,0:54:PASS:1932910:1817073:901901:1:
0:18:0:0 0/1:-14.3866,0,-50.0868:144:PASS:1384270:1282649:621820:1:9:4:0:0 0/1:-10.7753,0,-118.775:108:PASS:1939344:1999980:
1008238:1:21:3:0:0 1/1:-100.1,-5.41844,0:54:PASS:1655494:1489965:724297:1:0:18:0:0
Chr1 1394089 INV00011550 G <INV> 1829 PASS IMPRECISE;SVTYPE=INV;SVMETHOD=EMBL.DELLYv1.7.2;END=4556153;PE=37;MAPQ=60;
CT=3to3;CIPOS=-163,163;CIEND=-163,163 GT:GL:GQ:FT:RCL:RC:RCR:RDCN:DR:DV:RR:RV 0/1:-47.1887,0,-0.689909:8:LowQual:1933048:1816911:901744
:1:1:10:0:0 0/1:-29.1784,0,-83.6784:10000:PASS:1384338:1282553:621712:1:15:6:0:0 0/1:-42.776,0,-72.8757:10000:PASS:1939476:1999794
:1008088:1:14:11:0:0 0/1:-43.2887,0,-2.68995:27:PASS:1655597:1489849:724173:1:1:10:0:0
```

Maybe...
But it would be lost
among the false
positives!



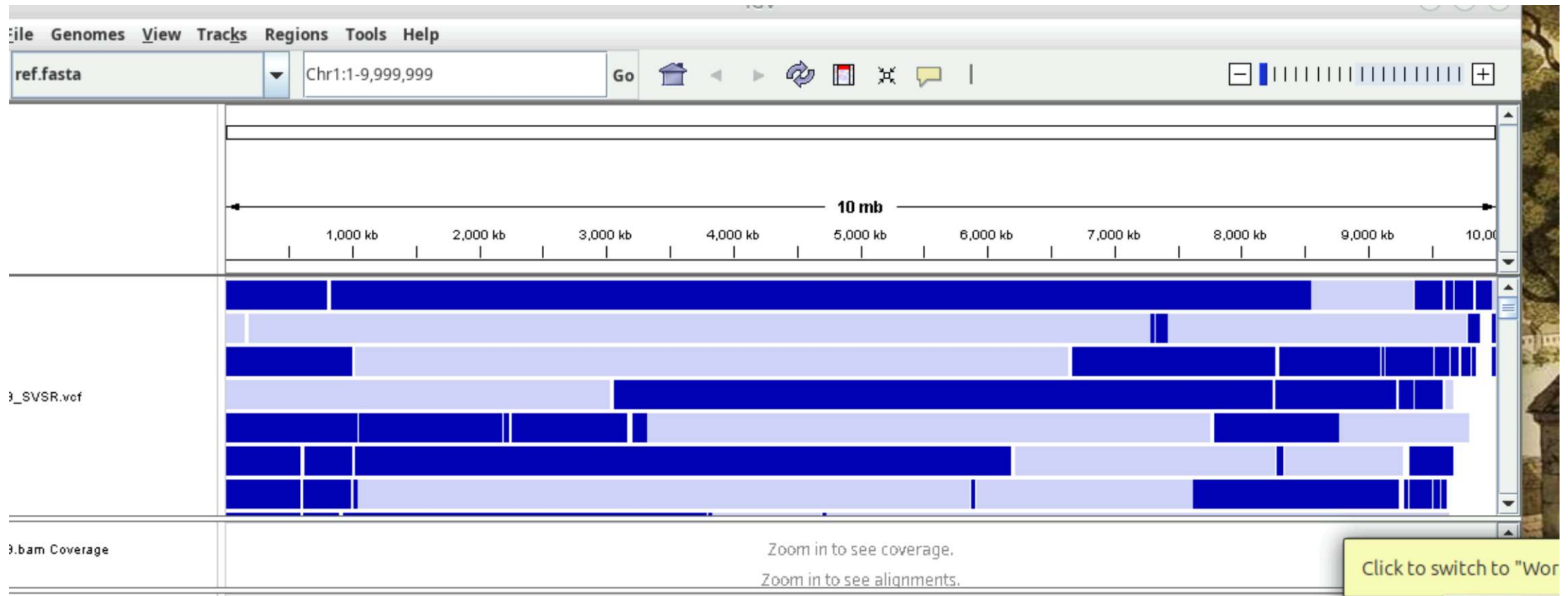
4 – SV call with short-reads

-> What do you think of the SVs? Is it readable?

Too many long SV for IGV

Lots of SVs at the same position

=> Probably a sign of lots of false positives!



4 – SV call with short-reads

-> Can you find some deletions well supported by reads?



4 – SV call with short-reads

-> How many SVs are you analysing in total?

16330

-> What do you think of allelic frequency? And missing data?

Little missing data

BUT : lots of allelic frequency at 0 (and genotypes are 0/0)
= ref/ref

⇒ Those are not even variants

⇒ Remove them

Chr1	5	INV00000000	9396125	0.75	0
Chr1	519	DEL00000001	552	0	0.125
Chr1	958	INS00000002	958	0	0
Chr1	5355	DUP00000003	826732	0	0.5
Chr1	5356	DUP00000004	180476	0	0
Chr1	5356	DUP00000005	616735	0	0.5
Chr1	5373	INV00000006	616199	0	0.5
Chr1	5375	INV00000007	894221	0	0
Chr1	5378	INV00000008	1021738	0	0.5
Chr1	5379	INV00000009	1018396	0	0
Chr1	5382	INV00000010	871395	0	0.5
Chr1	5383	DUP00000011	5547	0	0
Chr1	5383	DUP00000012	604520	0	0
Chr1	5383	DUP00000013	617527	0	0.5
Chr1	5383	INV00000014	816676	0	0
Chr1	5383	INV00000015	1024400	0	0.25
Chr1	5383	INV00000016	1068736	0	0
Chr1	5383	INV00000017	1143801	0	0
Chr1	5383	DUP00000018	1144327	0	0
Chr1	5383	INV00000019	3052463	0	0.25
Chr1	5383	INV00000020	3811013	0	0
Chr1	5383	INV00000021	8676150	0	0
Chr1	5384	DUP00000022	204296	0	0.25
Chr1	5384	INV00000023	4283625	0	0.5
Chr1	5386	INV00000024	968242	0	0
Chr1	5404	DUP00000025	1075475	0	0.125
Chr1	5405	INV00000026	1076829	0	0.125
Chr1	5425	DUP00000027	1021486	0	0.5
Chr1	5486	INV00000028	1096631	0	0
Chr1	5494	INV00000029	1010012	0	0
Chr1	5497	DEL00000030	5278163	0	0.25
Chr1	5503	INV00000031	744468	0	0.125
Chr1	5540	DEL00000032	959246	0	0
Chr1	5546	INV00000033	9567667	0	0.125
Chr1	5555	DEL00000034	8515626	0	0.125
Chr1	5557	DEL00000035	871141	0	0.5
Chr1	5621	DUP00000036	1046565	0	0
Chr1	5622	INV00000037	1046561	0	0.375
Chr1	5679	DUP00000038	6664587	0	0
Chr1	5728	INV00000039	958032	0	0.125
Chr1	5735	INV00000040	888458	0	0.5
Chr1	5757	INV00000041	1021865	0	0.5
Chr1	5776	DEL00000042	814677	0	0

4 – SV call with short-reads

- > How many SVs are you analysing in total? Same? **16330 - same**
- > Do you notice differences with the earlier vcf?

Chr	Start	ID.x	End	F_missing_before	MAF_before	ID.y	F_missing_after	MAF_after
1	Chr1	5	INV000000000 9396125	0.75	0	INV000000000	0.75	0
2	Chr1	113015	DUP00000129 2980316	0.50	0	DUP00000129	0.50	0
3	Chr1	143486	INS00000191 143486	0.75	0	INS00000191	0.75	0
4	Chr1	1060949	INV00010779 4104687	0.25	0.5	INV00010699	0.25	0.5
5	Chr1	1329380	DUP00011490 6847771	0.25	0.5	DUP00011392	0.25	0.5
6	Chr1	1419614	INS00011608 1419614	0.50	0	INS00011507	0.50	0
7	Chr1	1483914	INS00011643 1483914	0.25	0	INS00011541	0.25	0
8	Chr1	1488455	DEL00011646 1488499	0.25	0.5	DEL00011544	0.25	0.5
9	Chr1	1531849	DEL00011677 1532372	0.75	0	DEL00011574	0.75	0
10	Chr1	1537050	DEL00011680 1537103	0.25	0.166667	DEL00011577	0.25	0.166667
11	Chr1	1554194	DEL00011691 1554232	0.25	0.333333	DEL00011587	0.25	0.333333
12	Chr1	1567946	INS00011711 1567946	0.25	0	INS00011606	0.25	0
13	Chr1	1569316	DEL00011712 1569358	0.50	0	DEL00011607	0.50	0
14	Chr1	1584848	DEL00011729 1584932	0.25	0.166667	DEL00011624	0.25	0.166667
15	Chr1	1592686	DEL00011737 1706036	0.25	0.333333	DEL00011632	0.25	0.333333
16	Chr1	1607047	INV00011746 3686296	0.25	0.333333	INV00011640	0.25	0.333333
17	Chr1	1672225	INS00011781 1672225	0.25	0.333333	INS00011674	0.25	0.333333
18	Chr1	1691248	INV00011788 5153424	0.25	0.166667	INV00011681	0.25	0.166667
19	Chr1	1693676	INS00011793 1693676	0.25	0.166667	INS00011686	0.25	0.166667
20	Chr1	1733296	INV00011829 9367617	0.25	0.333333	INV00011720	0.25	0.333333
21	Chr1	1754730	DEL00011846 1754772	0.25	0.166667	DEL00011737	0.25	0.166667
22	Chr1	1796733	DEL00011874 1796793	0.25	0.333333	DEL00011765	0.25	0.333333
23	Chr1	1894880	DEL00011934 8803568	0.50	0.25	DEL00011825	0.50	0.25
24	Chr1	1899396	INS00011939 1899396	0.25	0.333333	INS00011830	0.25	0.333333

Not better for missing data

& re-genotyping has
renamed the SV... (hard to
compare!)

But breakpoints should
have been refined (yay!)

5- Towards genome graphs

We get some warnings. Why?

The graph needs the
sequence in the ALT field...

```
warning:[vg::Constructor] vcflib could not canonicalize some SVs to base-level sequence; skipping variants like: Chr1 129696 Sniffles2.DUP.240S0 A <DUP> 59
PASS PRECISE;SVTYPE=DUP;SVLEN=1390;END=131086;SUPPORT=10;COVERAGE=31,23,662,25,32;STRAND=+-;STDEV_LEN=4.593;STDEV_POS=0.408;VAF=0.312
Warning: deletion END and SVLEN do not agree [canonicalize] Chr1 519626 Sniffles2.DEL.1F9S0 A <DEL> 51 PASS PRECISE;SVTYPE=DEL;SVLEN=-
129928;END=649553;SUPPORT=11;COVERAGE=35,37,19,22,21;STRAND=+-;STDEV_LEN=0.000;STDEV_POS=0.000;VAF=0.423;SPAN=129928
END: 649553 SVLEN: 129928
```

-> open the vcf: how many variants could you genotype? What does the format look like?

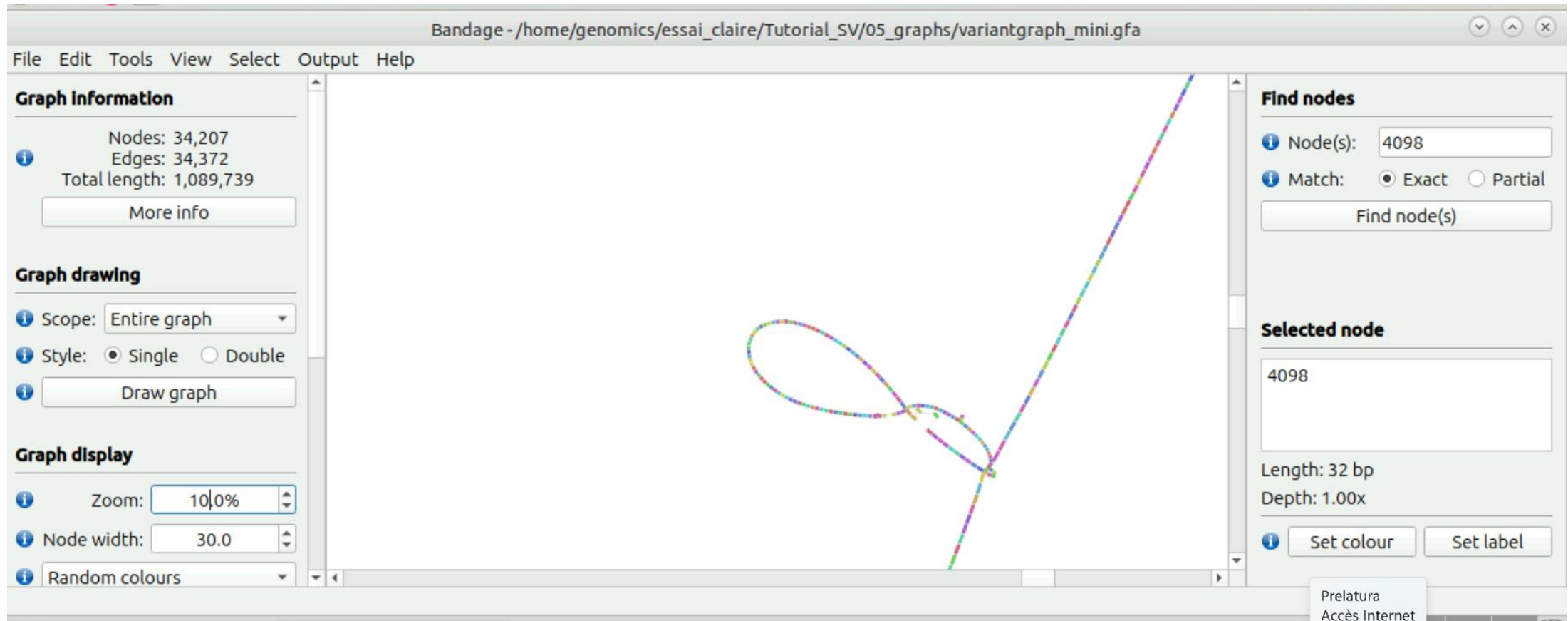
145 out of 170

5- Towards genome graphs

[illegible]

5- Towards genome graphs

Visualisation in Bandage – exemple of a bubble for a large SV



5- Towards (pan)genome graphs

-> What do you observe on the 2D representation?

Our big
inversion!

