



# Experimental Design

**Joana Meier**

**Tree of Life Programme, Wellcome Sanger Institute**

THE  
ROYAL  
SOCIETY

The  
Branco Weiss  
Fellowship  
Society in Science

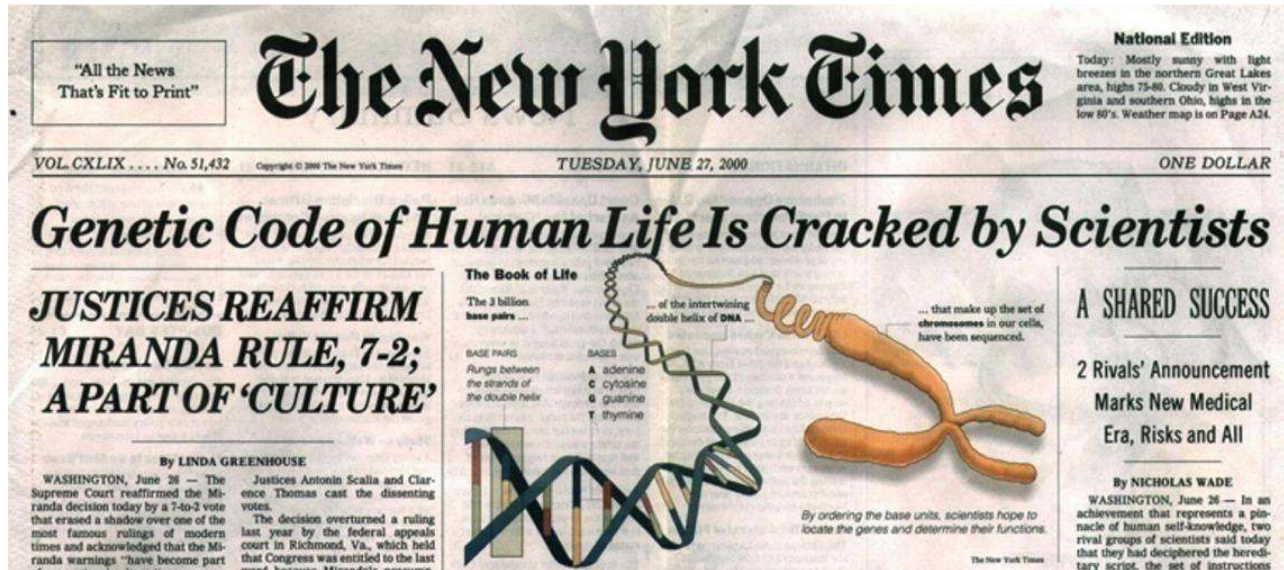


UNIVERSITY OF  
CAMBRIDGE

# Outline

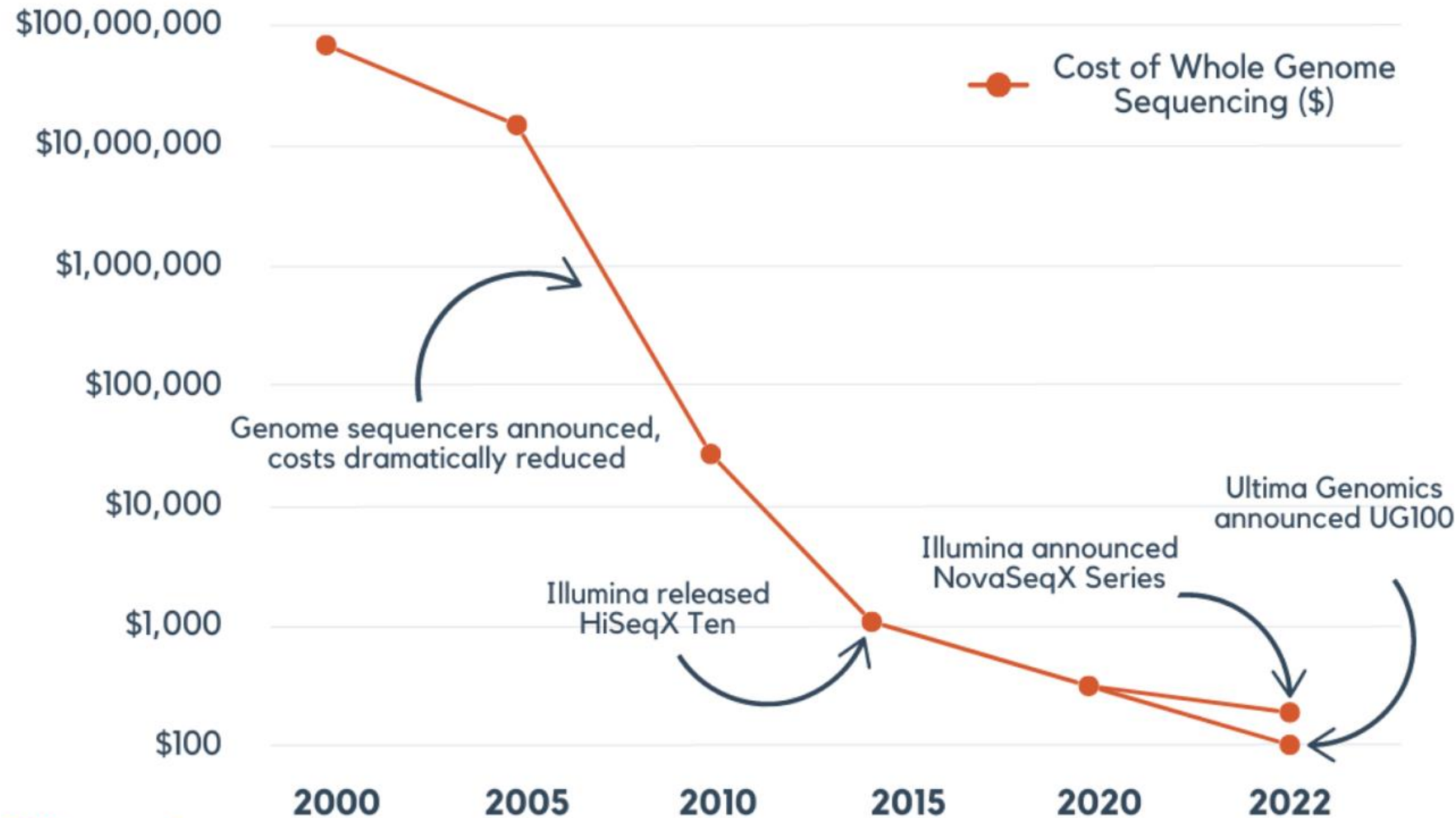
- Sequencing technologies
- Sampling and sequencing strategy
- Case studies
- Ensuring you trust your data
- Breakout groups to discuss your experimental design
- Reference genomes

# Human Genome Project



- Human genome project – started in 1990, completed in 2003
- Sequenced across ~20 institutions worldwide
- Cost an approximate \$5 billion US dollars

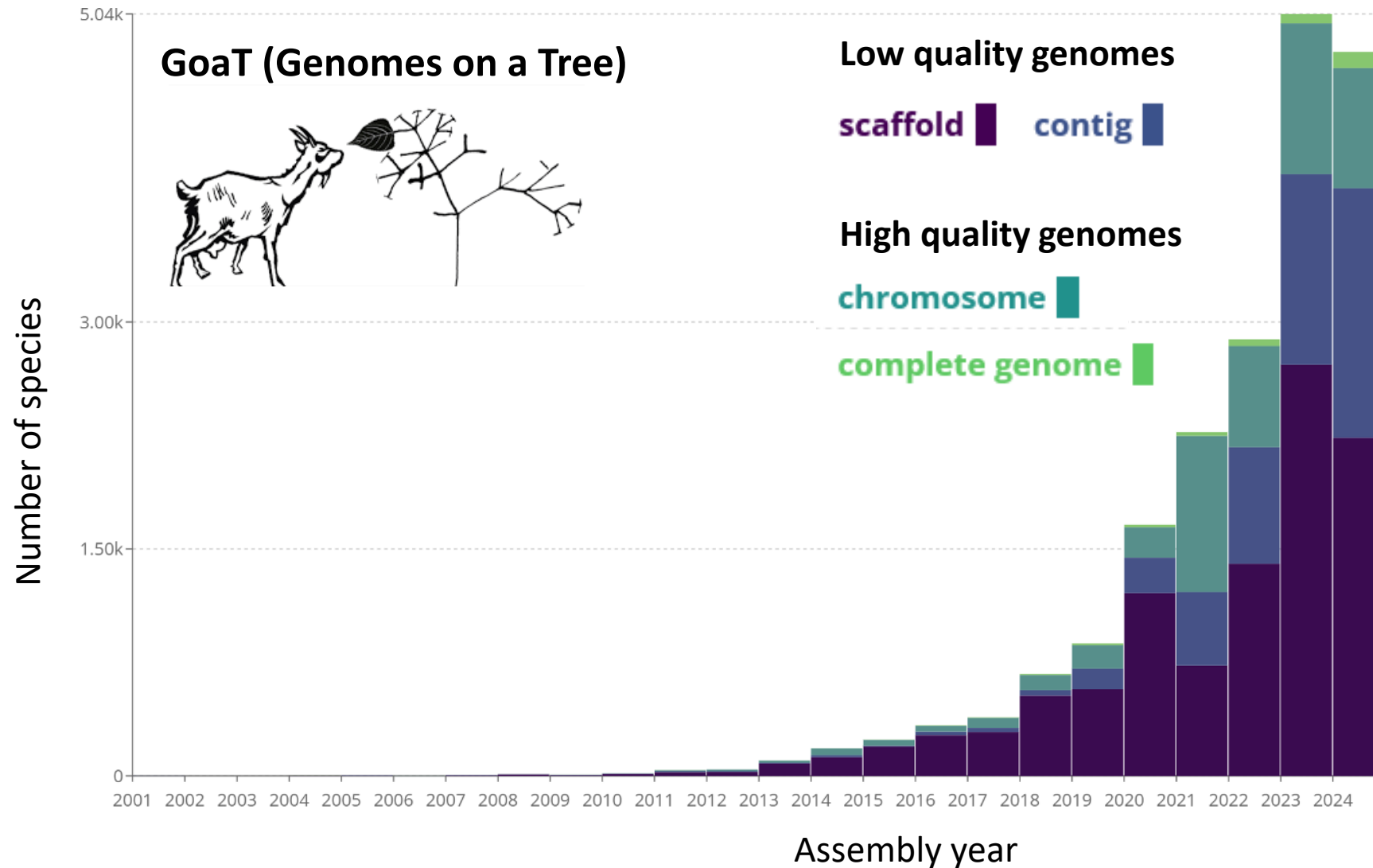
# Sequencing costs are decreasing rapidly



Since Oct 2023  
PacBio Revio  
(66 Gbp per lane  
in 15 kb reads)



# More and more species have reference genomes



# Two main types of high-throughput sequencing

- **Short-read sequencing**

- Reads are typically 150-300 bp long
- Cheaper than long-read sequencing
- E.g. Illumina, Ultima Genomics

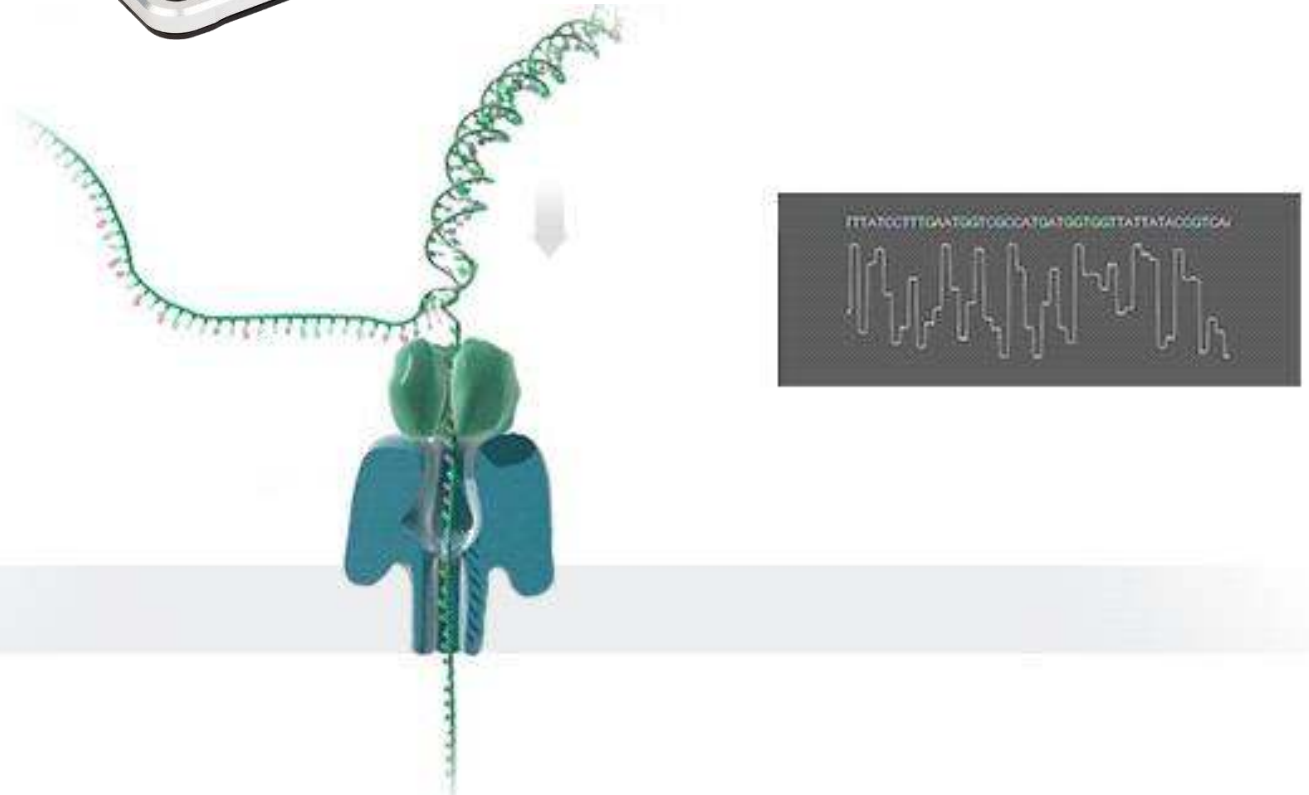
- **Long-read sequencing**

- Reads are typically >10 kb long (PacBio: 10-20 kbp, ONT/Nanopore: >100 kbp)
- More expensive than short-read technologies
- Required for making a reference genome
- E.g. PacBio or ONT/Nanopore

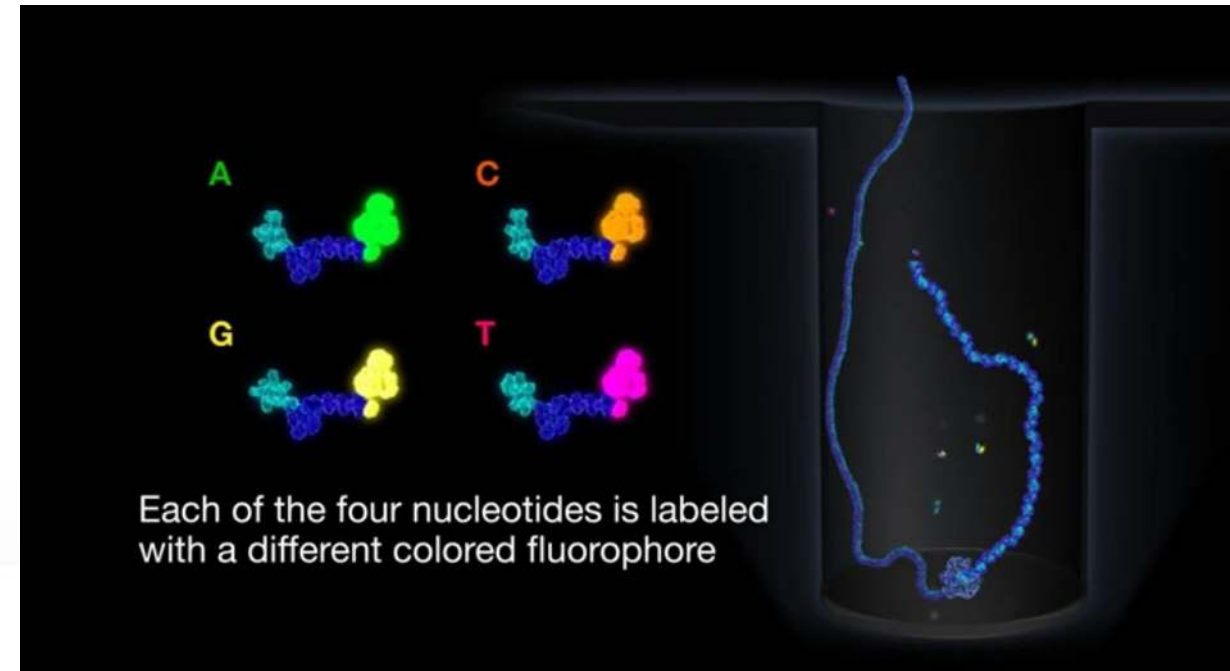
# Long read sequencing technologies



**ONT / Nanopore**  
**(>100 kb reads)**



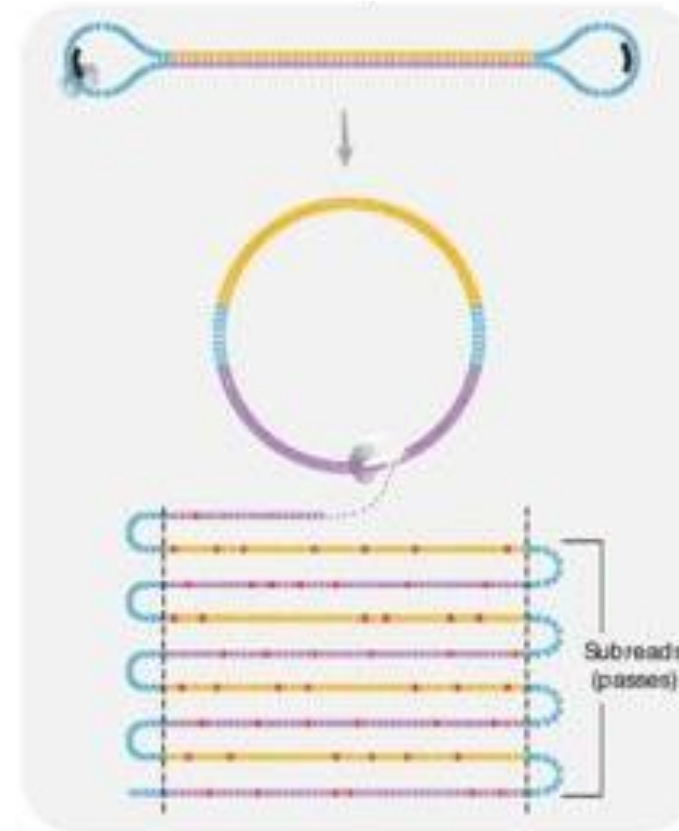
**PacBio**  
**~10-20 kb reads**



# PacBio HiFi reads (99.95% accuracy)

Each DNA-fragment is sequenced many times to get a high-quality consensus (=summary) read

Multi-pass sequencing  
on Sequel II System



HiFi Read Base Calling

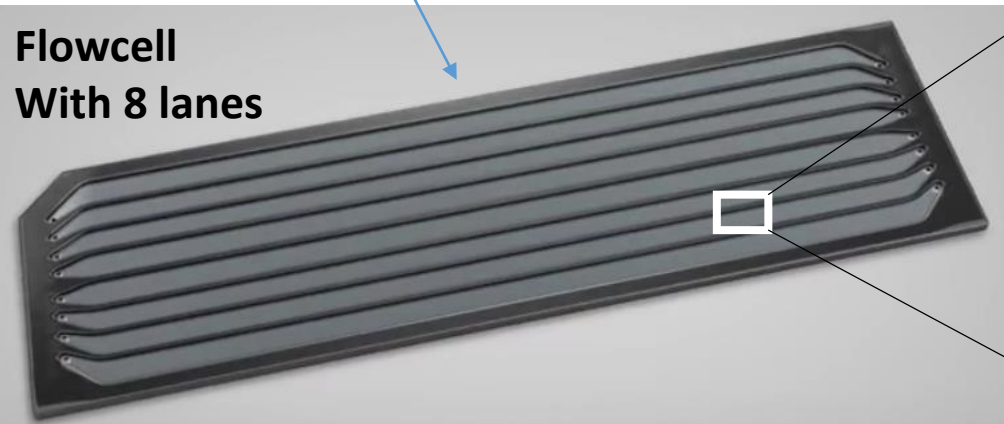
HiFi READ  
Read Lengths up to 25,000 bp  
Average Read Accuracy  $\geq 99.5\%$



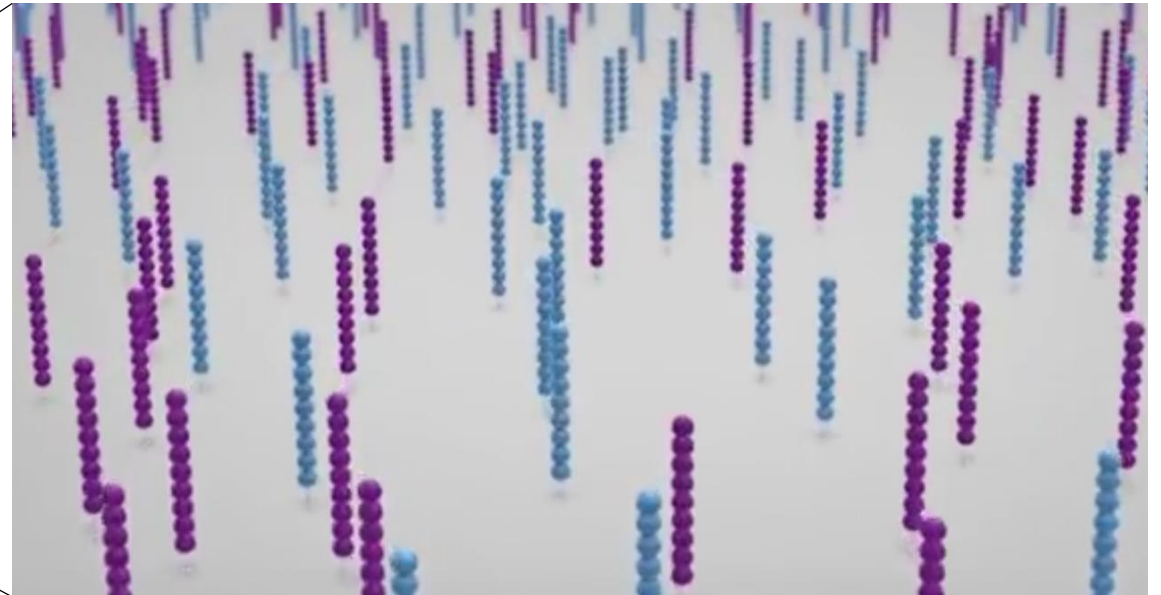
# Illumina flowcell

DNA fragments with Illumina adapters

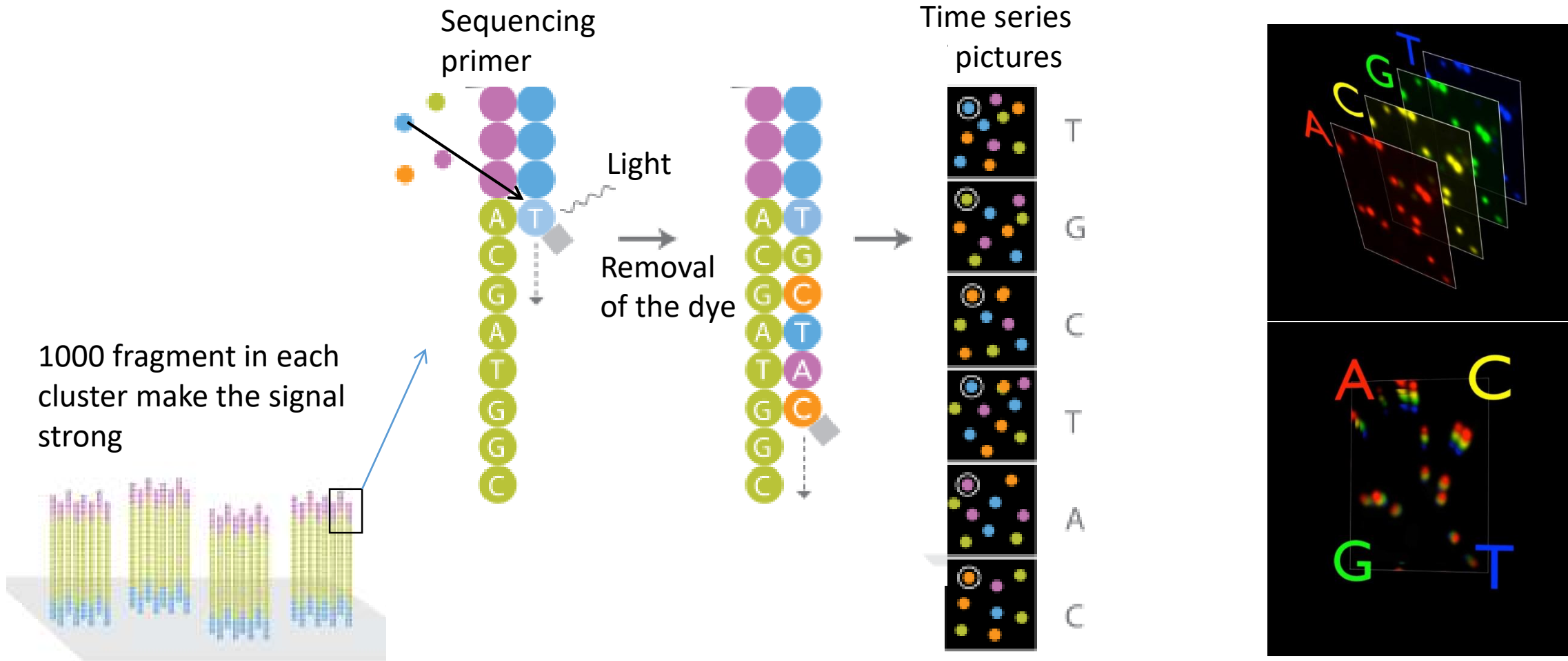
Flowcell  
With 8 lanes



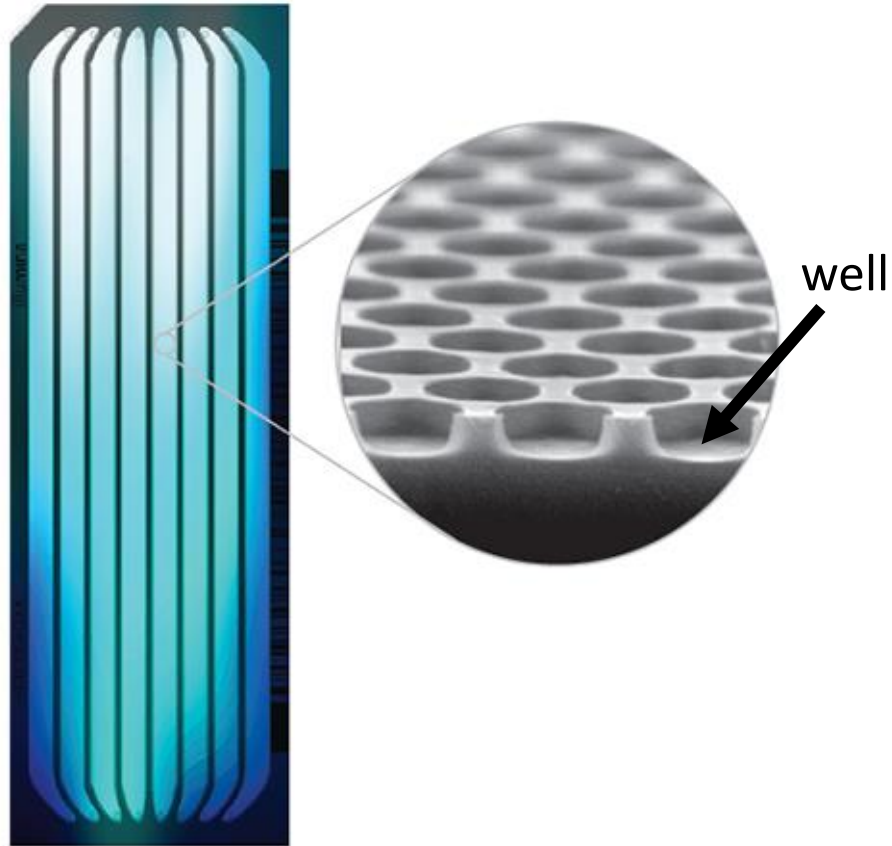
Each lane contains a dense lawn of Illumina primers



# Short-read sequencing with Illumina



**Newer Illumina machines use wells and only 2 colours**  
(e.g. Novaseq, Nextseq, MiniSeq. This makes it faster and cheaper)

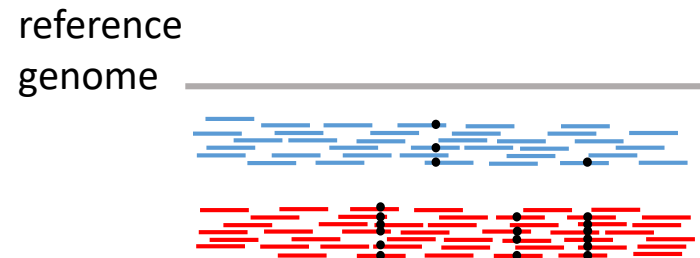


| 2-Channel Chemistry |                                                                                          |   |                                                                                          |                                                                                          |
|---------------------|------------------------------------------------------------------------------------------|---|------------------------------------------------------------------------------------------|------------------------------------------------------------------------------------------|
|                     | <br>A | G | <br>T | <br>C |
| Image 1             |       |   |       |                                                                                          |
| Image 2             |     |   |                                                                                          |     |
| Result              | A                                                                                        | G | T                                                                                        | C                                                                                        |

# Sequencing approaches for speciation genomics

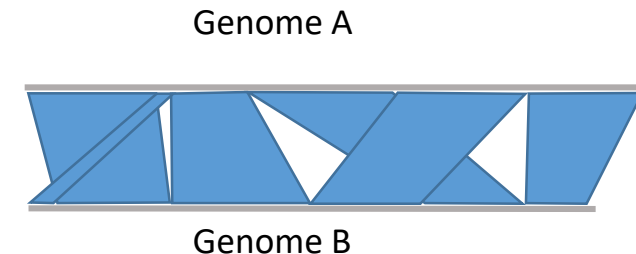
## Whole-genome resequencing (short-read data)

- Requires a reference genome
- individuals need to be from the same or closely related species
- Complete genome sequenced



## Genome assembly comparisons (long-read data)

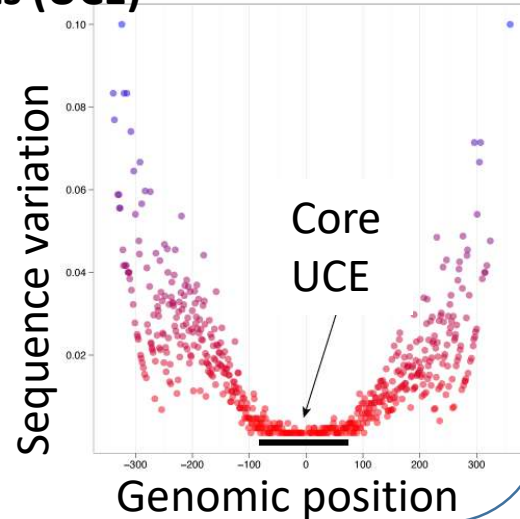
- Comparative genomics – studying structural variation between species, can be distantly related
- Gene expansions, transposable elements etc
- Phylogenomics across deeply divergent species
- Pangenomics – multi-genome assemblies to study within-species variation in structural variants



# Reduced-representation techniques (only parts of the genome sequenced)

## Ultra-conserved elements (UCE)

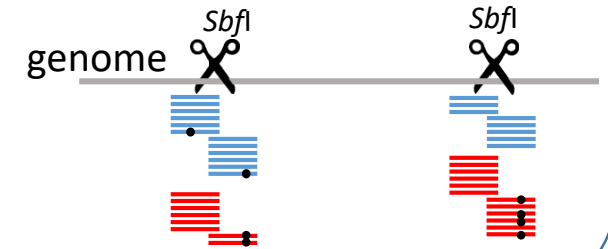
- Sequence capture with baits based on genomic regions that are conserved across many species
- Works with highly divergent species



## Restriction Associated Sequencing (RAD)

(similar methods: GBS, ddRAD)

- does not require primers/baits or reference genome
- individuals need to be from the same or closely related species
- Information from thousands of loci distributed across the genome

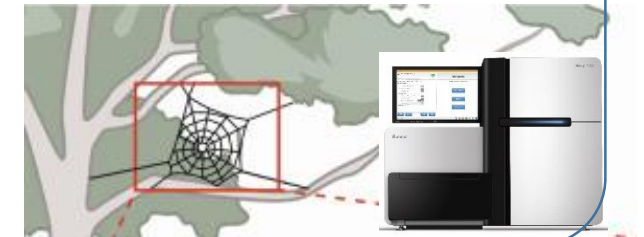


## Targeted or amplicon sequencing, e.g. barcoding

- Sequencing one or few genes
- requires primers
- e.g. CO1 (mitochondrial barcoding region), advantage: large database (BOLD) available to compare to for species identification

## Environmental DNA (eDNA)

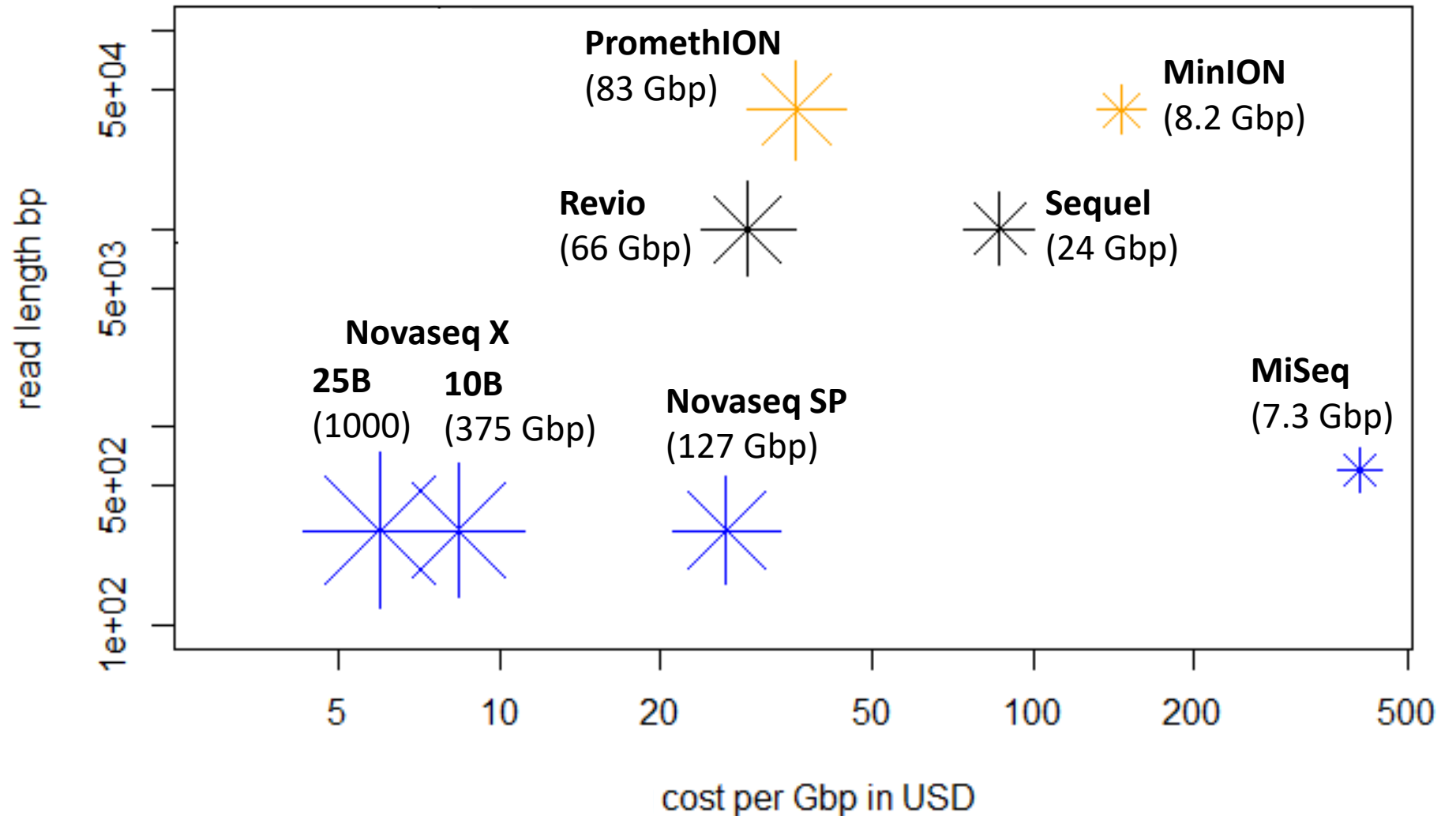
- Mostly CO1 sequencing from soil, water, air (spider webs)
- Studying local species richness



# Read length versus per Gbp sequencing costs for different sequencing machines (note the axes are in logarithmic scale)

\* ONT  
\* PacBio  
\* Illumina

Star size shows the total throughput per lane, also given in parentheses ( )



# Sequencing strategy

## Total sequencing data gets divided by:

- Number of sites to sequence
  - Depends on genome size and sequencing strategy, e.g. RAD versus whole-genome
- Number of specimens to sequence
  - and how many specimens per population, how many populations per species
- Sequencing depth (e.g. sequencing at 10x depth of coverage)
- Example: 1 NovaseqX 10B lane
  - ~2.5 billion paired-end reads of 150 bp each -> 375 Gbp data
    - 100 whole-genomes of a species with 0.375 Gbp genome size at 10x coverage
    - 19 whole-genomes of a species with 1 Gbp genome size at 20x coverage
    - 375 individuals sequenced with a RAD sequencing approach resulting in 50 Mbp at a sequencing depth of 20x

# Different questions require different experimental design

## Resolving the taxonomy

- Placing potentially new species
- Species delineation



## Adaptation to climate change

- Identifying genomic regions involved in adaptation

Pfenninger  
et al. 2021



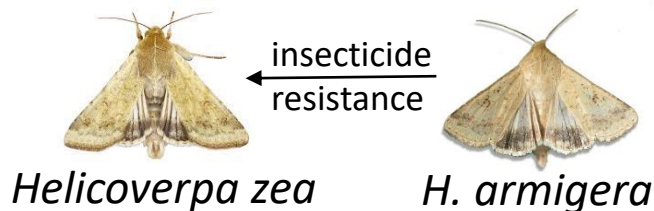
## Are the species declining?

- Detecting past inbreeding
- Assessing genetic diversity



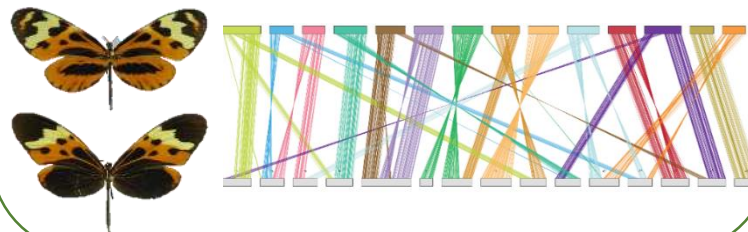
## Detecting introgression

- Are populations/species hybridising now or in the past?
- Finding regions of adaptive introgression



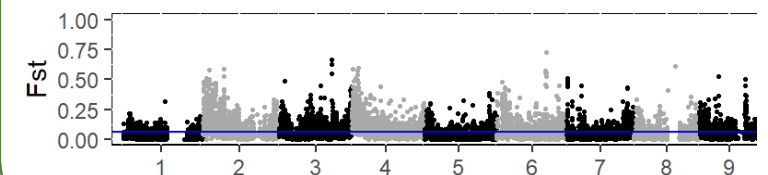
## Studying genome evolution

- Gene expansions, e.g. olfactory
- Chromosomal rearrangements
- Genome size evolution (TEs, etc)



## What genes are under divergent selection

- Short-term (recent sweep)
- Long-term (fixed substitutions)



# Sampling considerations

- Knowing the natural history and morphologically inferred taxonomy if your helps massively
- If you want to test for introgression, you need outgroups
- Aim to get even sampling of populations/species to compare
- Sampling sites matter:  
e.g. if you want to test for ongoing gene flow, you need specimens from sympatry or parapatry
- High number of specimens needed for:
  - Inferring recent demographic changes
  - Studying rare alleles
  - GWAS (inferring the genetic basis of traits, particularly if polygenic)

# Case studies to discuss

Three examples with very different optimal sampling designs

- **Identifying barriers to gene flow and the genetic basis of relevant traits in a hybrid zone**

# *H. erato* and *H. melpomene* in Ecuador



## *Heliconius erato*



Highland



Lowland

## *Heliconius melpomene*



Highland



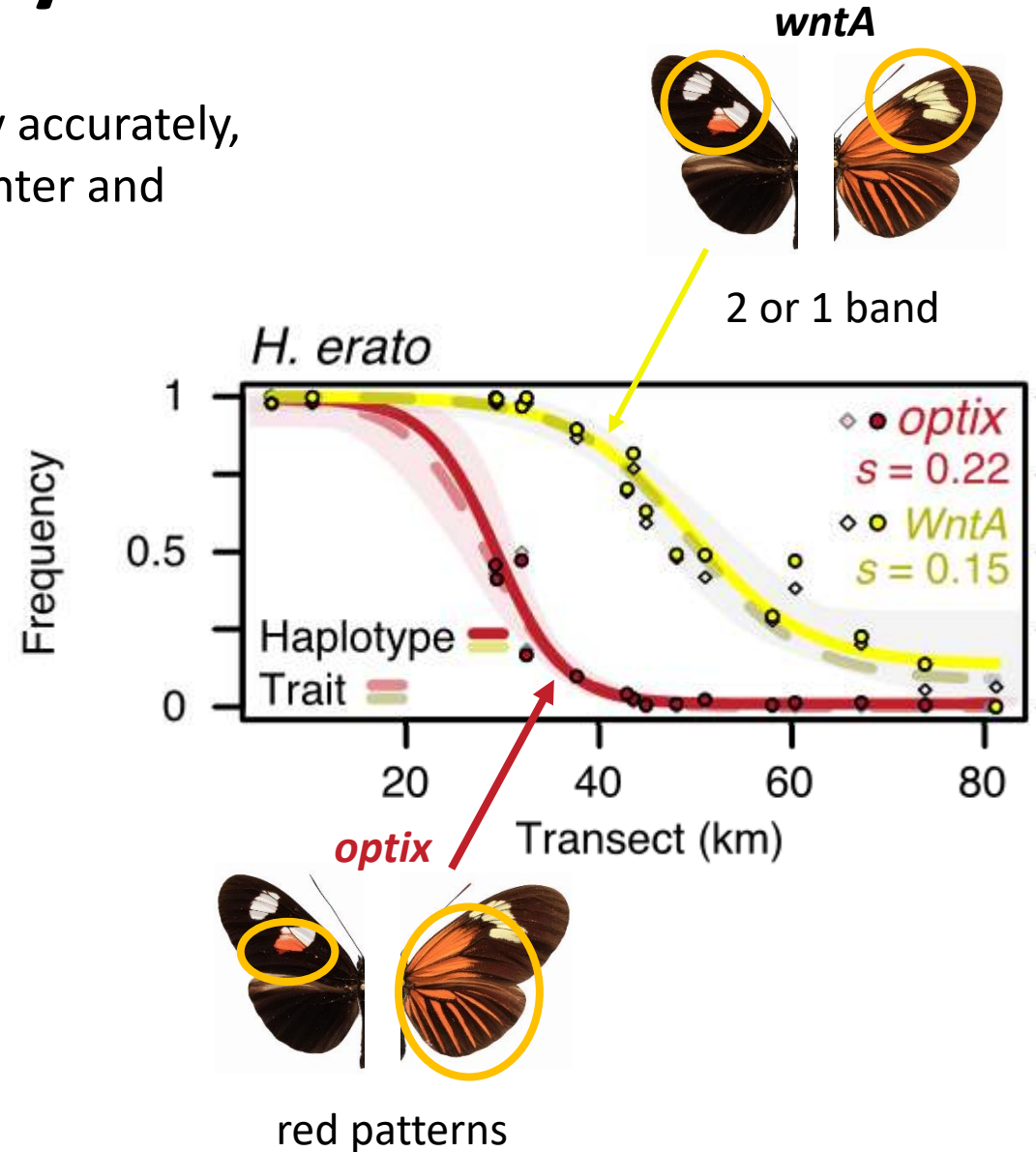
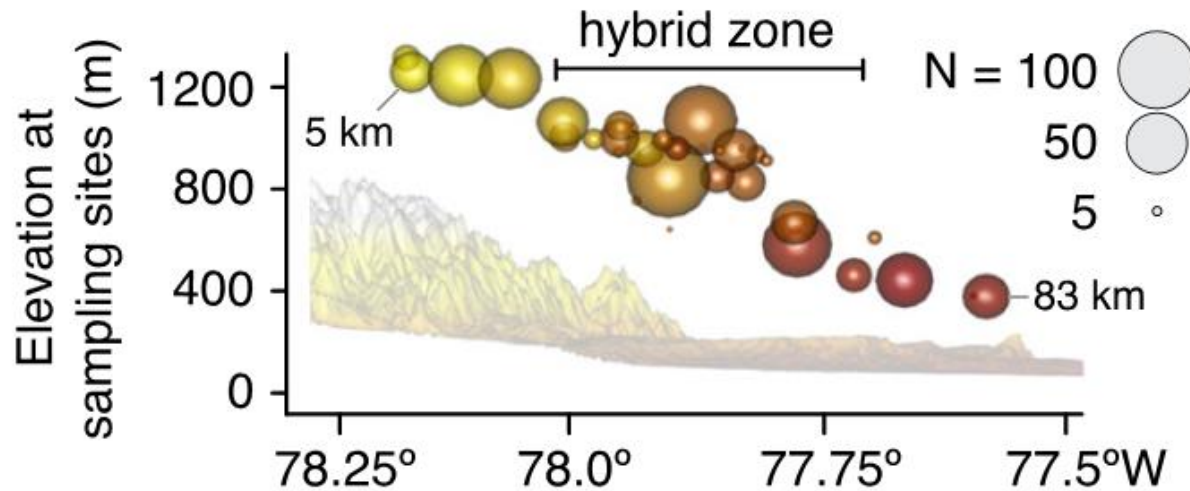
Lowland

Patricio Salazar Chris Jiggins

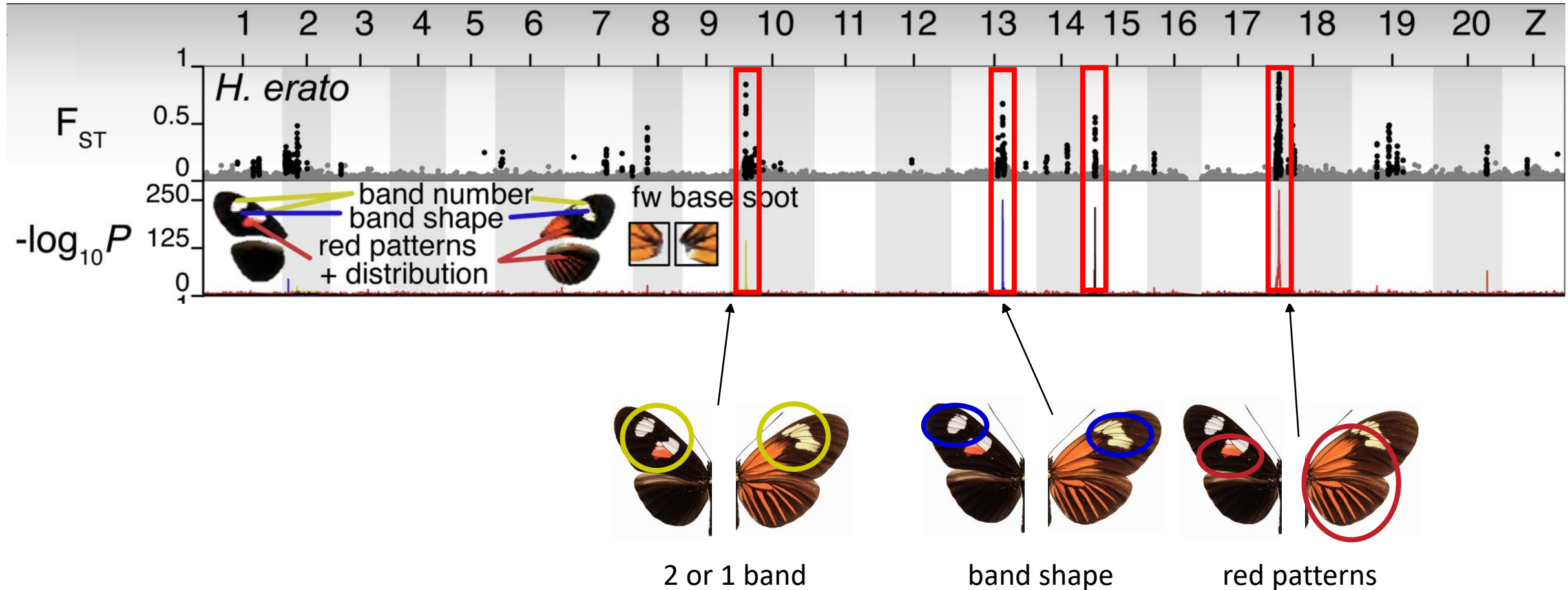


# *Heliconius erato* hybrid zone

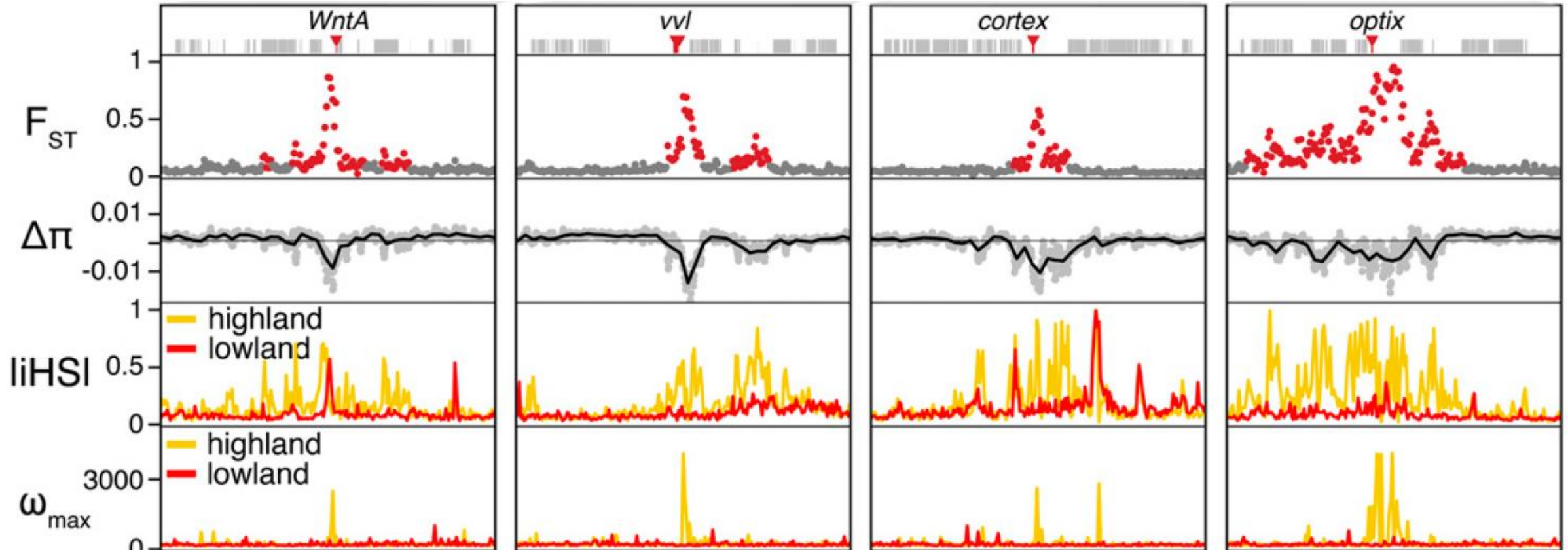
Many individuals per population to estimate allele frequency accurately,  
many populations along the hybrid zone to infer the cline center and  
shape accurately



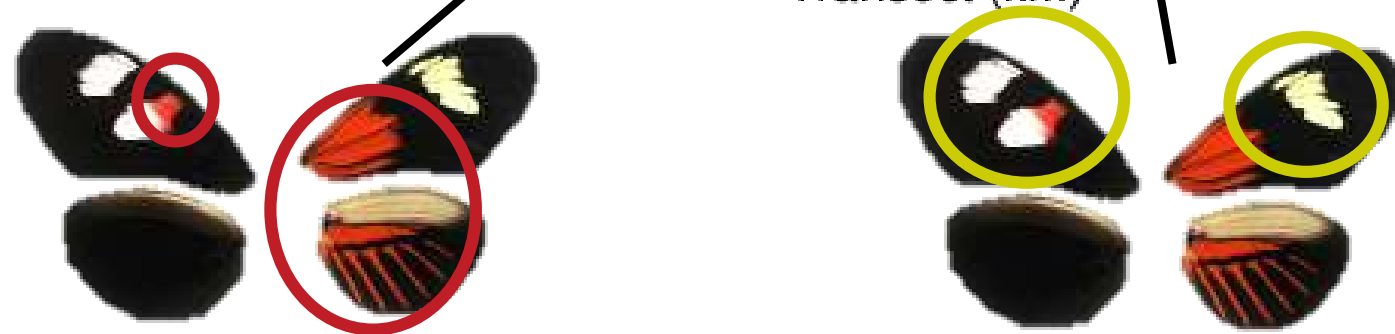
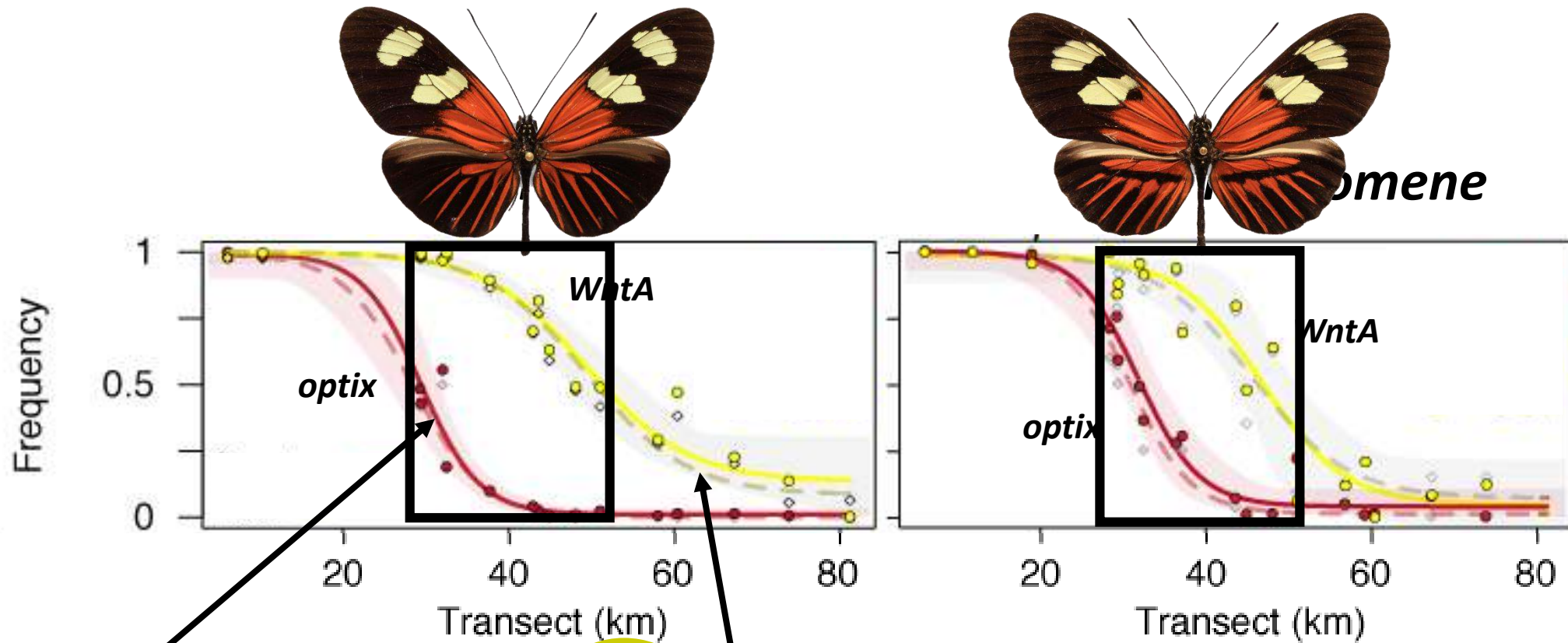
# $F_{ST}$ peaks coincide with GWAS peaks and show signatures of selective sweeps



# $F_{ST}$ peaks coincide with GWAS peaks and show signatures of selective sweeps



In both *Heliconius erato* and *H. melpomene*, the clines at the major colour loci are shifted by 18 km

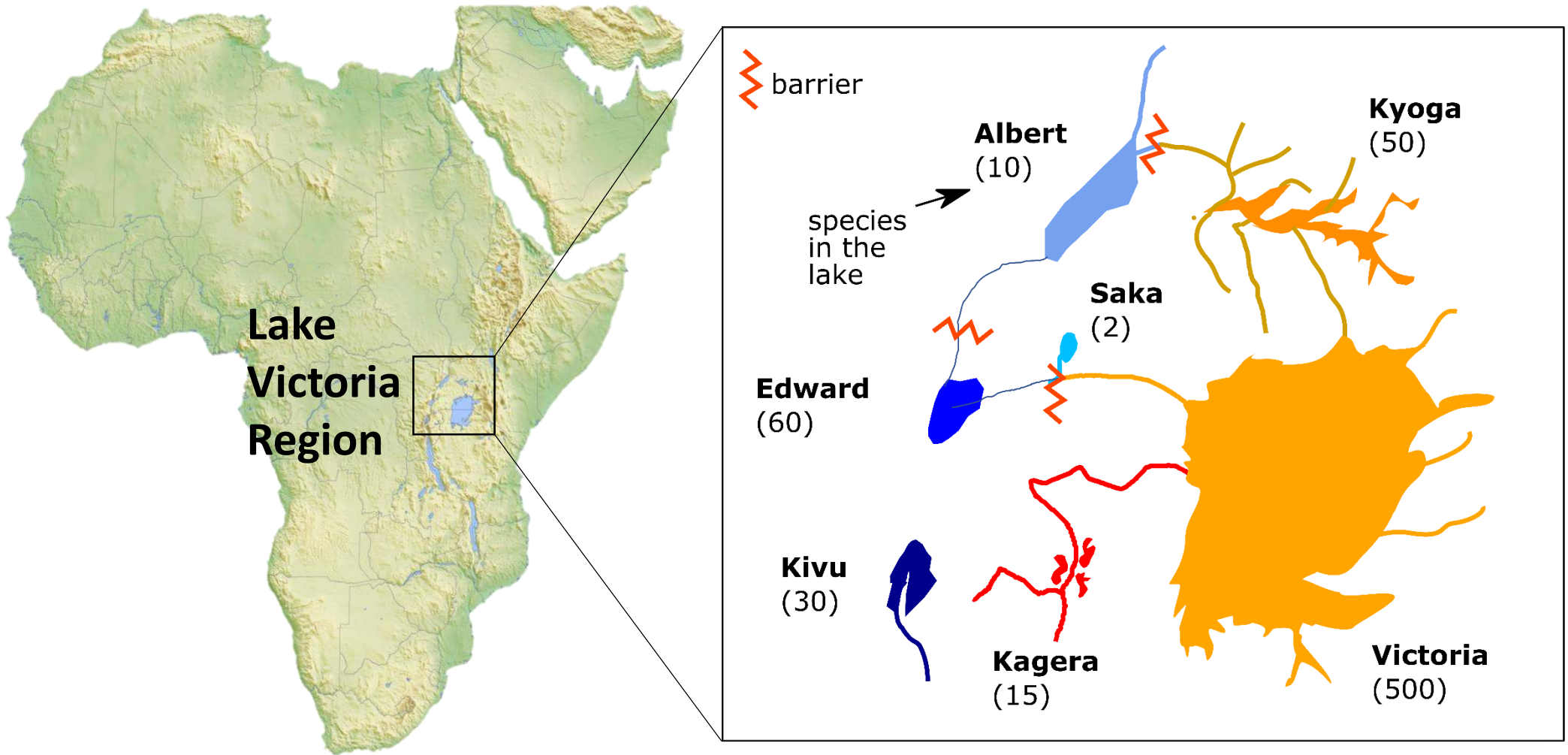


# Case studies to discuss

Three examples with very different optimal sampling designs

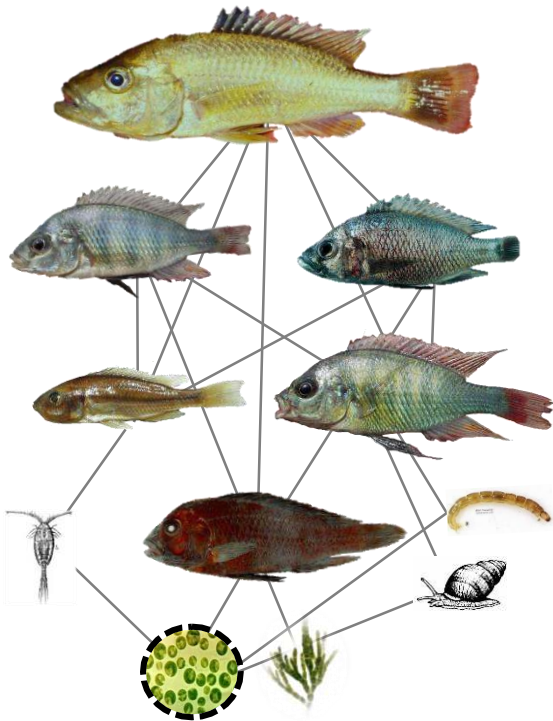
- Identifying barriers to gene flow and the genetic basis of relevant traits in a hybrid zone
- **Inferring if a fish species community evolved in a lake or if the different species independently colonised the lake.**

# The same lineage of cichlid fishes diversified in all major lakes in the Lake Victoria Region in only 150,000 years

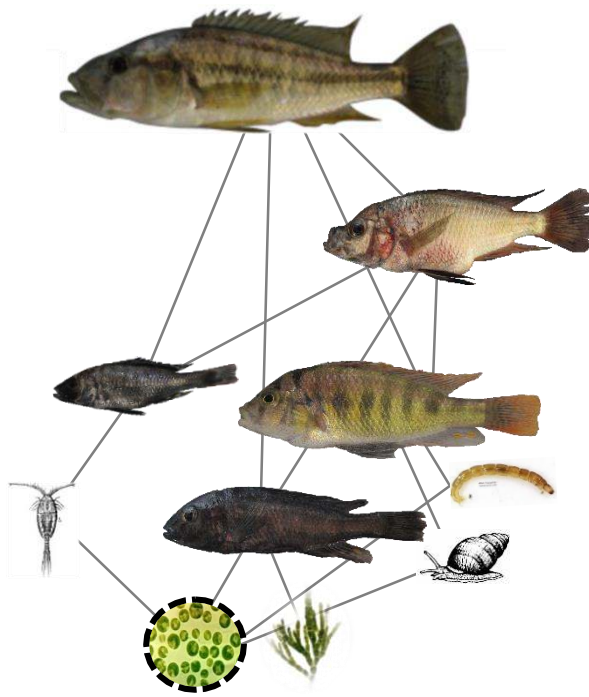


**In each lake, cichlids occupy a wide range of ecological niches.  
But Lake Victoria is much younger! Only 16,000 years!**

**Lake Edward**

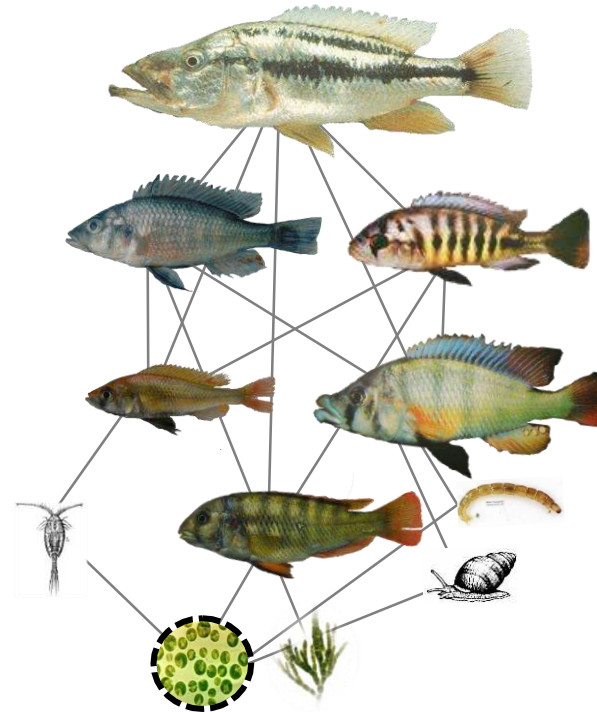


**Lake Kivu**

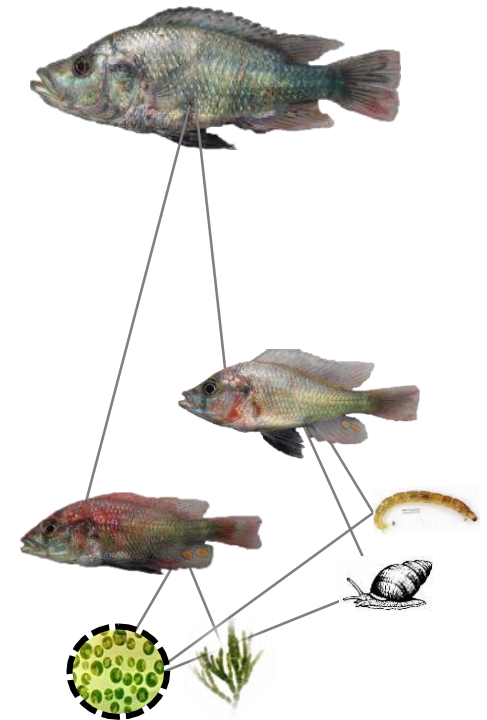


**Less than 16,000 years old!**

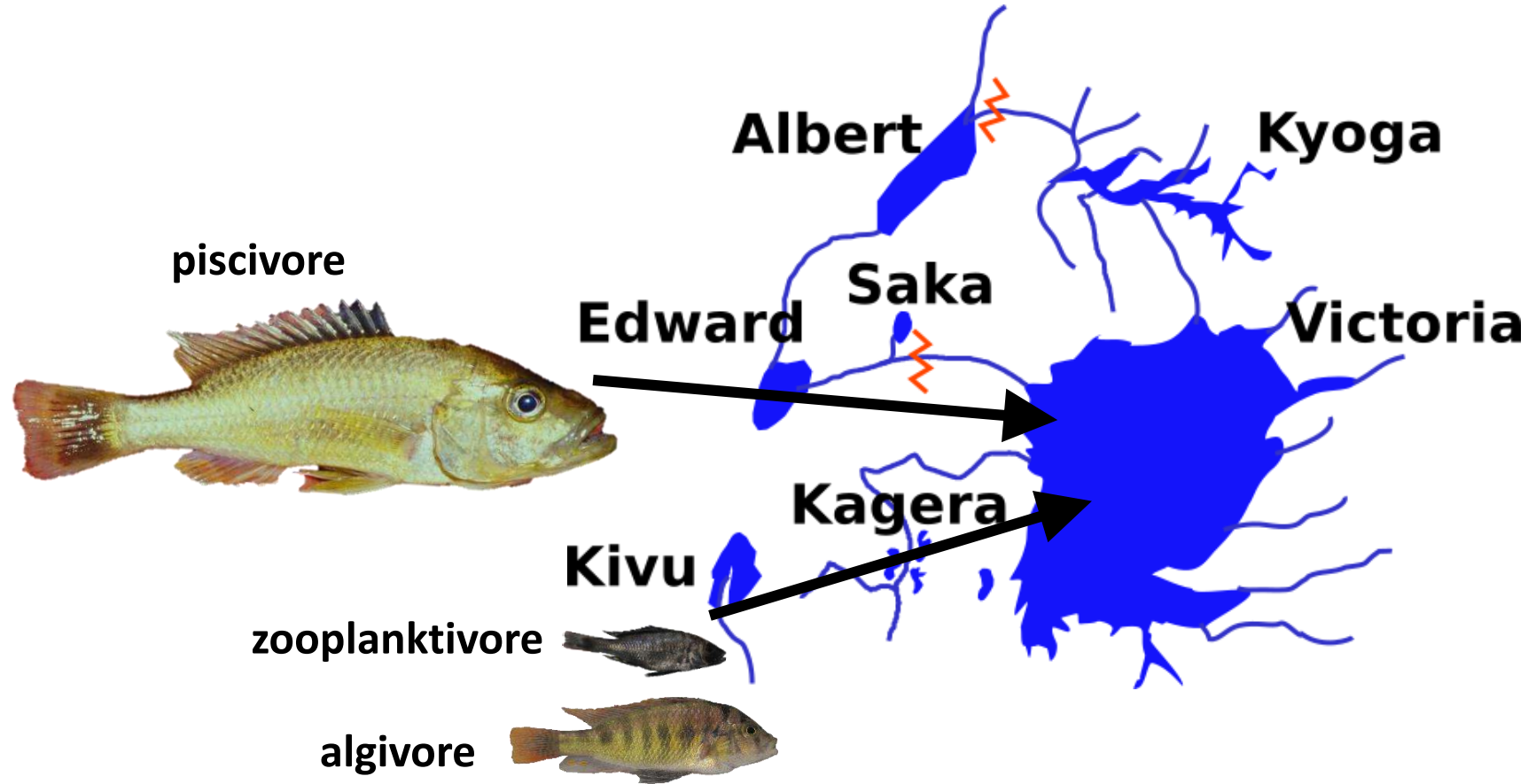
**Lake Victoria  
(500 species)**



**Kagera Lakes**

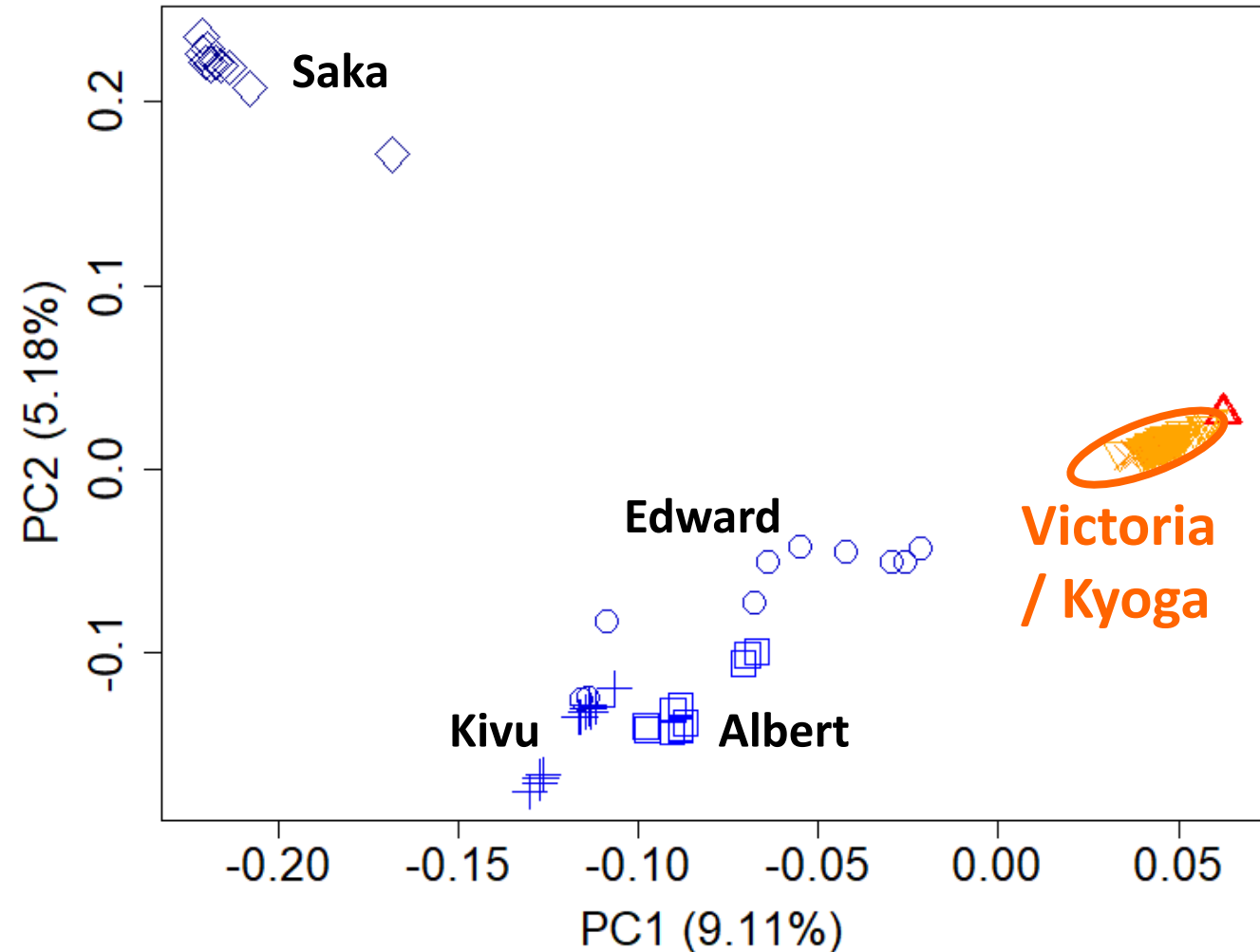
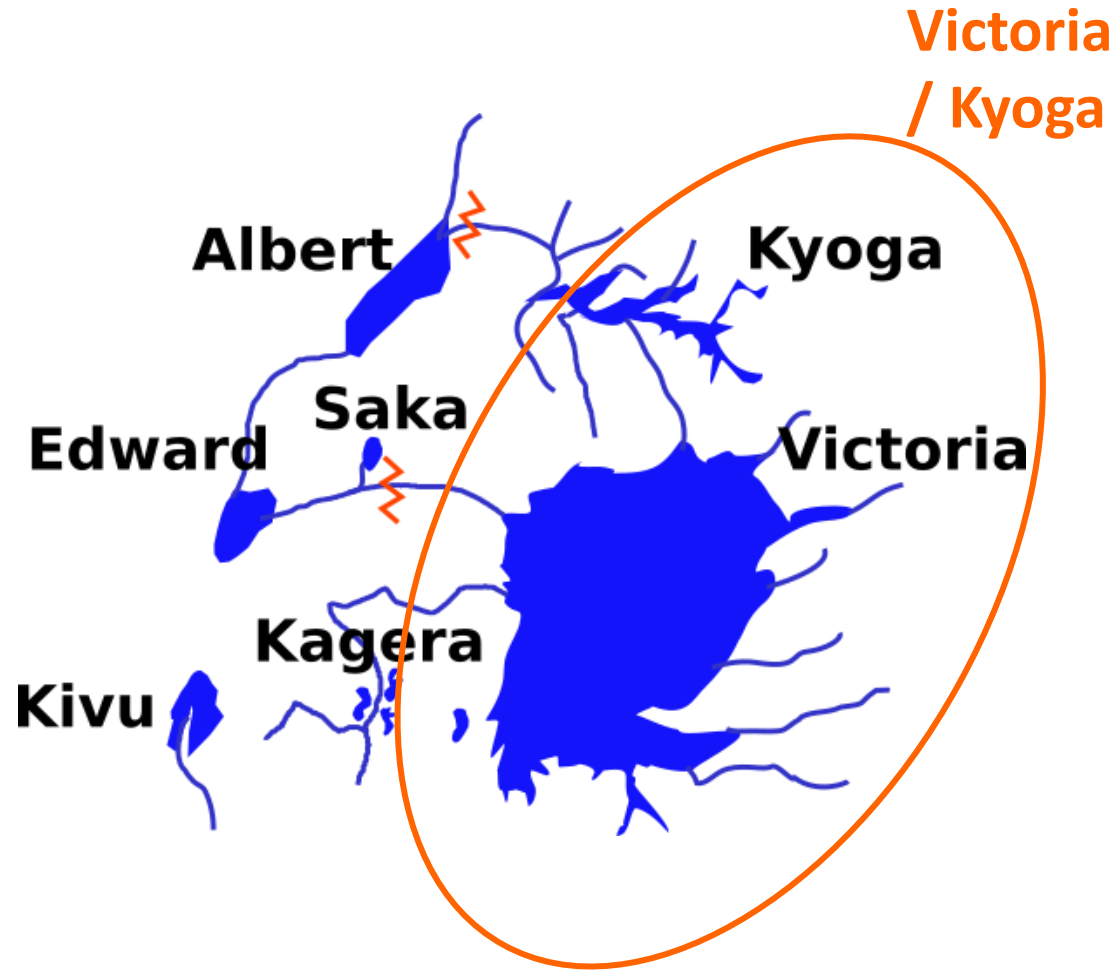


# Did the different ecological groups colonise Lake Victoria independently?



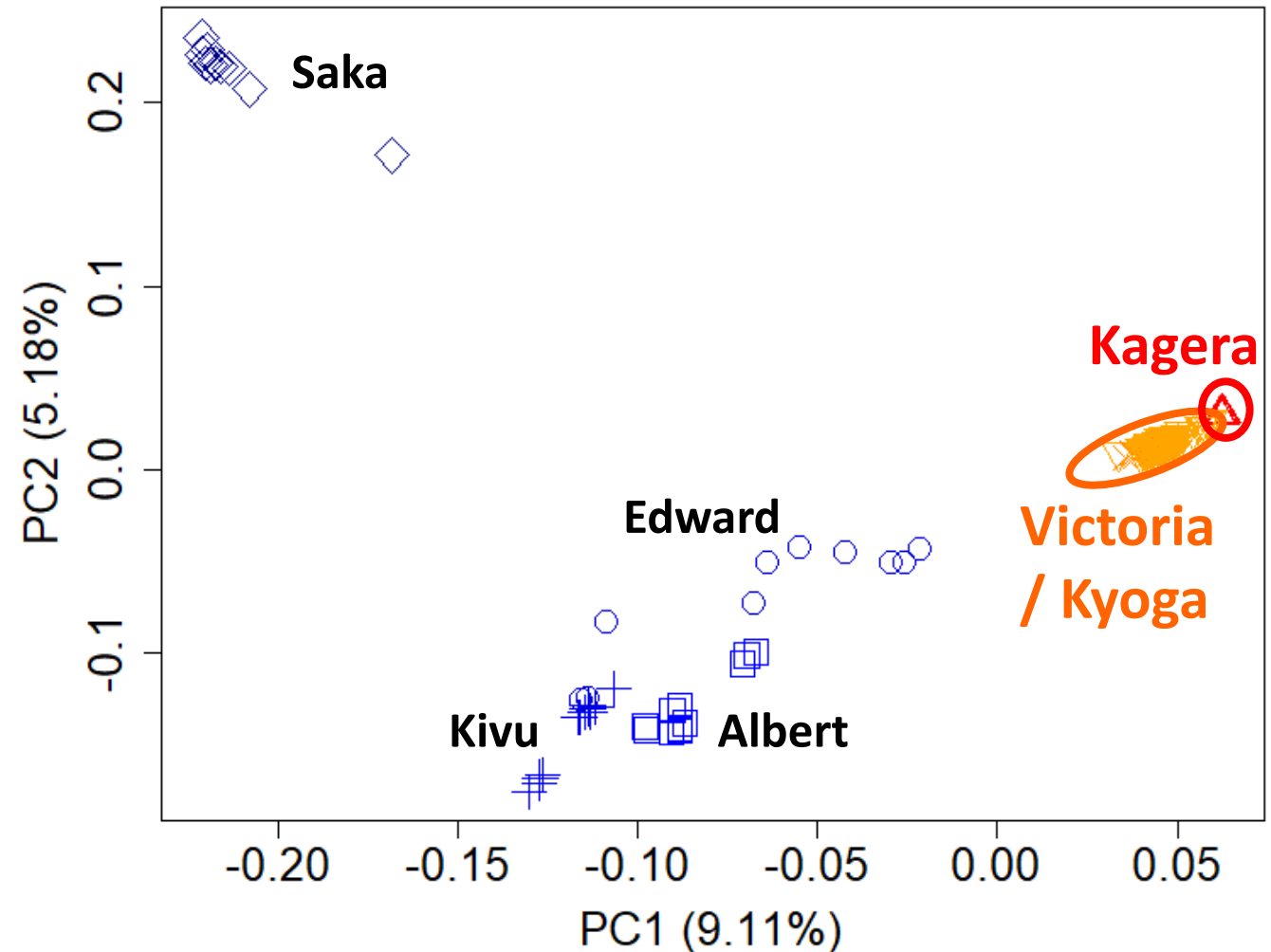
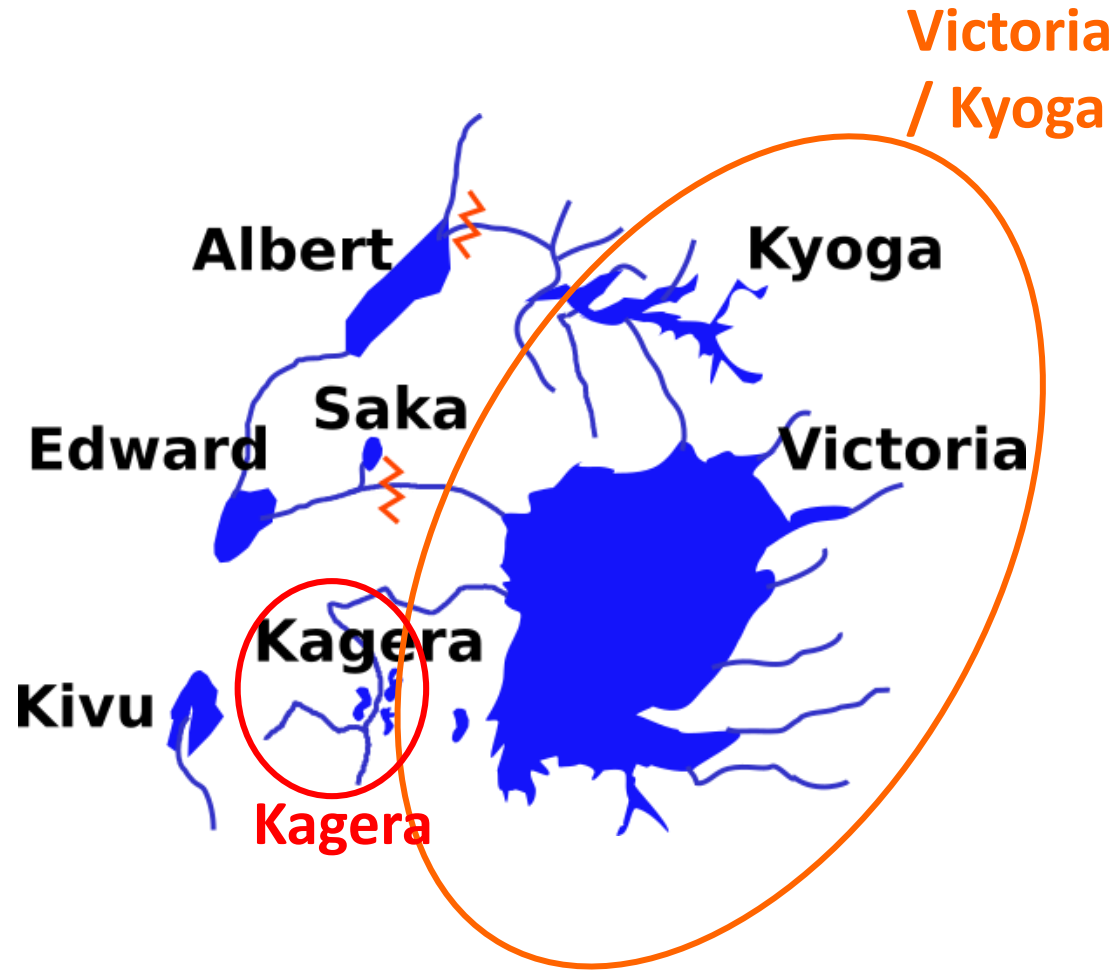
# PCA on whole-genomes separates Lake Victoria/Kyoga cichlids from others

(152 genomes, 1.6M LD-pruned SNPs)



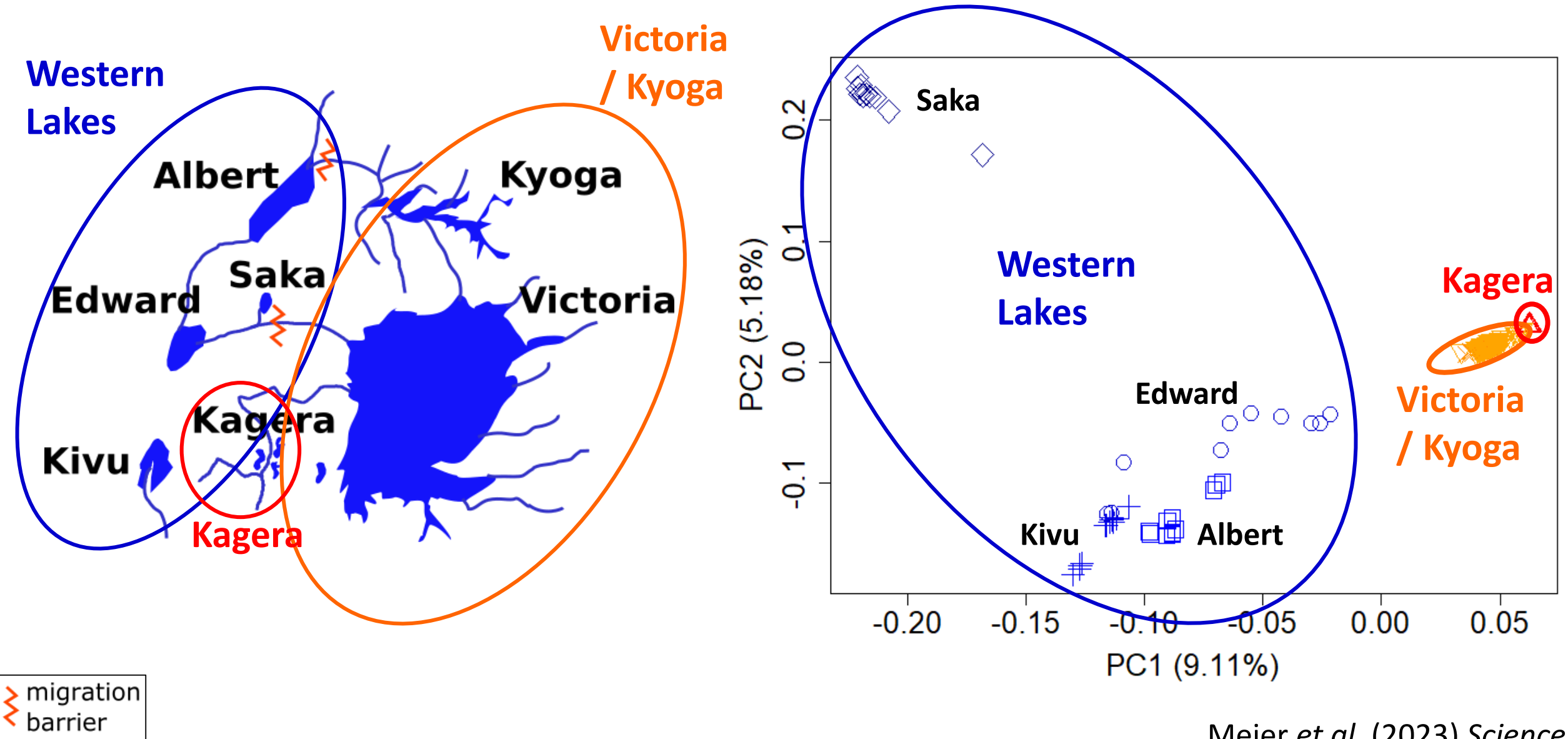
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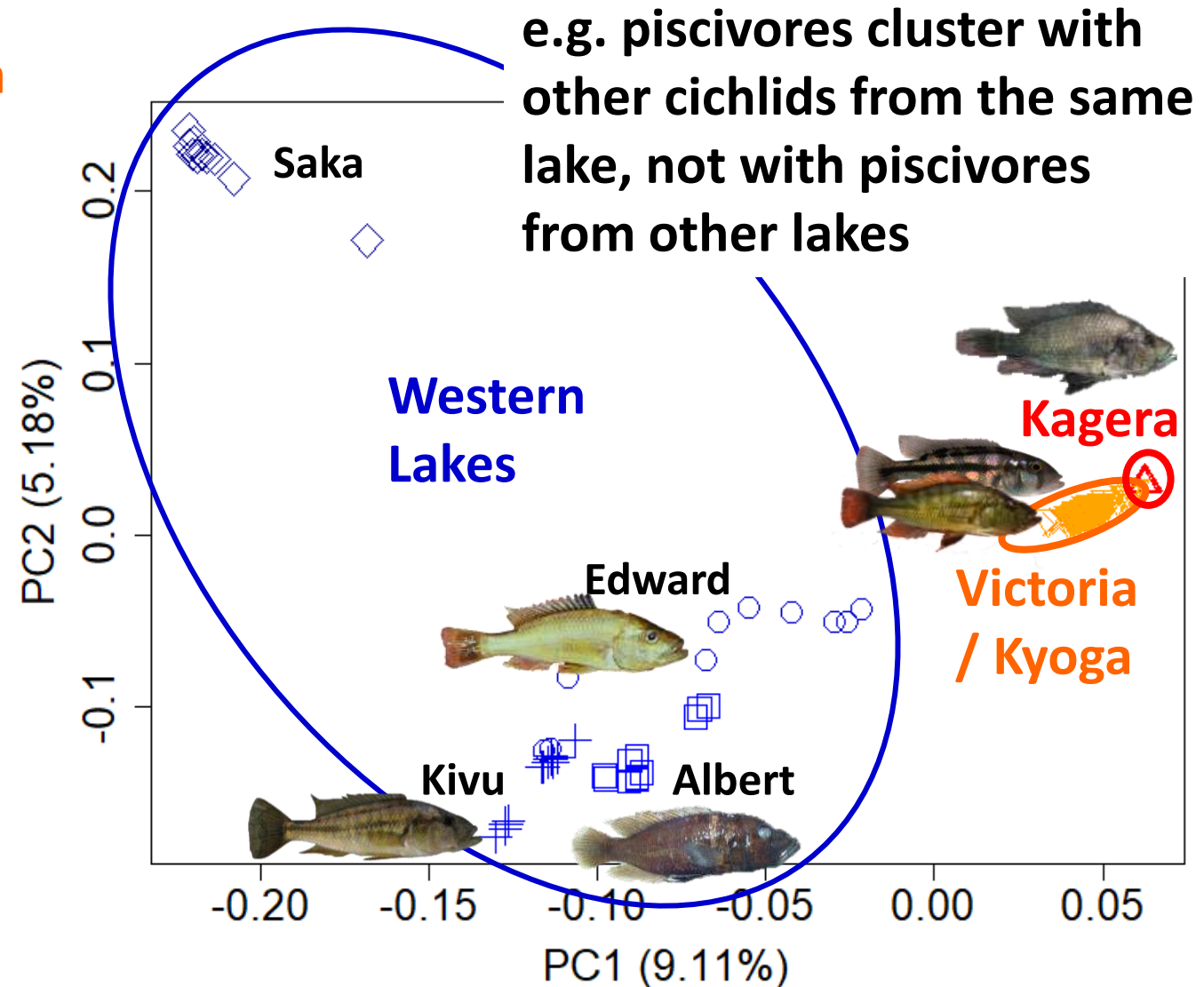
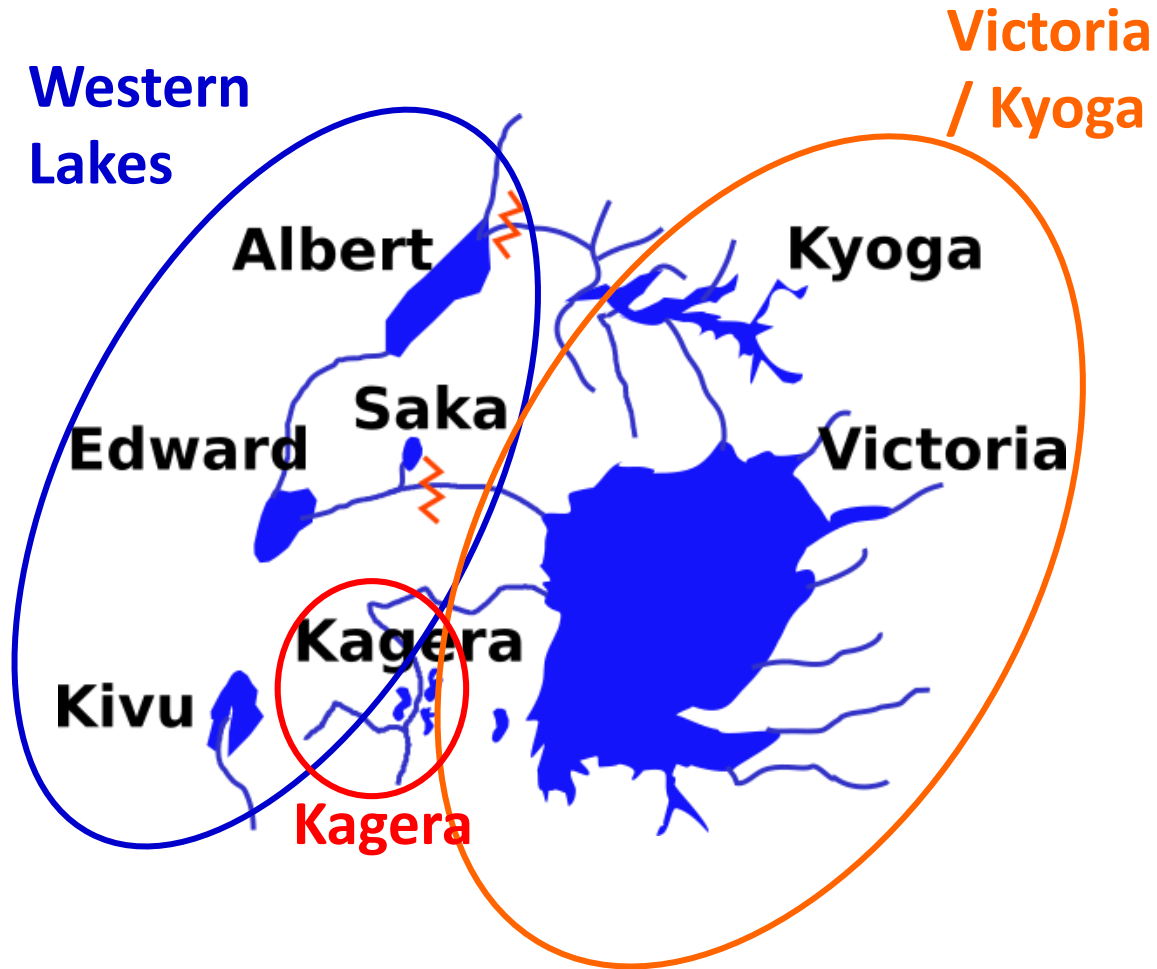
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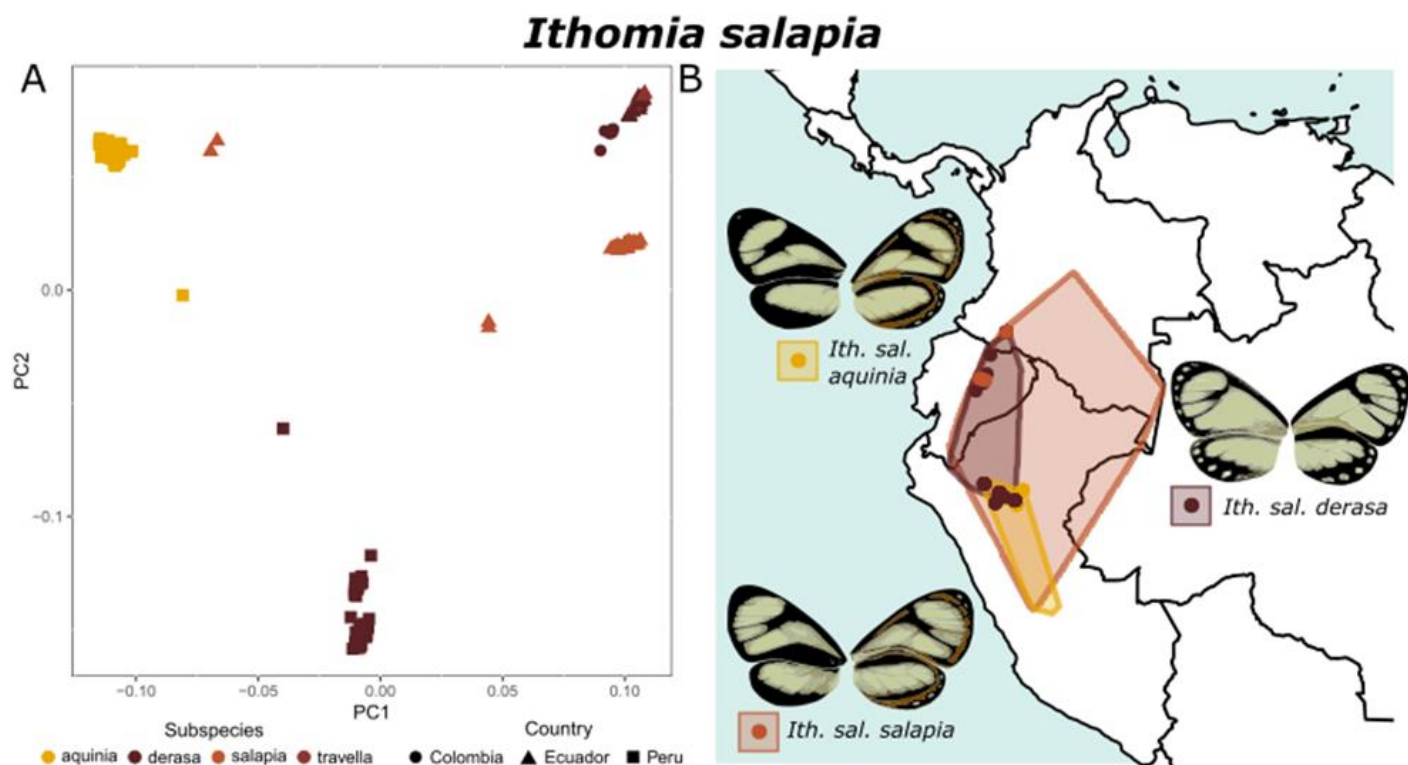
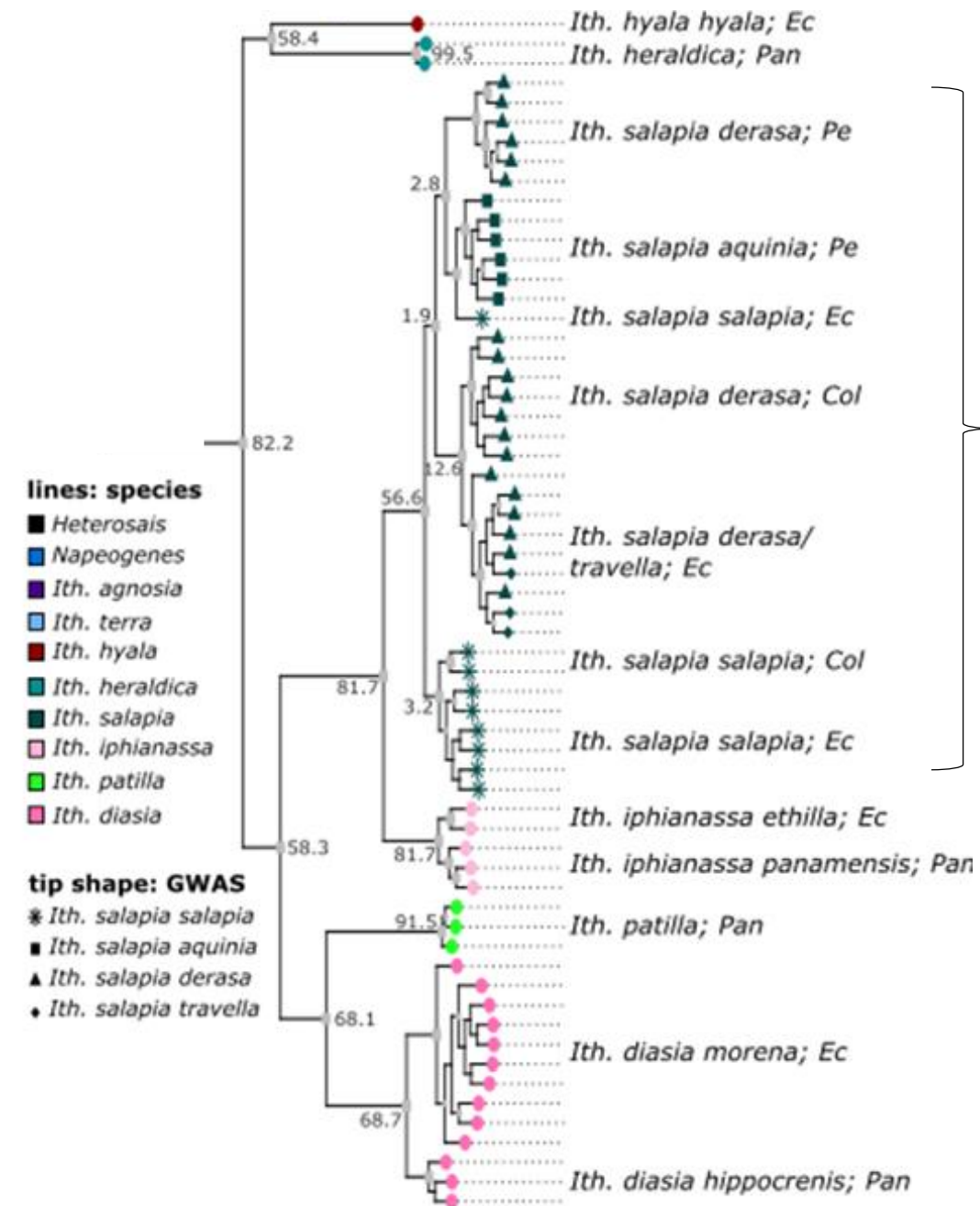
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# Case studies to discuss

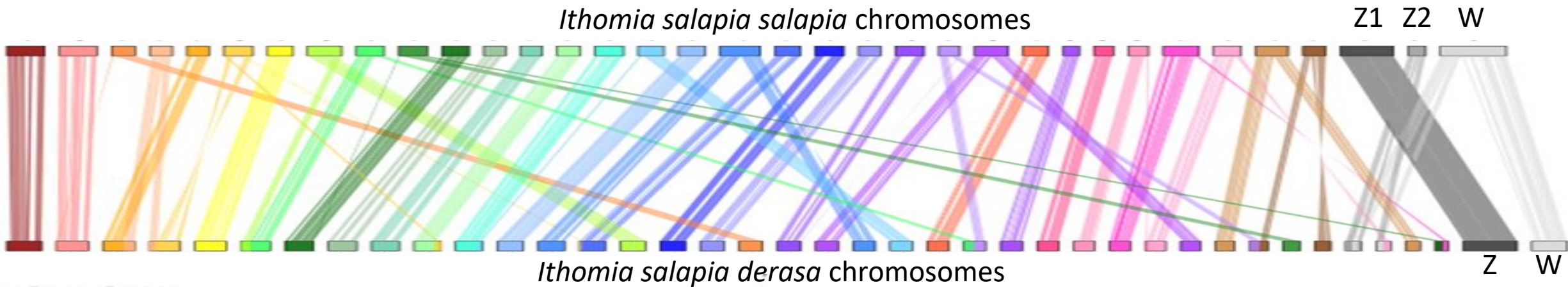
Three examples with very different optimal sampling designs

- Identifying barriers to gene flow and the genetic basis of relevant traits in a hybrid zone
- Inferring if a fish species community evolved in a lake or if the different species independently colonised the lake
- **Inferring if chromosomal rearrangements contribute to speciation in a butterfly species complex**

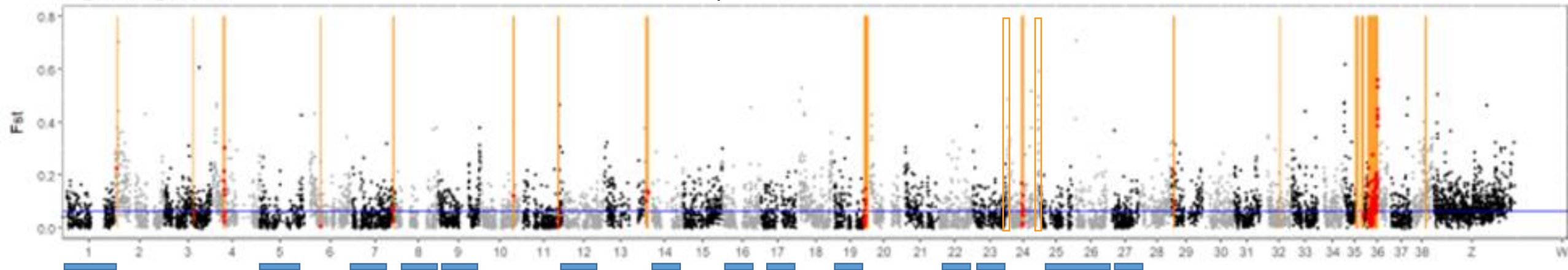


# *Ithomia salapia* «subspecies» show massive chromosomal rearrangements, likely contributing to speciation

*Ithomia salapia salapia* chromosomes



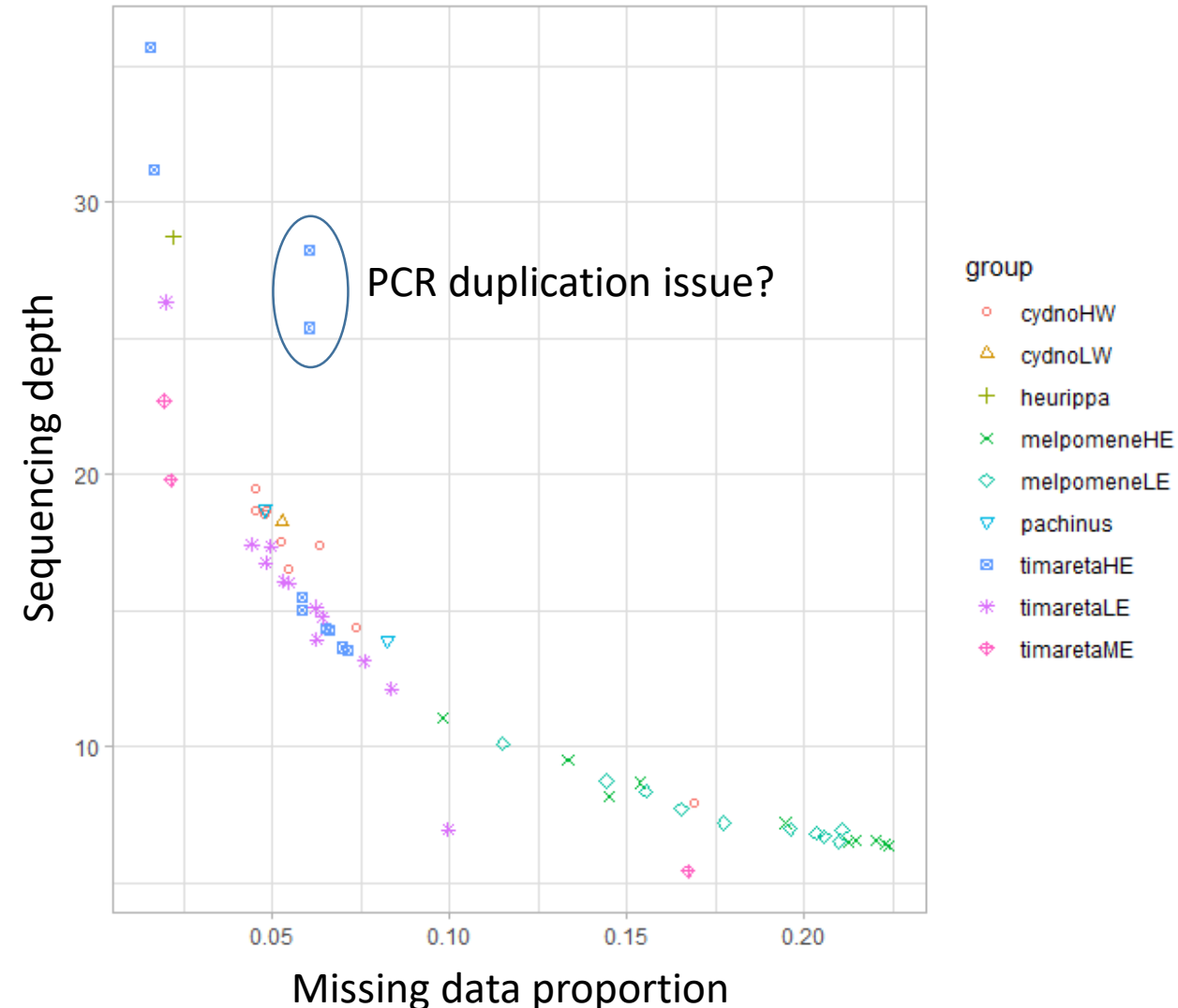
*Ithomia salapia derasa* chromosomes



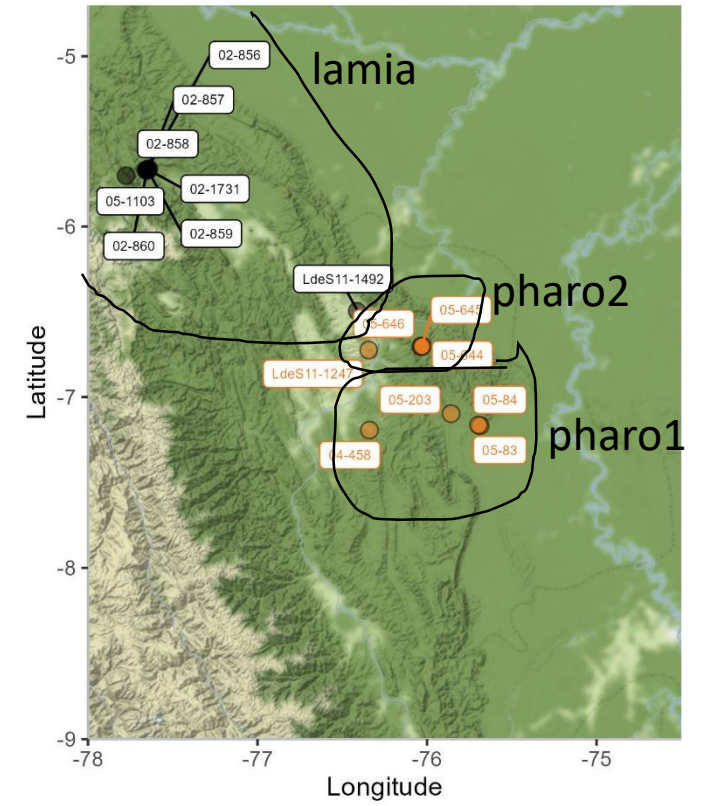
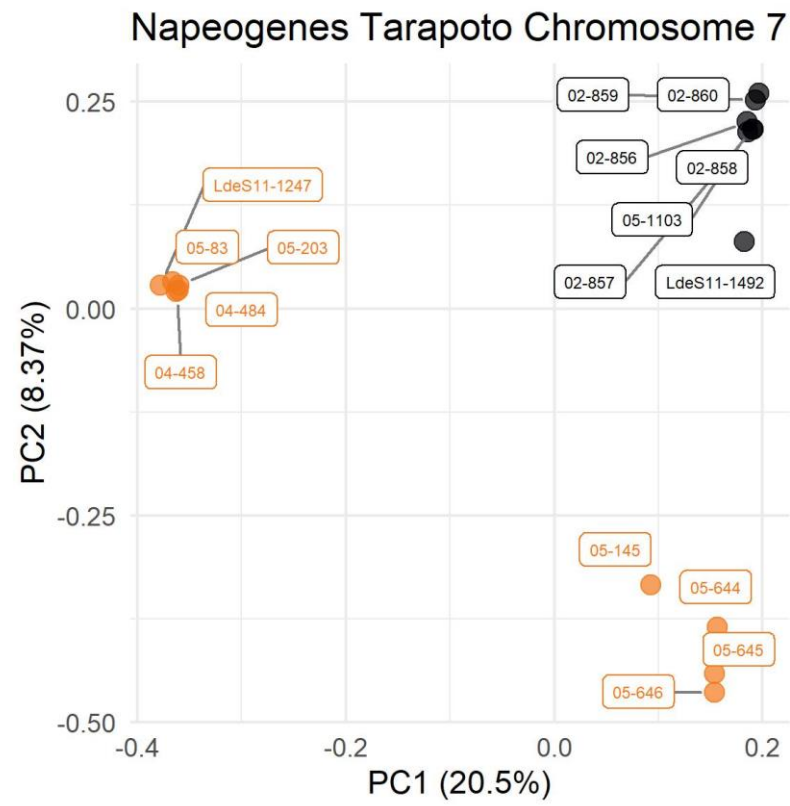
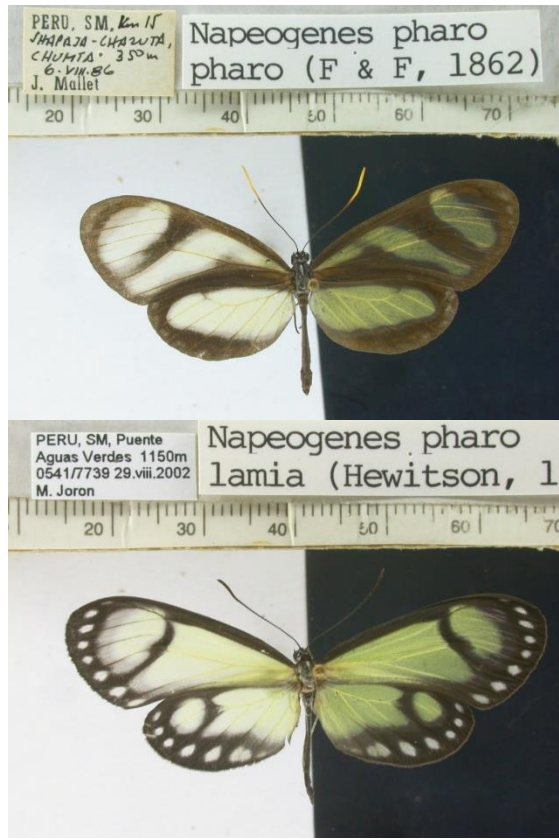
# Once you get your data, make sure you trust it

Get a feeling for your data, do quality checks at all stages and plot your data

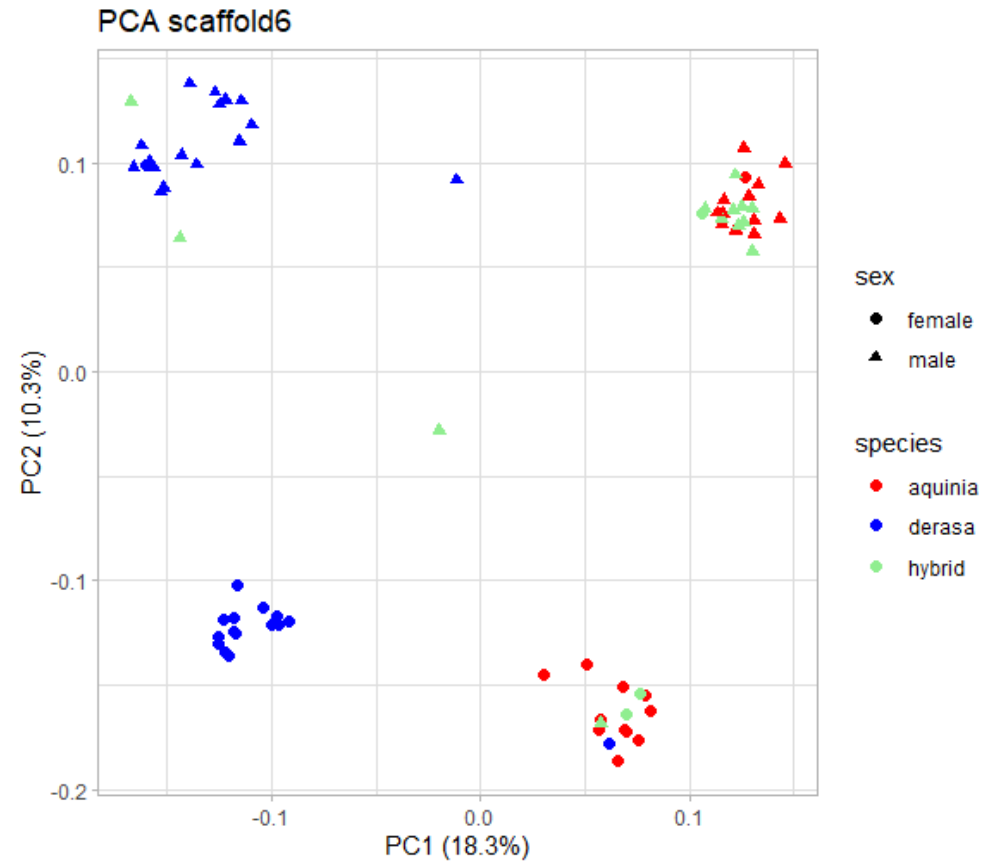
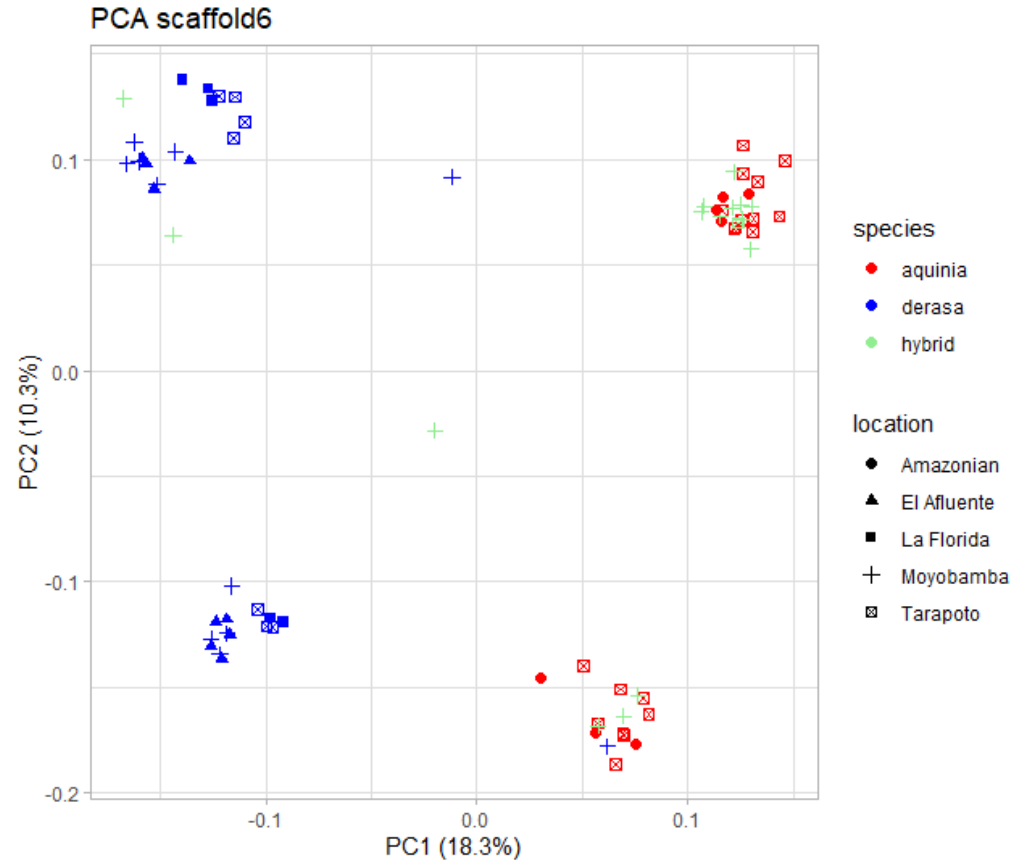
- Do any individuals show low mapping proportion?
- Do any individuals show high proportion of missing data or low sequencing depth?



# Once you get your data, make sure you trust it



# Once you get your data, make sure you trust it



*Ithomia salapia aquinia* (Amazon)



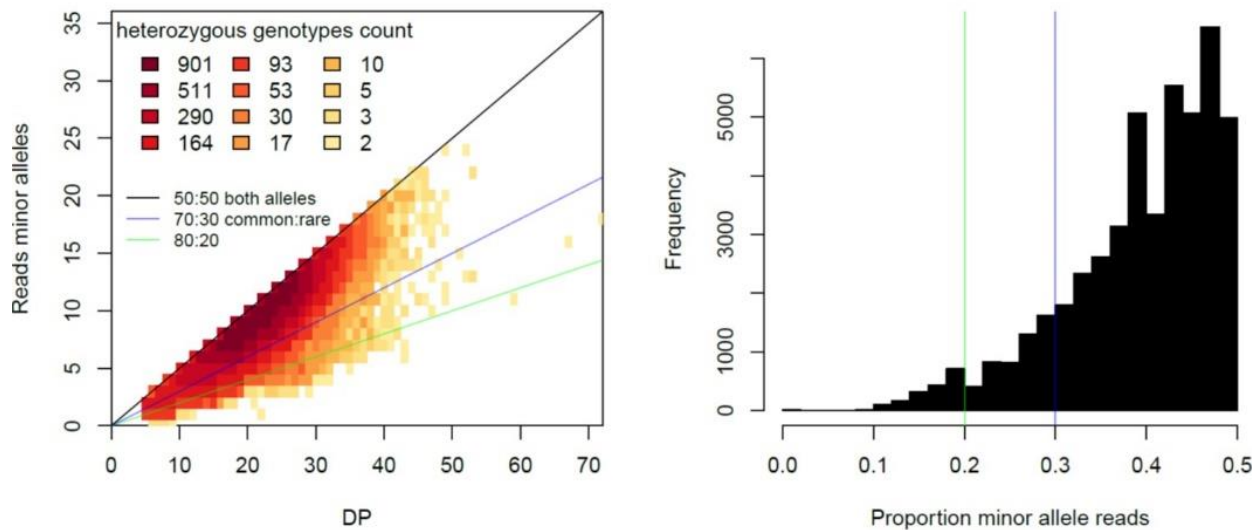
*Ithomia salapia derasa* (Andes)



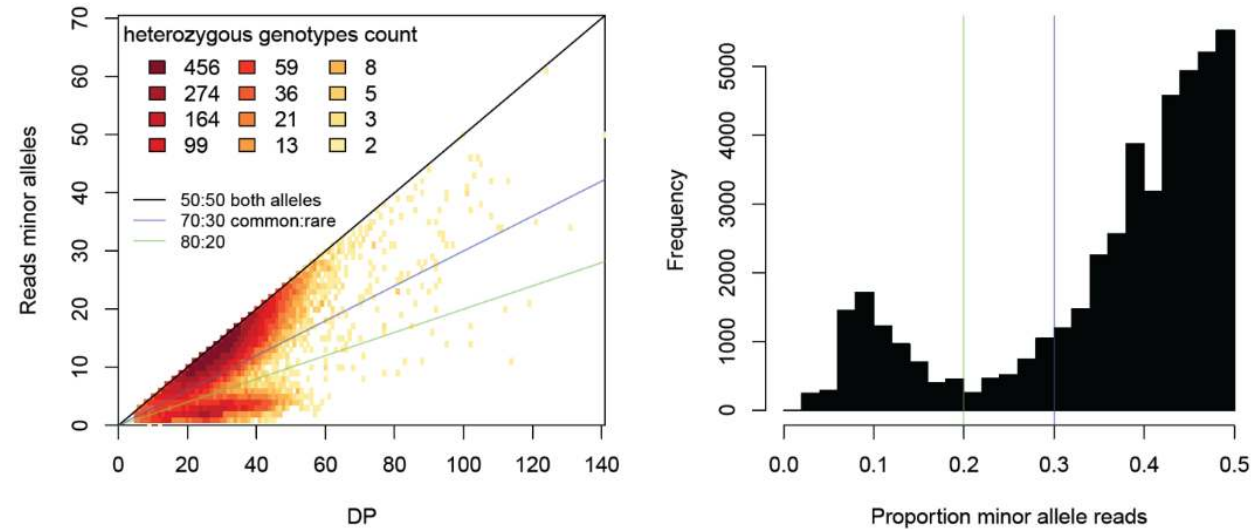
# Once you get your data, make sure you trust it

Contamination can cause issues, including erroneous inference of introgression

well-sequenced individual

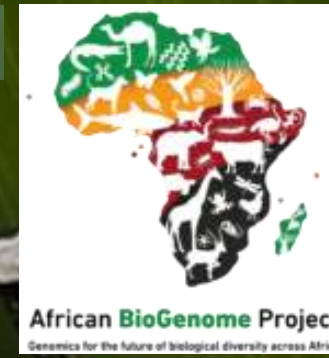


Individual with likely contamination





Project Psyche



ATLASea  
Atlas des génomes marins



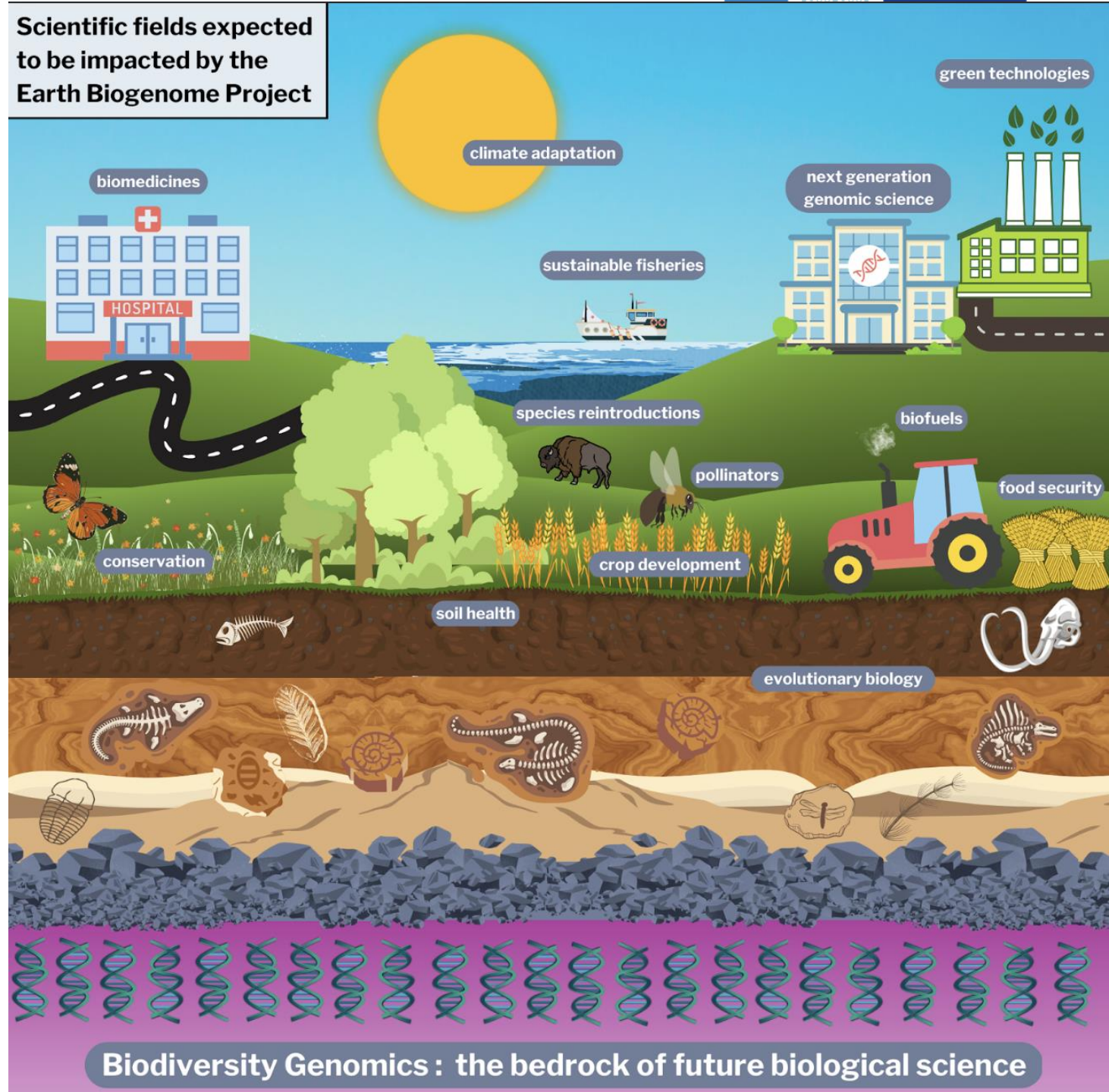
CREATING A NEW FOUNDATION FOR BIOLOGY

# Sequencing Life for the Future of Life

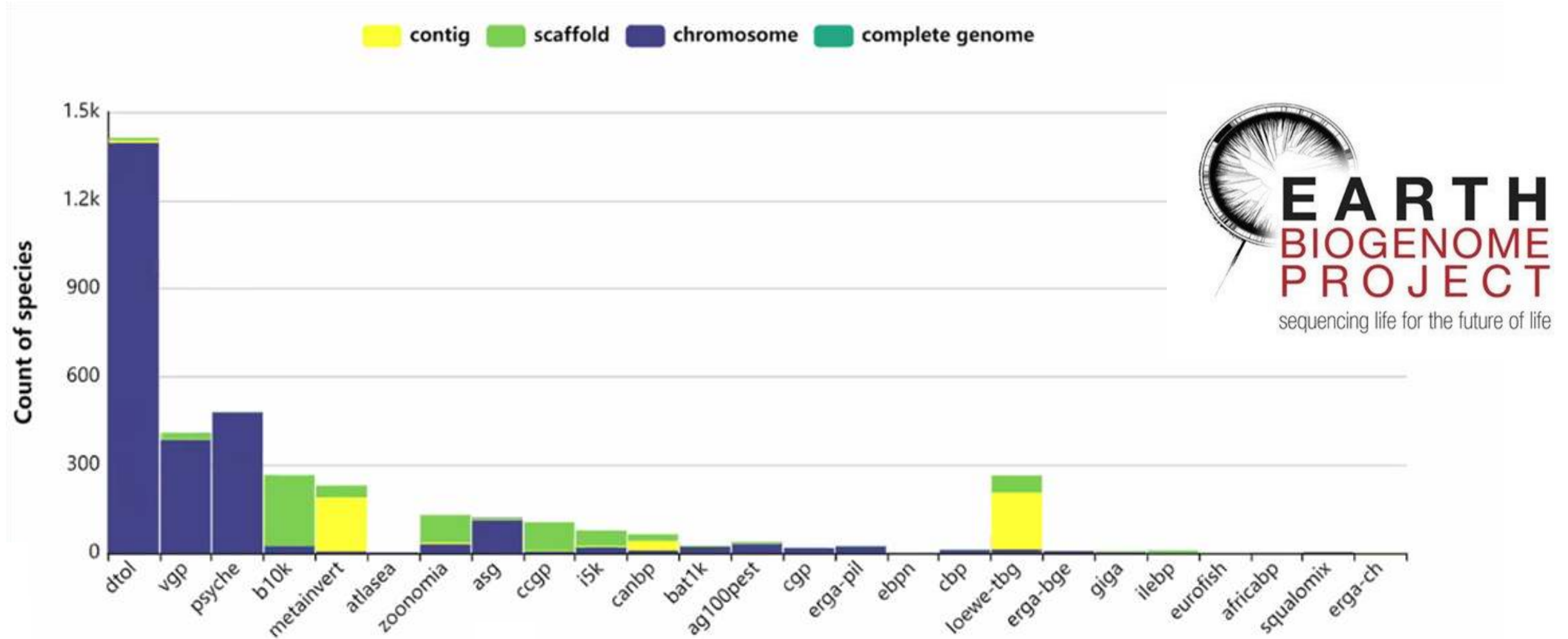


# Reference genomes have global impact in a wide range of areas important for planetary health:

- Biomedicines
- Climate adaptation
- Biomonitoring
- Green technologies
- Next generation genomic science
- Sustainable fisheries
- Species reintroductions
- Conservation
- Pollinators
- Biofuels
- Food security
- Crop development
- Soil health
- Evolutionary biology



# Large reference genome projects



# Major Projects at the Tree of Life Programme at the Wellcome Sanger Institute, Cambridge, UK



## ★ **Darwin Tree of Life Project**

- 70,000 species from Britain and Ireland [funded for ~8,000 species to 2026 currently]



## ★ **Aquatic Symbiosis Genomics**

- 1,000 species (500 symbiotic systems) from marine and freshwater



## ★ **Project Psyche – Lepidoptera genomes for Europe**

- Sequencing all 11,000 moths and butterflies of Europe



## ★ **Vertebrate Genomes Project**

- VGP Phase 1 (ordinal - 260 species) and Phase 2 (family) goals



## ★ **European Reference Genome Atlas**

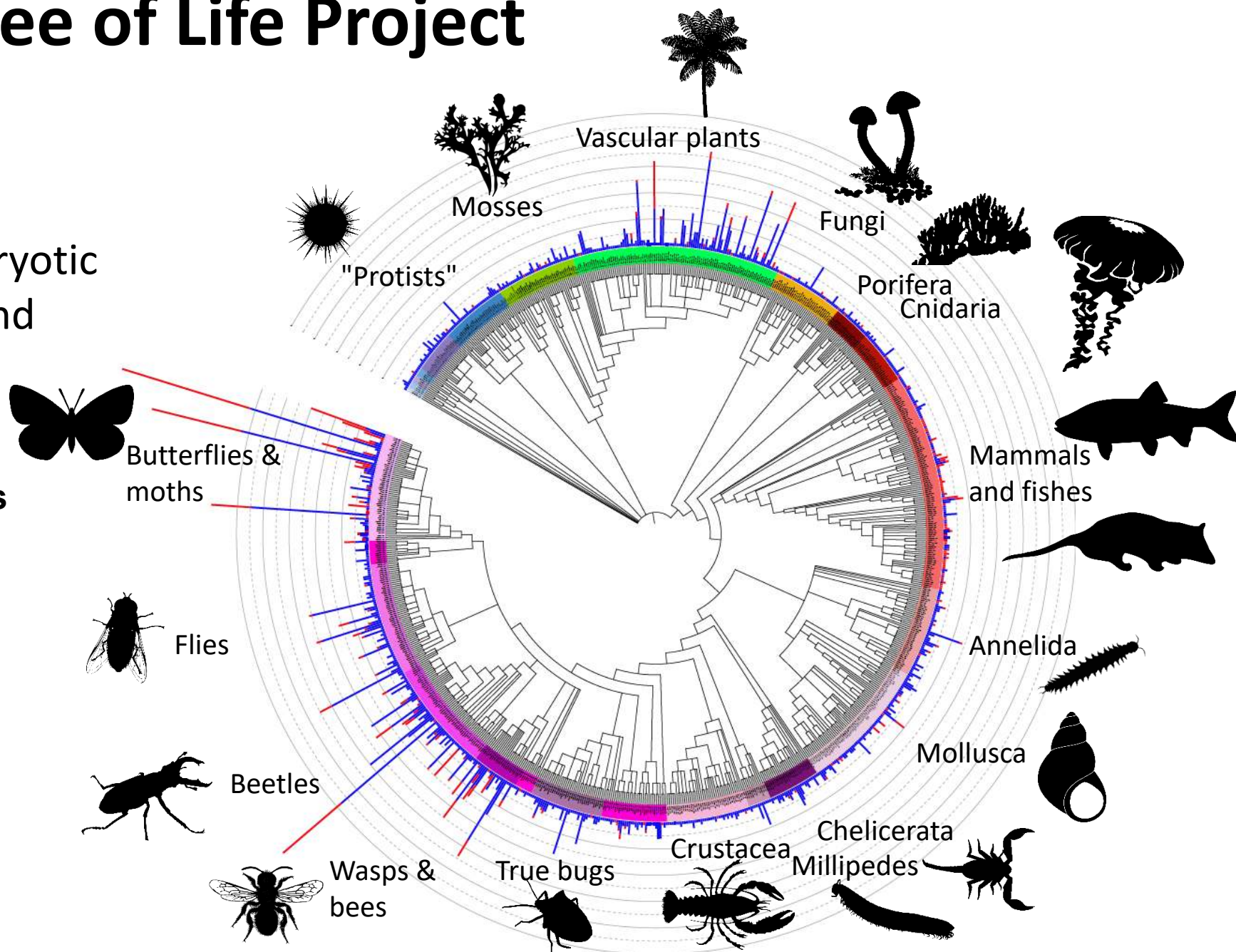
- Sequencing the genomes of all species in the European continent (~500 species)



# Darwin Tree of Life Project

sequence all ~70,000 eukaryotic species in Britain and Ireland

 >5000 species in progress  
 >1500 genomes complete





## Project Psyche

Generate and explore reference genomes for all **~11,000 species** of **butterflies and moths (Lepidoptera)** of **Europe**.

Named after Psyche -  
Greek goddess of the soul,  
often depicted with butterfly  
wings



Credit: Sidon, Sarcophagus relief of  
Psyche. Livius.org

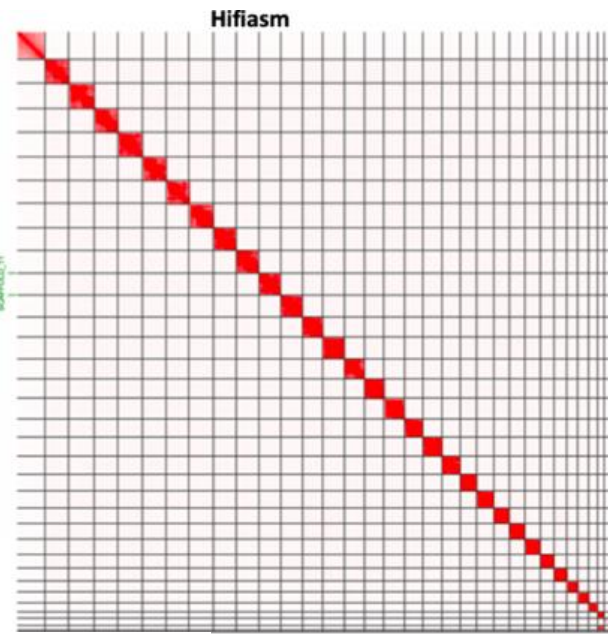
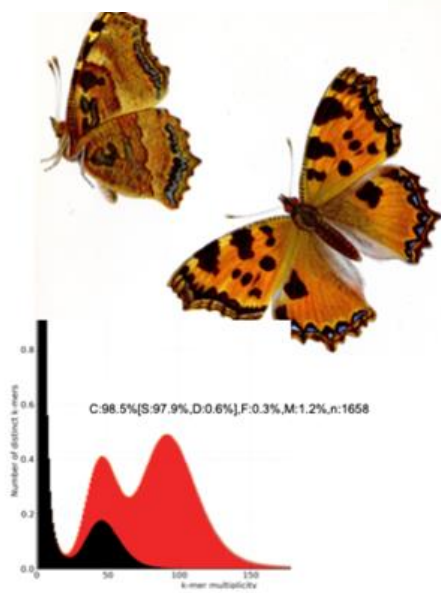


# Why Lepidoptera?

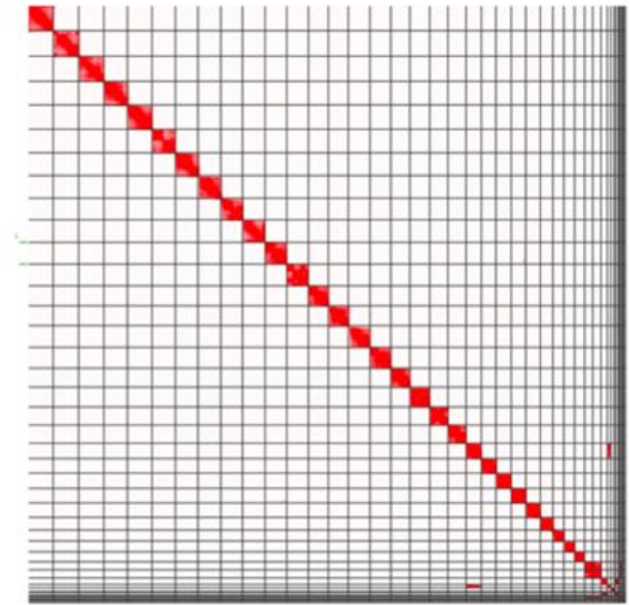
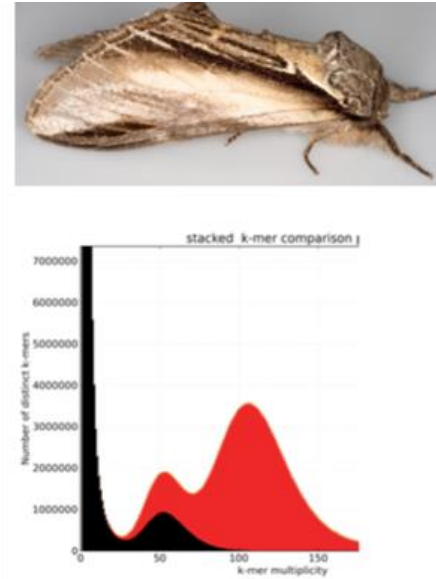


# Lepidoptera genomes tend to assemble very easily

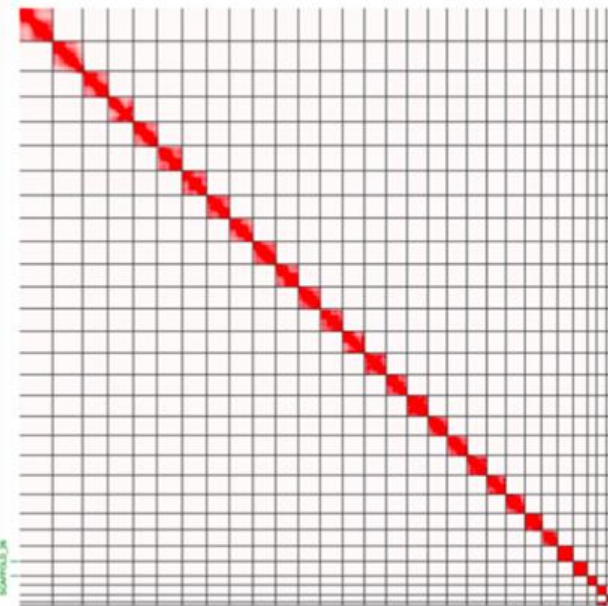
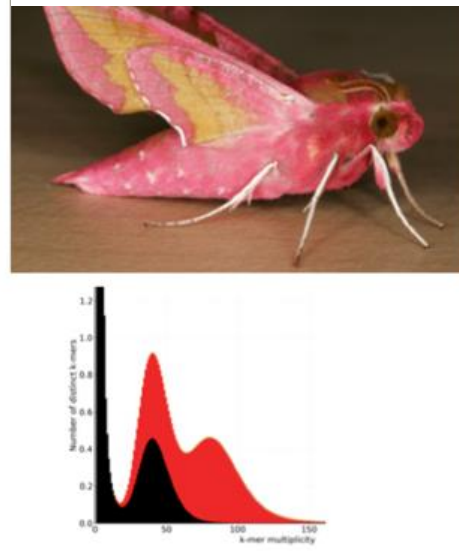
*Nymphalis polychloros*



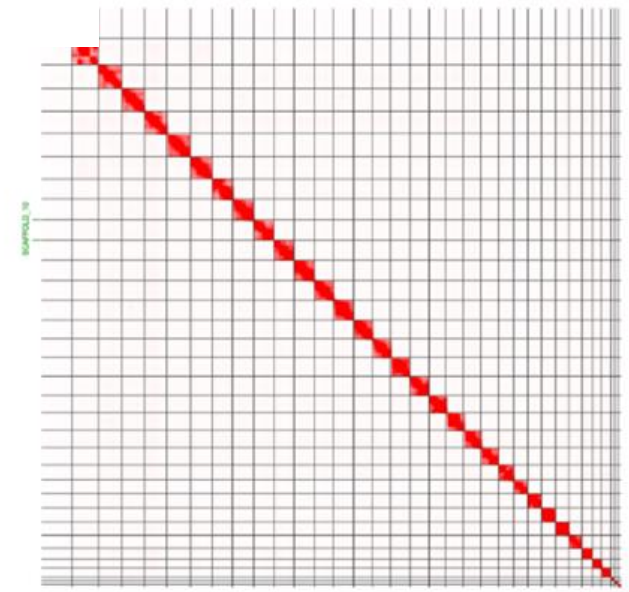
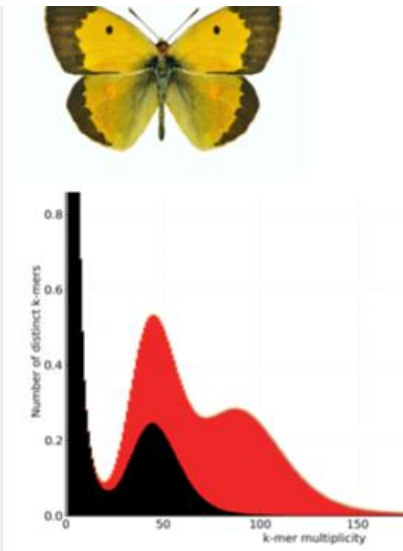
*Pheosia tremula*



*Deilephila porcellus*



*Colias croceus*



# Why sequence all species of Lepidoptera in Europe?

To answer wide-reaching questions:

- Chromosome evolution
- Evolution of opsin genes
- Genetic basis of migration
- Population genetics
- Genetic diversity predictors
- & many more...

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Article | [Open access](#) | Published: 21 February 2024

### Comparative genomics reveals the dynamics of chromosome evolution in Lepidoptera

[Charlotte J. Wright](#) ✉, [Lewis Stevens](#), [Alexander Mackintosh](#), [Mara Lawniczak](#) & [Mark Blaxter](#) ✉

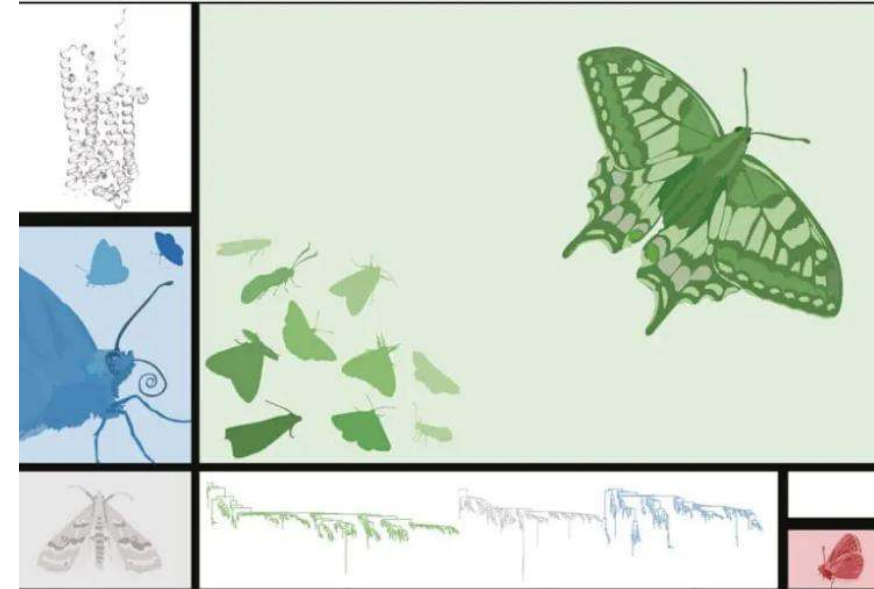
[Nature Ecology & Evolution](#) **8**, 777–790 (2024) | [Cite this article](#)

**16k** Accesses | **13** Citations | **255** Altmetric | [Metrics](#)

Volume 40 • Number 12 • December 2023

# MOLECULAR BIOLOGY AND EVOLUTION

academic.oup.com/mbe



## Opsin Gene Duplication in Lepidoptera: Retrotransposition, Sex Linkage, and Gene Expression

[Peter O Mulhair](#) ✉, [Liam Crowley](#), [Douglas H Boyes](#), [Owen T Lewis](#), [Peter W H Holland](#) [Author Notes](#)

*Molecular Biology and Evolution*, Volume 40, Issue 11, November 2023,

# Psyche community



Charlotte Wright



Mark Blaxter

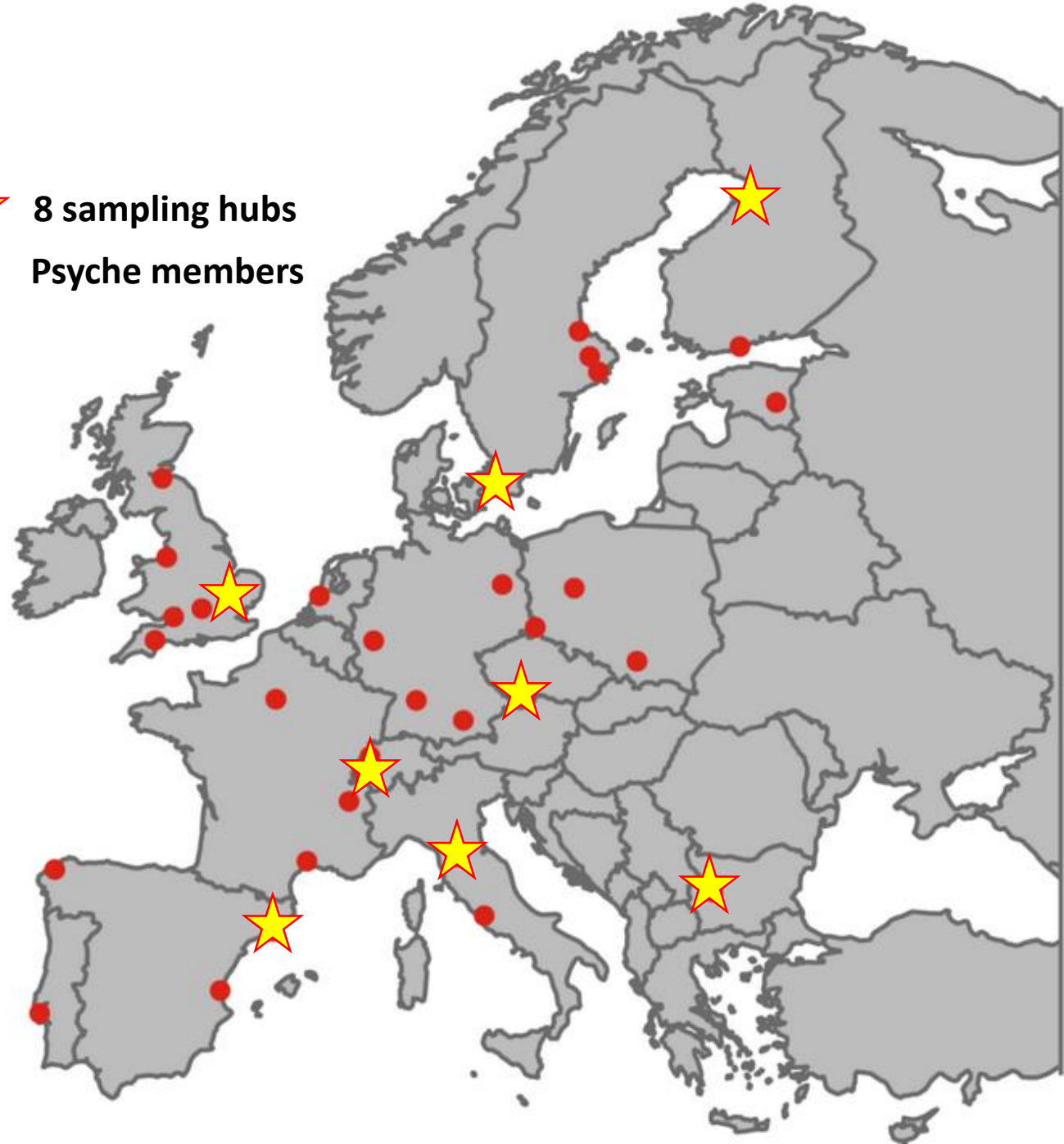


Joana Meier

Join us if you are interested  
in butterflies and moths



- ★ 8 sampling hubs
- Psyche members





# Phases of Project Psyche

Generate and explore reference genomes for all **~11,000 species** of **butterflies and moths (Lepidoptera)** of **Europe**.

## Phase 1: Generate 2,000 genomes

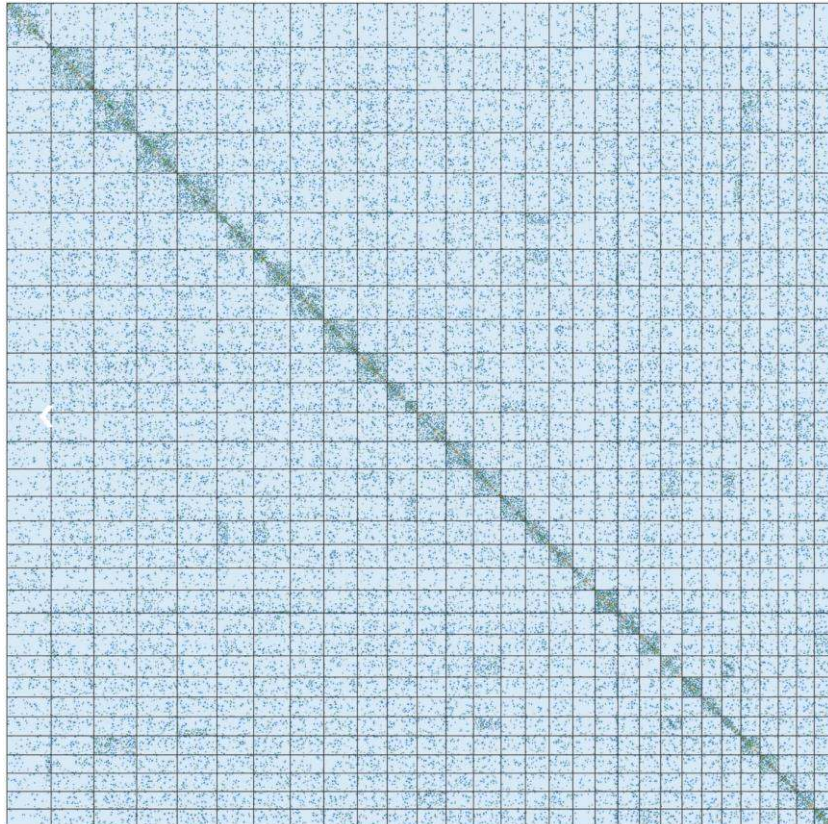
- Focus on taxonomic diversity
- Samples from sampling hubs
- Sequencing at the Wellcome Sanger Institute
- Writing a whitepaper to celebrate the first 1000 genomes
- Writing a set of 3-5 publications on the first 1000 genomes

## Plans for later phases:

- Establishing more sampling hubs
- Establishing more sequencing hubs

# Genome assembly of the moth and all its co-bionts

***Phalera bucephala*:**  
one moth, five genomes



31 nuclear chromosomes



mitochondrial  
genome

*Wolbachia* A, B1, B2

# Openly published genomes and Genome Notes: Collector(s) as first author(s)

<https://www.darwintreeoflife.org/genomes/genome-notes>

Wellcome Open Research

Wellcome Open Research 2023, 8:278 Last updated: 18 SEP 2023



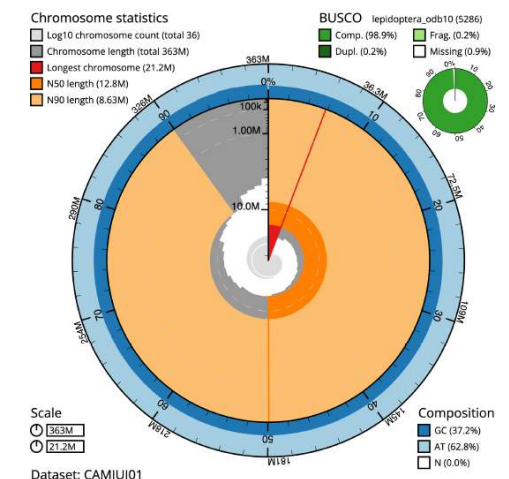
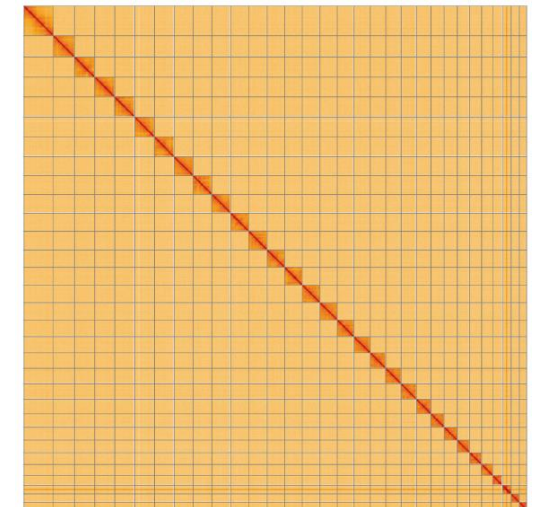
DATA NOTE

## The genome sequence of the Dark Spectacle, *Abrostola triplasia* (Linnaeus, 1758) [version 1; peer review: 2 approved]

Douglas Boyes<sup>1+</sup>, Owen T. Lewis<sup>2</sup>,  
University of Oxford and Wytham Woods Genome Acquisition Lab,  
Darwin Tree of Life Barcoding collective,  
Wellcome Sanger Institute Tree of Life programme,  
Wellcome Sanger Institute Scientific Operations: DNA Pipelines collective,  
Tree of Life Core Informatics collective, Darwin Tree of Life Consortium

<sup>1</sup>UK Centre for Ecology & Hydrology, Wallingford, England, UK

<sup>2</sup>University of Oxford, Oxford, England, UK



# Psyche enables LepEU: population genomics across Lepidoptera

LepEU: European Lepidopteran  
Population Genomics Consortia

<https://lepeu.github.io/>

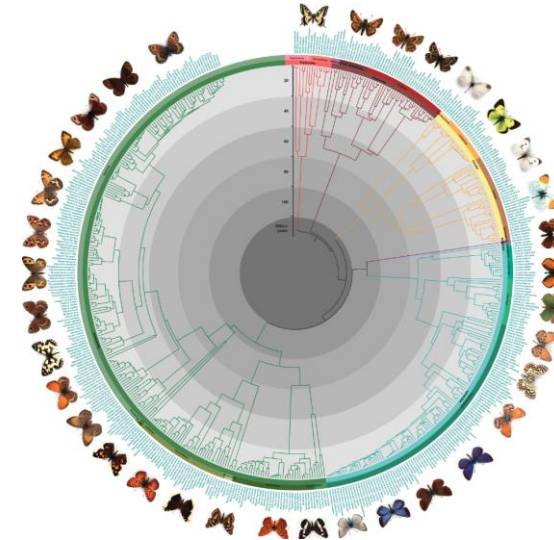
*Quantifying regional genetic diversity, inbreeding,  
and local adaptation of leps to inform biodiversity  
management in Europe*



**Project Psyche**  
Reference genomes of  
all European Lepidoptera



**Patrícia Beldade**  
Lisbon University, Portugal



**Chris Wheat**  
Stockholm University,  
Sweden

## COST action:

### Lep10KGenomes: Utilizing 10,000 genomes of European Lepidoptera

- Organise workshops and training courses
- Research exchanges for early career researchers
- Work with external stakeholders to address societal needs using genomic information
- Devise best practices
  - For reference genomes - **Psyche**
  - For population genomics - **LepEU**



**Niklas Wahlberg**  
Lund University,  
Sweden



Funded by the Horizon 2020 Framework Programme  
of the European Union

Find out more here:

<https://tinyurl.com/Lep10k-COST>

Join Project Psyche here:

<https://tinyurl.com/projectpsyche>

**How to generate a  
reference genome?**

# Check if a reference genome exists



Genomes on a Tree ([GoaT](https://goat.genomehubs.org/), <https://goat.genomehubs.org/>)



## Genomes on a Tree (GoaT)

**Genomes on a Tree (GoaT): A versatile, scalable search engine for genomic and sequencing project metadata across the eukaryotic tree of life.** Challis *et al.* 2023. Wellcome Open Res 2023, **8**:24. [doi:10.12688/wellcomeopenres.18658.1](https://doi.org/10.12688/wellcomeopenres.18658.1)

GoaT needs been built using [GenomeHubs](#) to help coordinate efforts across the [Earth Biogenome Project](#) (EBP) Network at all stages from planning through sequencing and assembly to publication. [read more...](#)

## Search GoaT

include descendants

☐

Off

include estimates

☒

On

empty columns

☐

Off

result columns

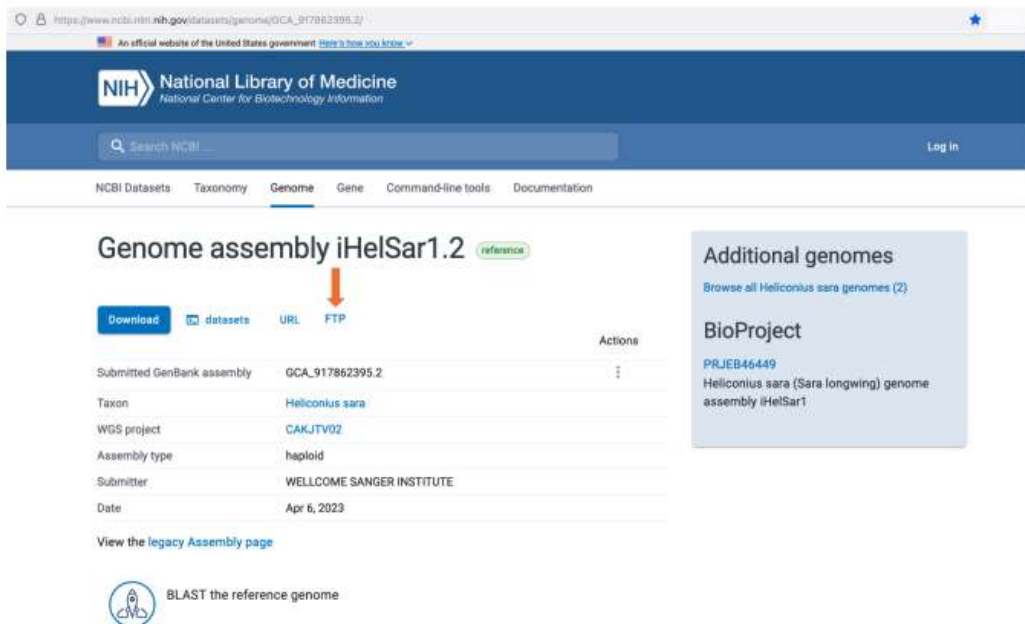
☐

query builder

clear all

# First, check if a reference genome already exists

- Genomes on a Tree ([GoaT, https://goat.genomehubs.org/](https://goat.genomehubs.org/))
- If yes, get it e.g. from NCBI ([www.ncbi.nlm.nih.gov/home/genomes](https://www.ncbi.nlm.nih.gov/home/genomes))



https://www.ncbi.nlm.nih.gov/datasets/genome/GCA\_917862395.2/

National Library of Medicine  
National Center for Biotechnology Information

Search NCBI

Log In

NCBI Datasets Taxonomy Genome Gene Command-line tools Documentation

### Genome assembly iHelSar1.2 reference

**Download** datasets URL FTP Actions

|                            |                           |  |
|----------------------------|---------------------------|--|
| Submitted GenBank assembly | GCA_917862395.2           |  |
| Taxon                      | Heliconius sara           |  |
| WGS project                | CAKJTV02                  |  |
| Assembly type              | haploid                   |  |
| Submitter                  | WELLCOME SANGER INSTITUTE |  |
| Date                       | Apr 6, 2023               |  |

[View the legacy Assembly page](#)

 BLAST the reference genome



https://ftp.ncbi.nlm.nih.gov/genomes/all/GCA/917/862/395/GCA\_917862395.2\_iHelSar1.2/

### Index of /genomes/all/GCA/917/862/395/GCA\_917862395.2\_iHelSar1.2

| Name                                           | Last modified    | Size |
|------------------------------------------------|------------------|------|
| Parent Directory                               | -                | -    |
| GCA_917862395.2_iHelSar1.2_assembly_report.txt | 2023-04-18 17:05 | 34K  |
| GCA_917862395.2_iHelSar1.2_assembly_stats.txt  | 2023-04-18 17:05 | 31K  |
| GCA_917862395.2_iHelSar1.2_feature_count.txt   | 2024-02-15 01:58 | 168  |
| GCA_917862395.2_iHelSar1.2_genomic.fna.gz      | 2023-04-18 17:05 | 187M |
| GCA_917862395.2_iHelSar1.2_genomic.gbff.gz     | 2023-04-18 17:05 | 130M |
| GCA_917862395.2_iHelSar1.2_genomic.gaps.txt.gz | 2023-04-18 17:05 | 1.4K |
| GCA_917862395.2_iHelSar1.2_wgmaster.gbff.gz    | 2023-04-18 17:05 | 1.4K |
| README.txt                                     | 2024-04-11 16:11 | 54K  |
| annotation_hashes.txt                          | 2024-02-17 10:02 | 410  |
| assembly_status.txt                            | 2024-07-15 13:53 | 14   |
| md5checksums.txt                               | 2024-02-17 10:02 | 625  |
| uncompressed_checksums.txt                     | 2024-06-14 01:21 | 167  |

[HHS Vulnerability Disclosure](#)

wget

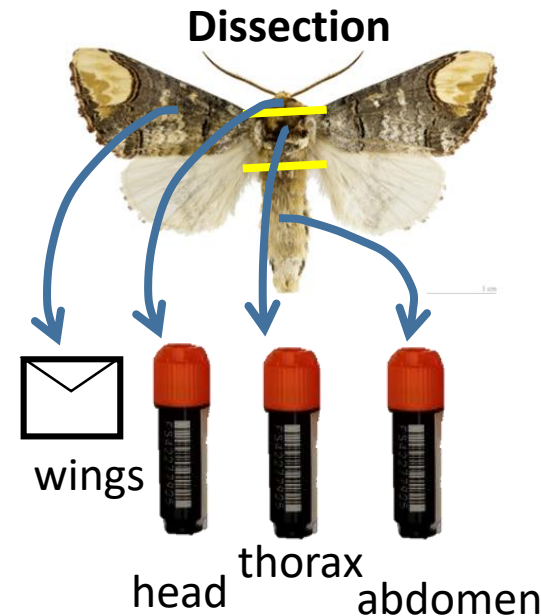
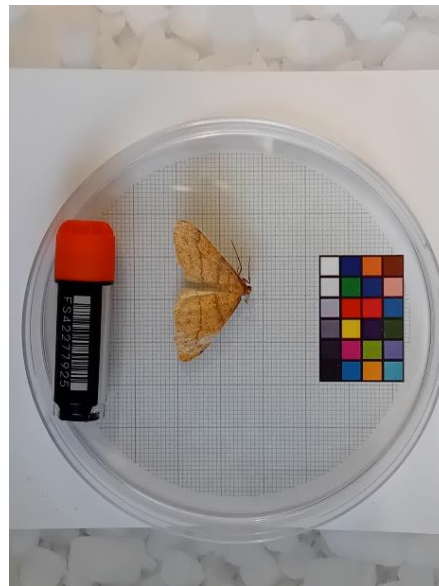
```
https://ftp.ncbi.nlm.nih.gov/genomes/all/GC  
A/917/862/395/GCA_917862395.2_iHelSar1.2/GC  
A_917862395.2_iHelSar1.2_genomic.fna.gz
```

# 1. Getting specimens with well-preserved DNA

- Ideally:
  - Fresh tissue (or blood for birds or fishes)
  - Flash-frozen (in liquid nitrogen or cryogenic dry shipper) and kept at max  $-70^{\circ}\text{C}$
- ensure you have all metadata for the specimen (GPS coordinates, photo of the specimen, etc)

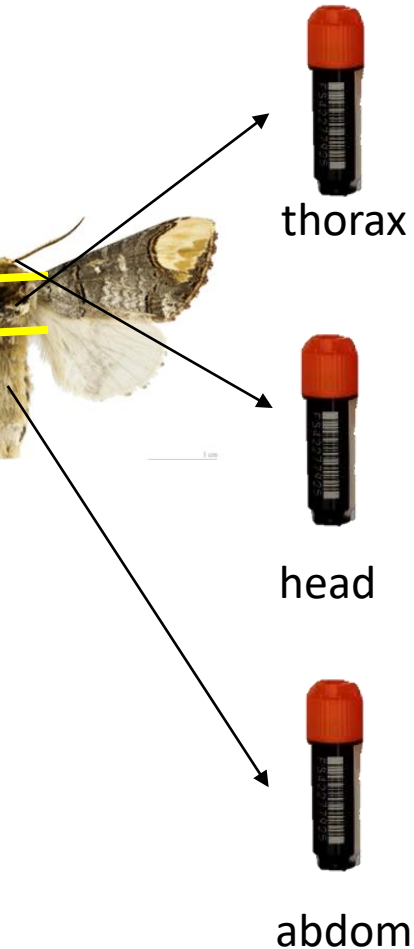


**Dry shipper ( $-150^{\circ}\text{C}$ )**  
(with liquid nitrogen in the walls)



## 2. Sequencing the specimen

For Project Psyche  
we use these tissues:



### PacBio sequencing (long reads) to assemble contigs

ATGTGTCATGGGACATATGTGTCATGGGACATGAGAGAGAGA ← contig = consensus sequence of aligned reads  
GAGAGAGAATGTGTCATGGGACAT  
GGACATATGTGTCATGGGACATGA  
TATGTGTCATGGGACATGAGAGAGAGA

### HiC sequencing to scaffold contigs together

GTGTCATG ATGTGTCATG ATGAGAGAG GAGAGGGA  
ATGTGTCATGGGACATATGTGTCATGGGACATGAGAGAGAGAGAGA GAGAGAGGGACATAAGAGTGTGTCATG  
ATGTGTCATGGGACATATGTGTCATGGGACATGAGAGAGAGAGAGAGANNNGAGAGAGGGACATAAGAGTGTGTCATG ← scaffold = sequence with gaps

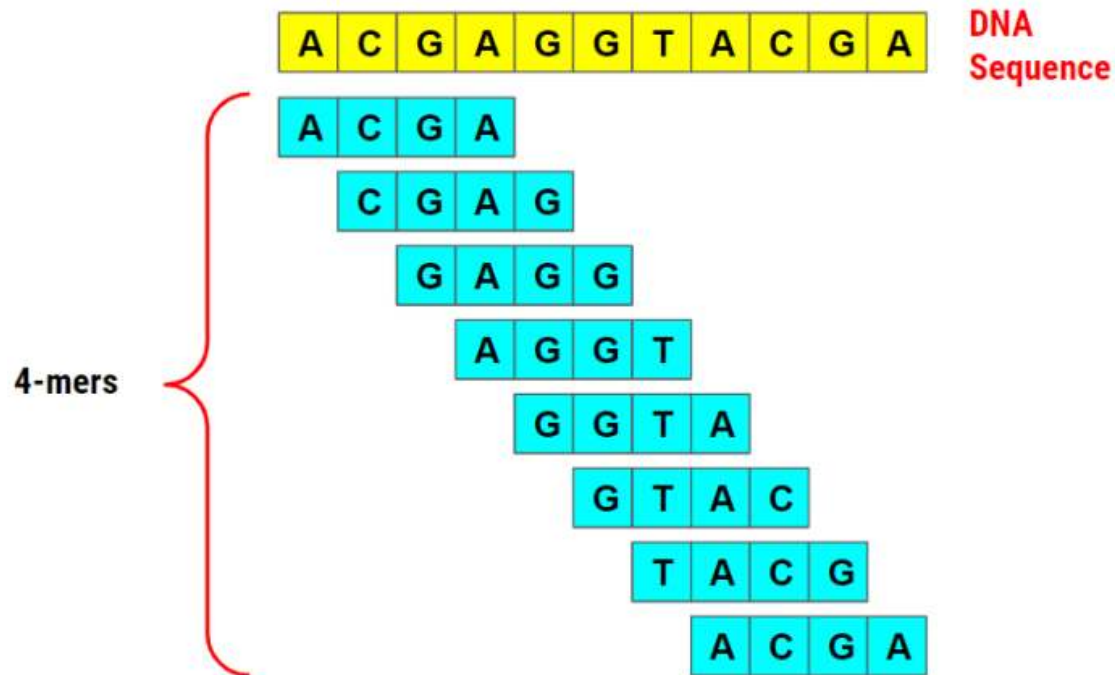
### RNA sequencing for genome annotation

GTGAGTCATGGGAGAGGGACATCAAAAAAAAAAAAAAAAAA  
AGAGGTCATGTGAGTACATCGATCGGAAAAAAAAAAAAAAAAA

For T2T (telomere-to-telomere) genomes,  
add ONT ultra-long reads (>150 kb)

### 3. First statistics with kmer analyses

Estimating the genome size, depth of coverage, ploidy, levels of heterozygosity & duplication, identifying contamination

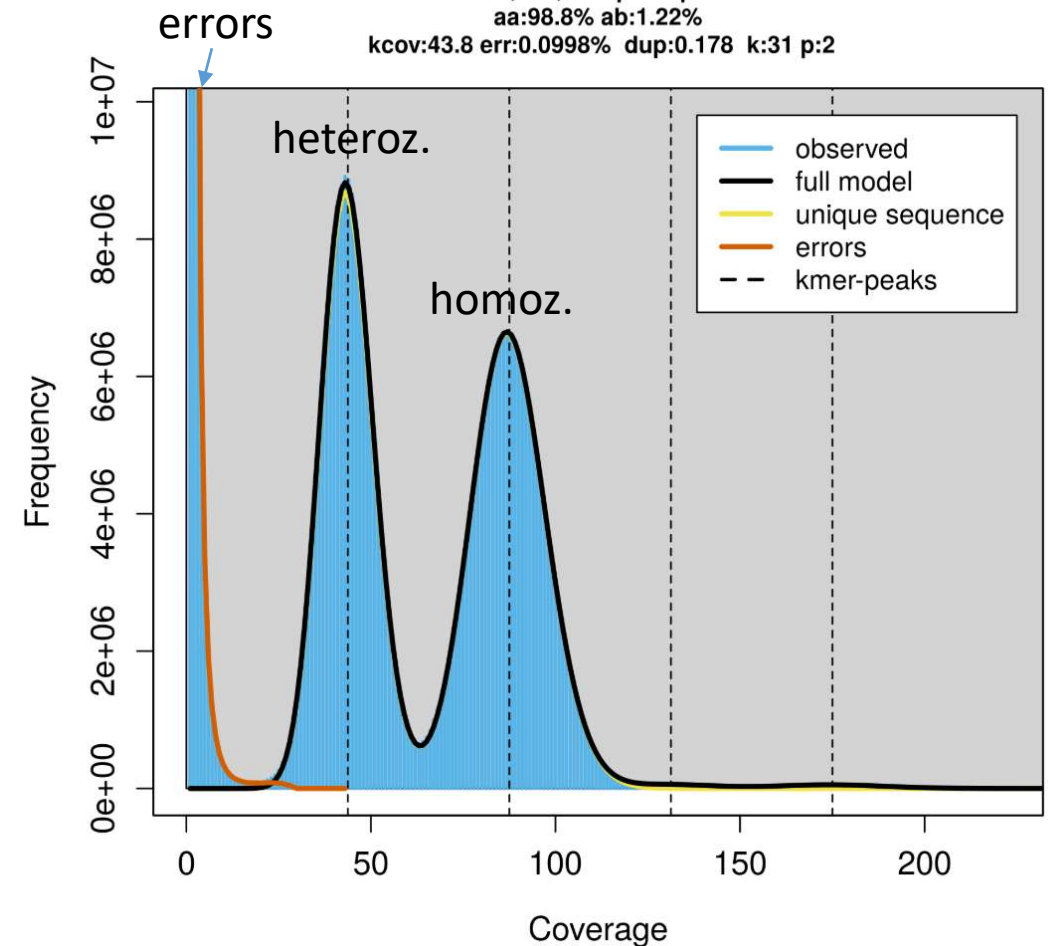


Genomescope

#### Histogram of frequency of k-mers

##### GenomeScope Profile

len:281,671,112bp uniq:87.5%  
aa:98.8% ab:1.22%  
kcov:43.8 err:0.0998% dup:0.178 k:31 p:2



# 4. Assembling the genome

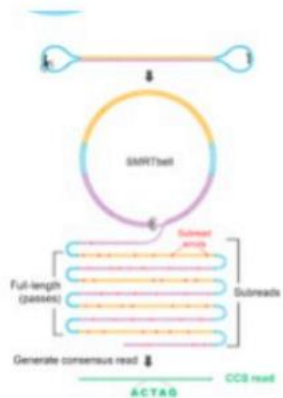
## DToL Current Pipeline

- Sequencing technologies: PacBio HiFi + HiC (Arima or Qiagen)

Slide from Marcela Uliano-Silva

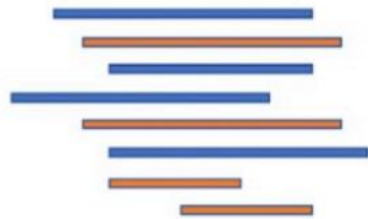


*For mitochondria genome assembly*



### Assembly

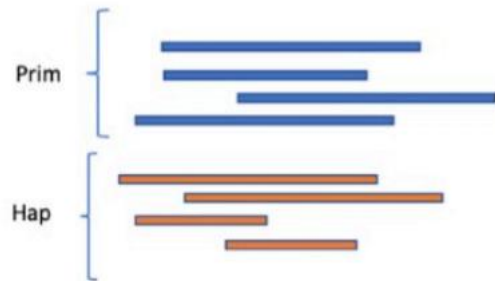
Hicanu  
or Hifiasm



2 - asmstats,  
BUSCO, merquy

### Haplotype separation

Purge dups



3 - asmstats,  
BUSCO, merquy

### Scaffolding

Yahs scaffolding  
(Arima or Qiagen  
HiC)



4 - asmstats,  
BUSCO,  
merquy, HiC  
heatmap

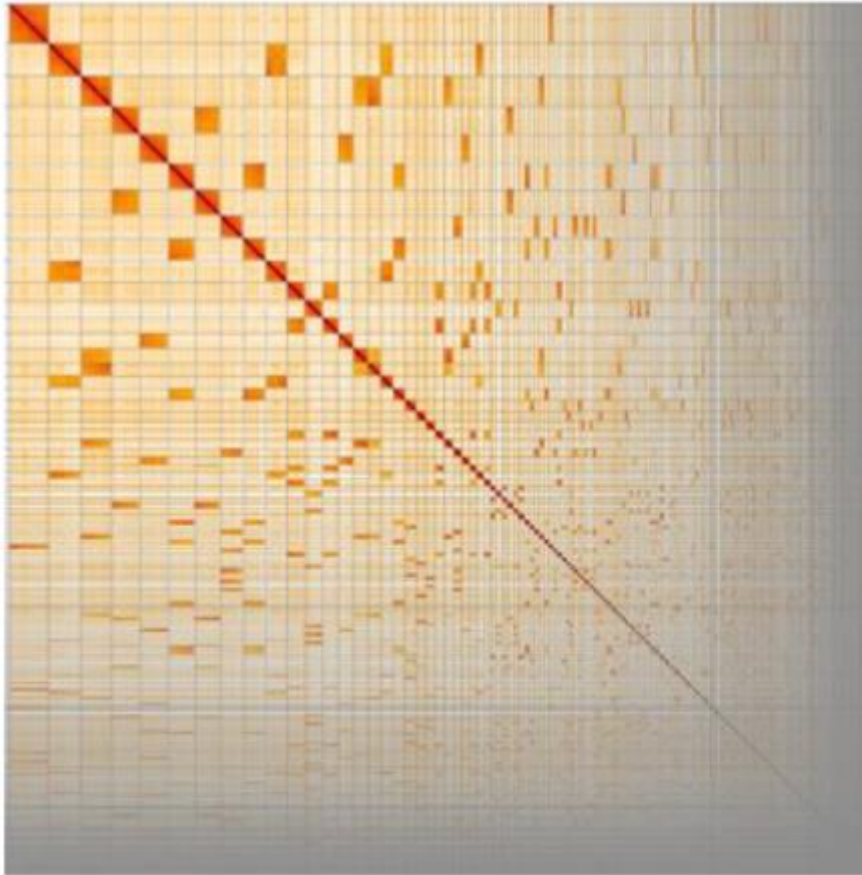
*Curated assembly*

5 - asmstats,  
BUSCO,  
merquy, HiC  
heatmap

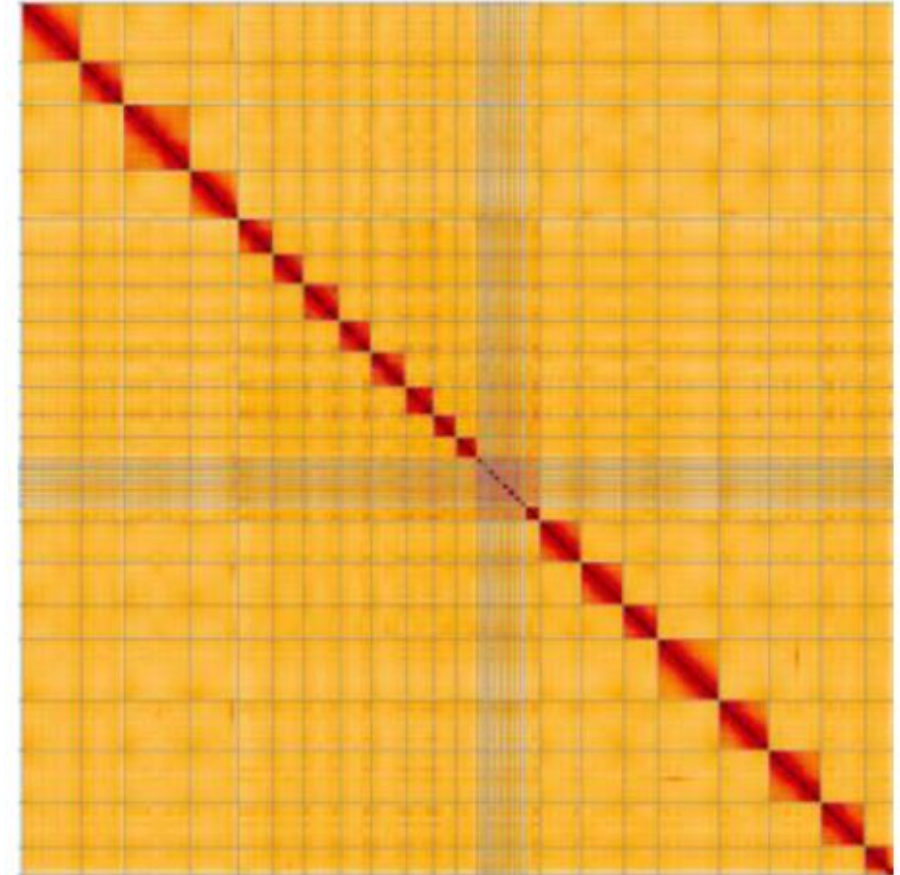
1- Kmer  
Jellyfish/  
GenomeScope,  
asmstats,  
smudgeplot

# Manual curation to correct errors by assemblers

HiC map before manual curation



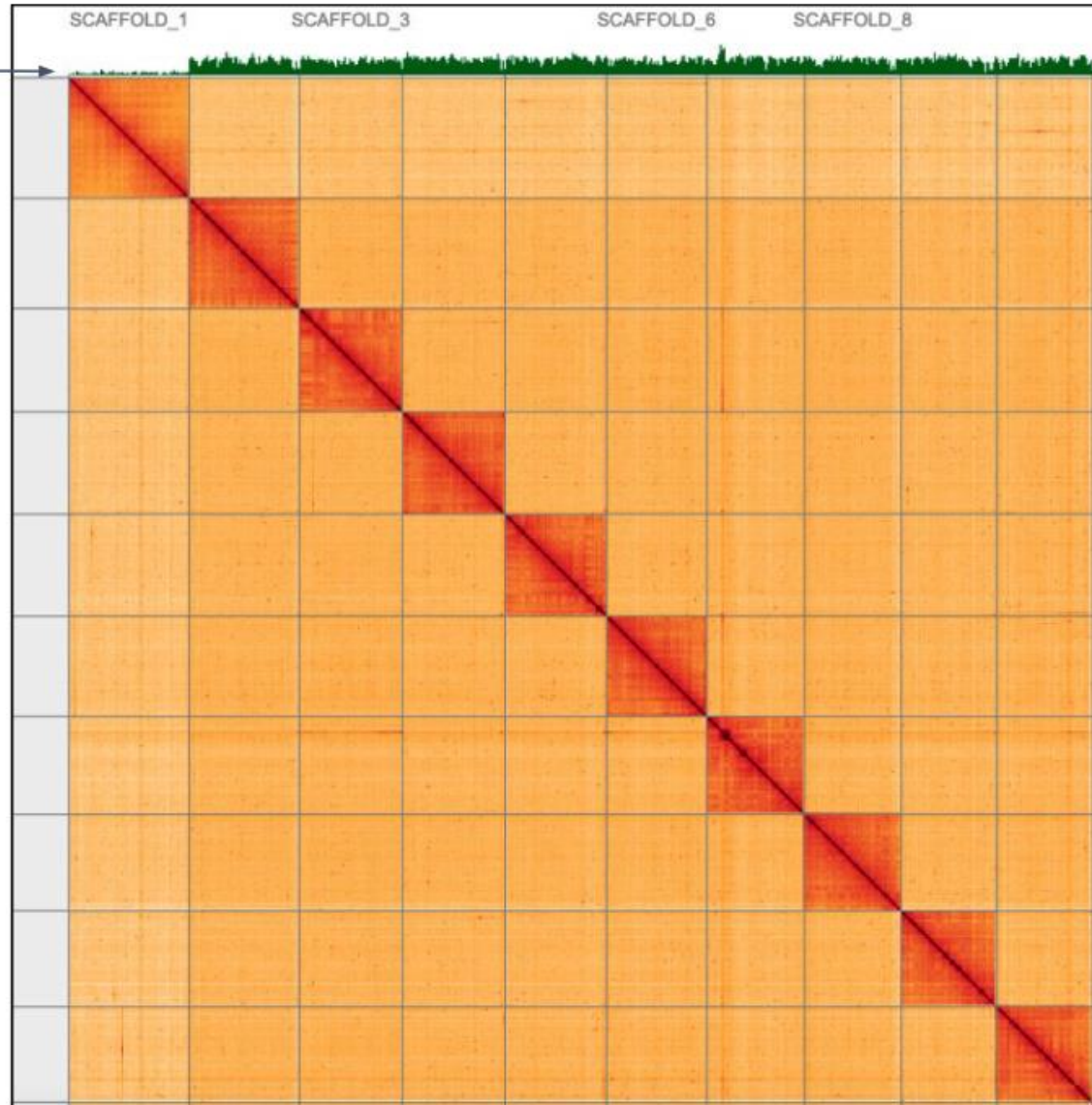
After manual curation with PretextView



[https://gitlab.com/wtsi-grit/rapid-curation/-/blob/main/Interpreting HiC Maps guide.pdf](https://gitlab.com/wtsi-grit/rapid-curation/-/blob/main/Interpreting%20HiC%20Maps%20guide.pdf)

# HiC maps will also show interesting features

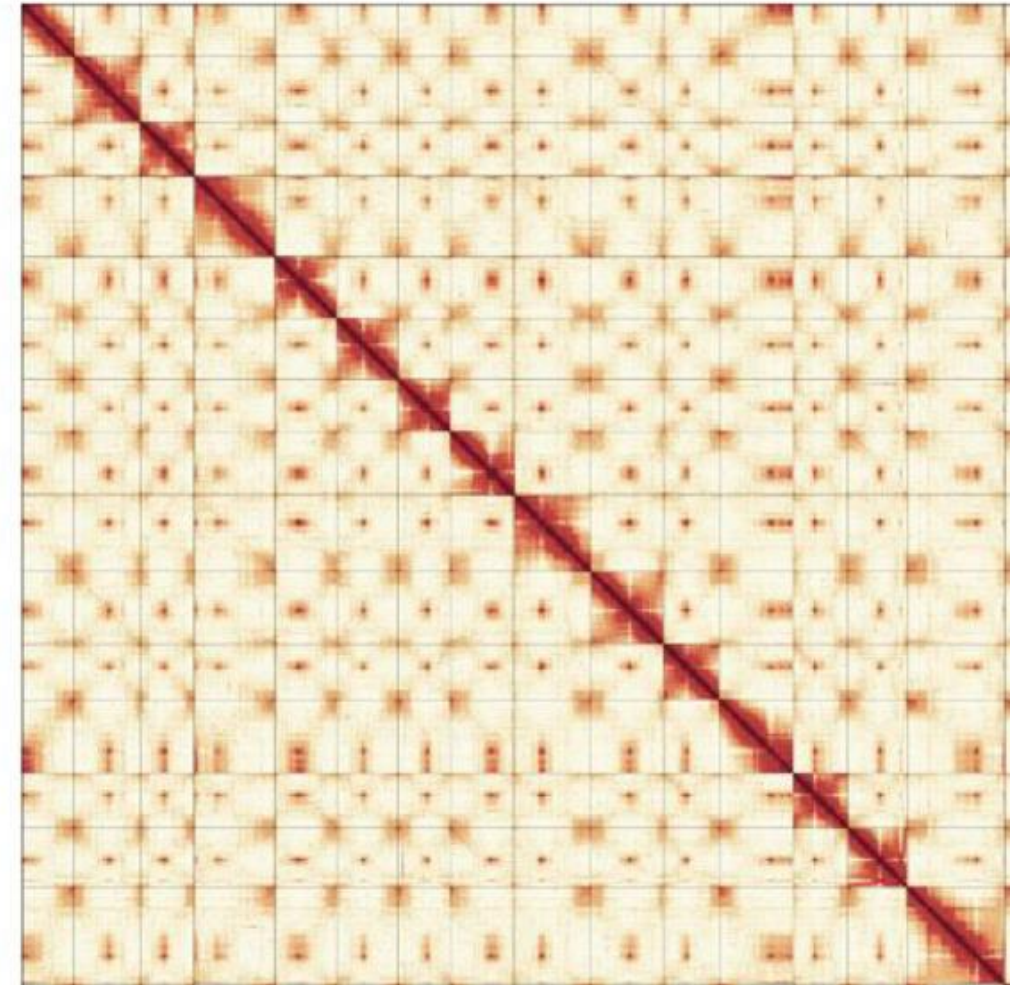
Identifying  
sex  
Chromosomes  
(half coverage)



Centromeres, eg

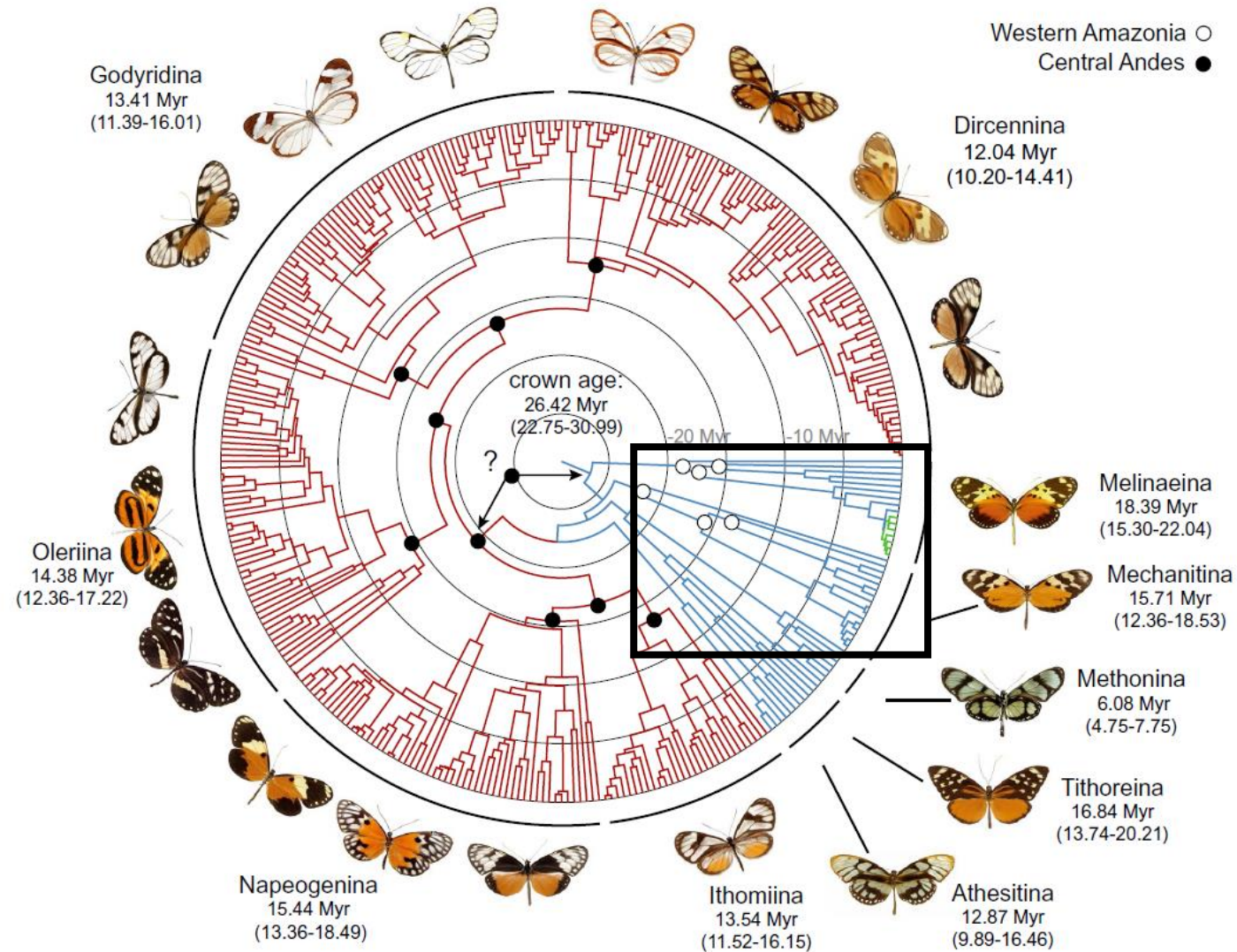


Telomeres, eg

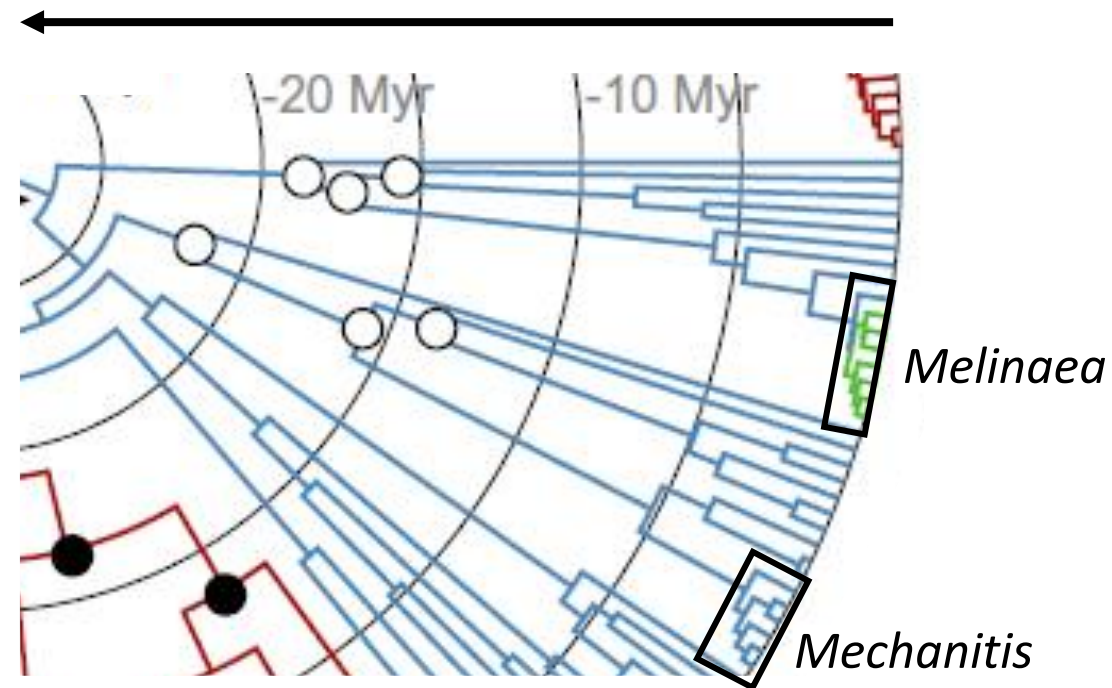


**An example of using reference  
genomes from my own work**

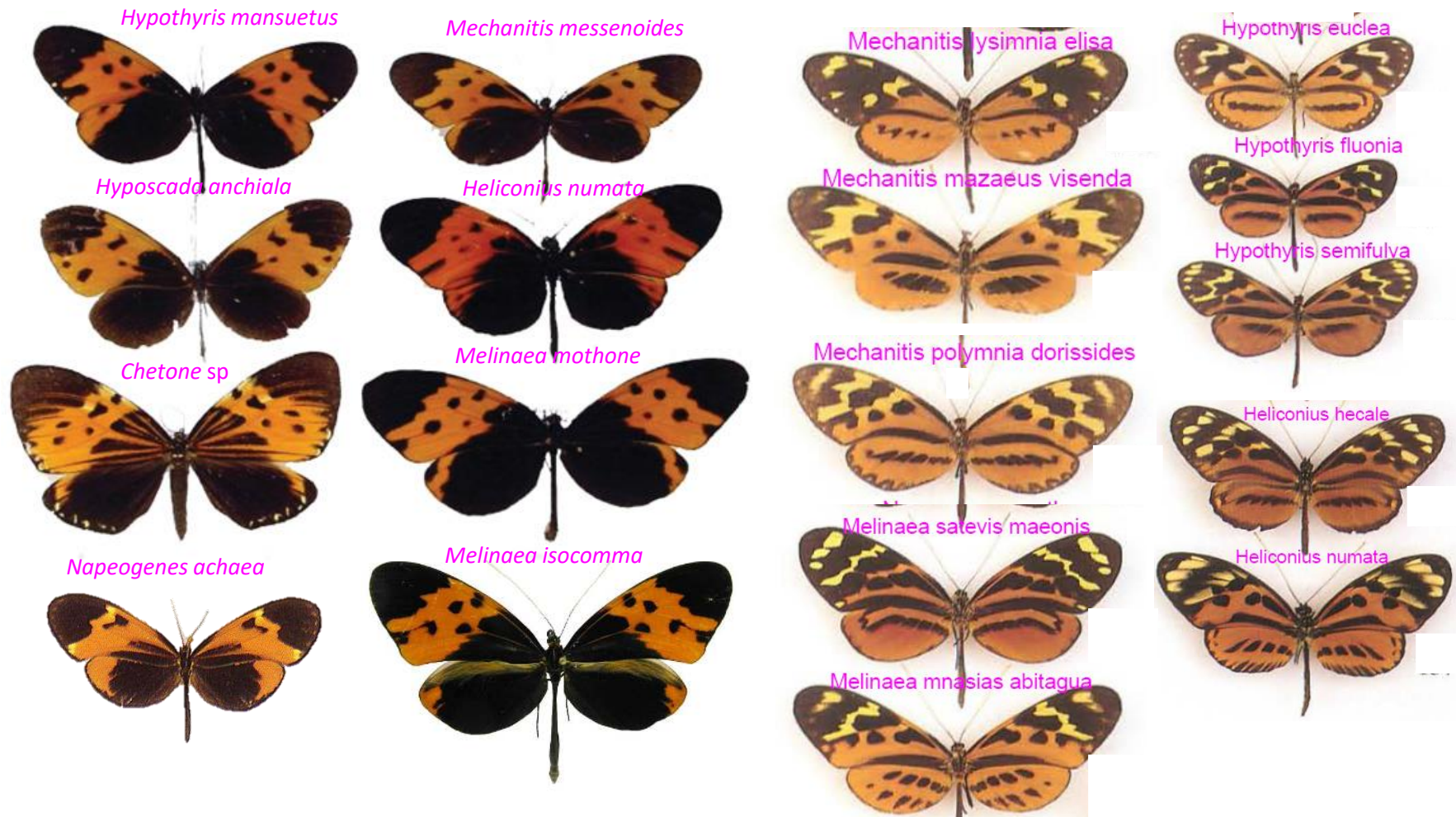
# Comparing rapidly and slowly speciating Ithomiini genera



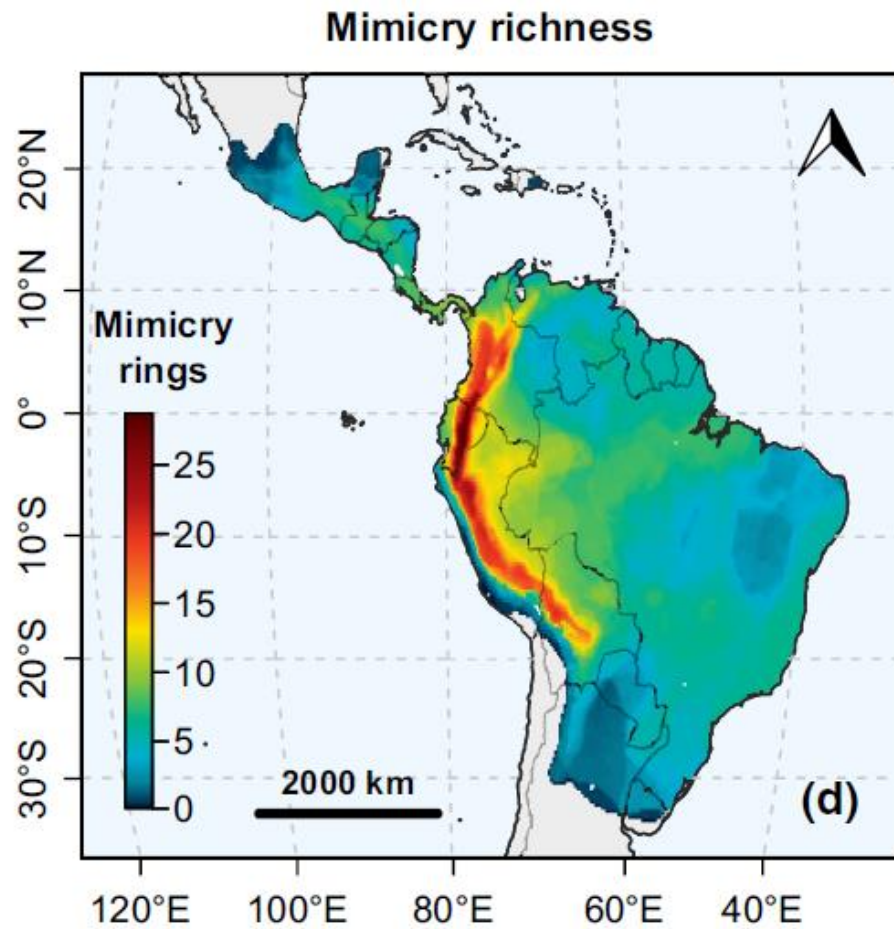
Time since last common ancestor



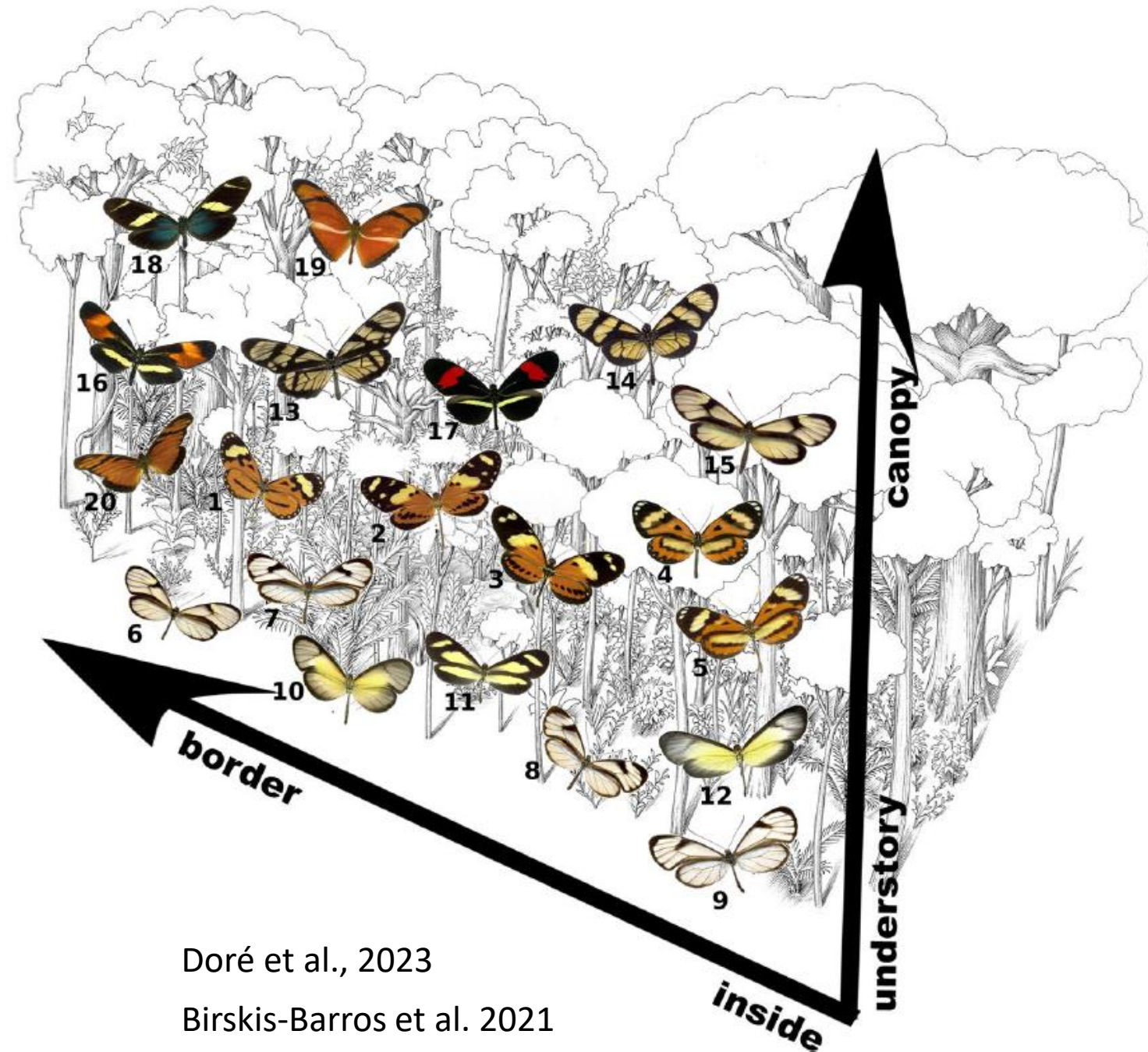
# Müllerian mimicry rings of ithomiini and others



# Co-mimics converge in microhabitat use and associated traits



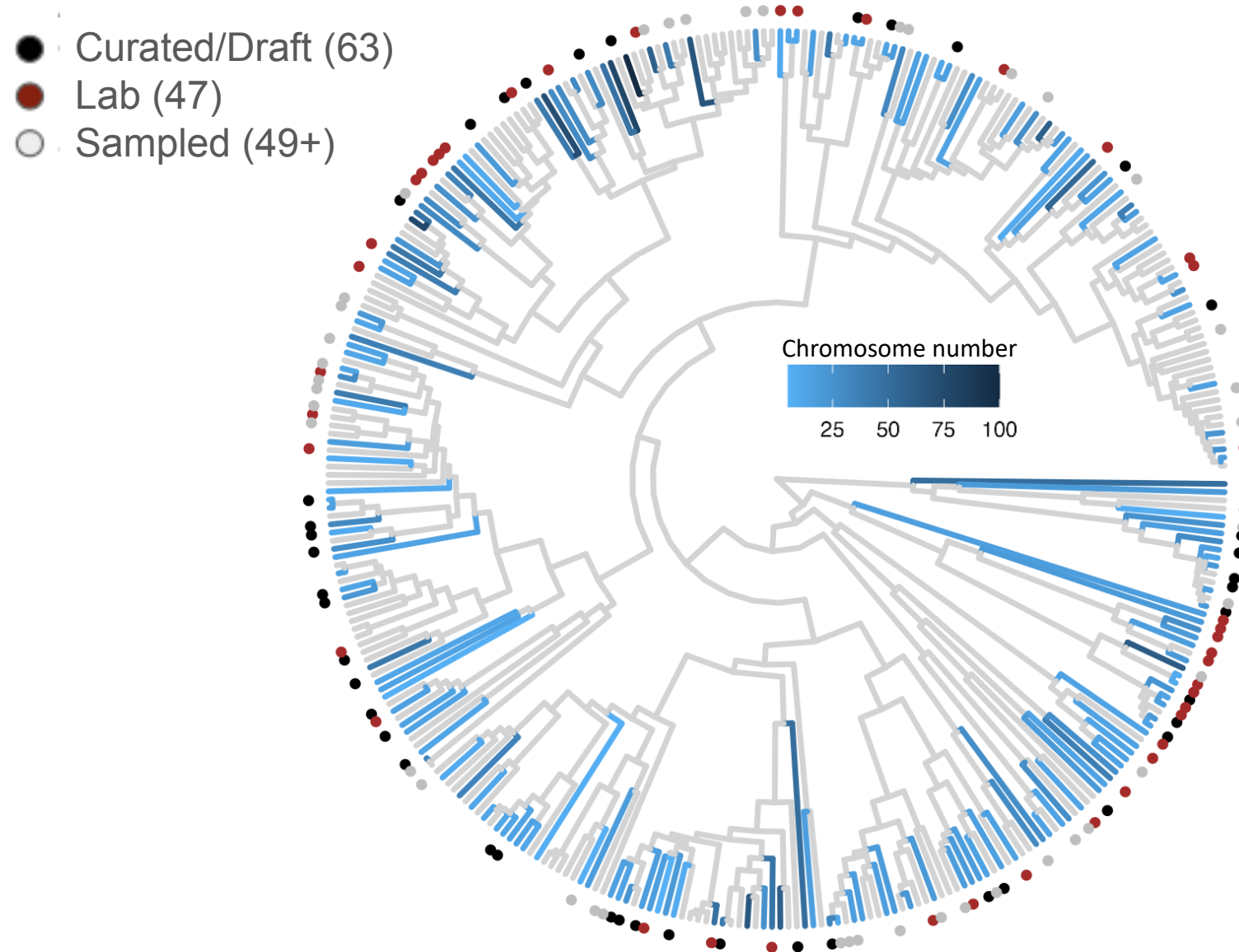
Doré et al., 2021



Doré et al., 2023

Birskis-Barros et al. 2021

# Sequencing 200 ithomiini genomes



**Patricio  
Salazar**



**Karin  
Näsval**

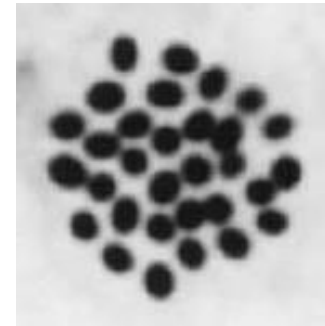
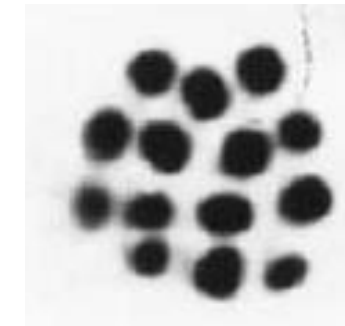
# Do chromosomal rearrangements contribute to rapid diversification?

most butterflies have 30-31 chromosomes

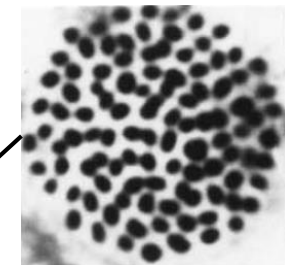
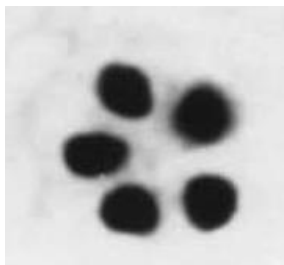
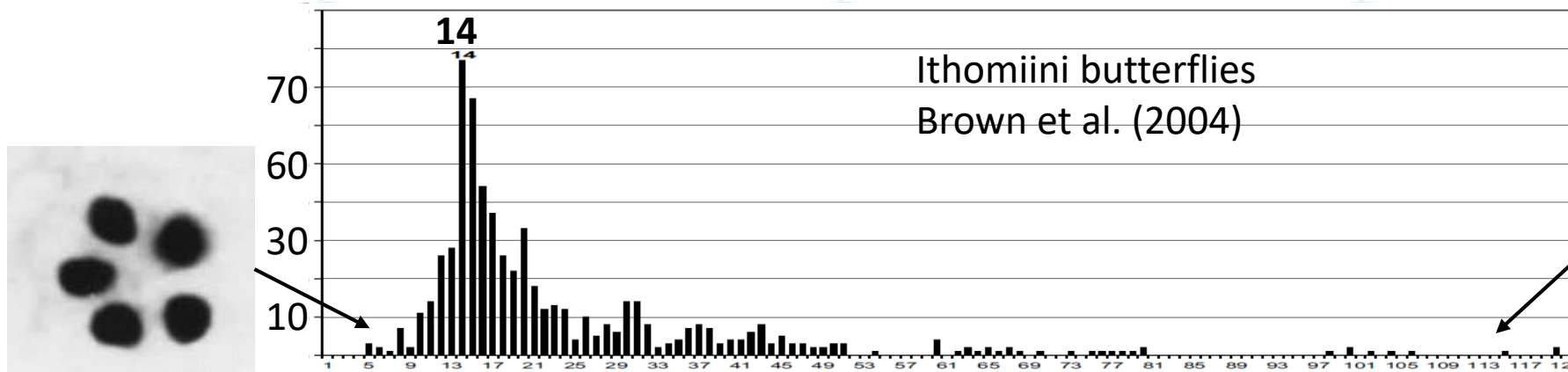
De Vos et al. (2020): 2400 Lepidoptera (including 171 ithomiini)

***Melinaea* species:**

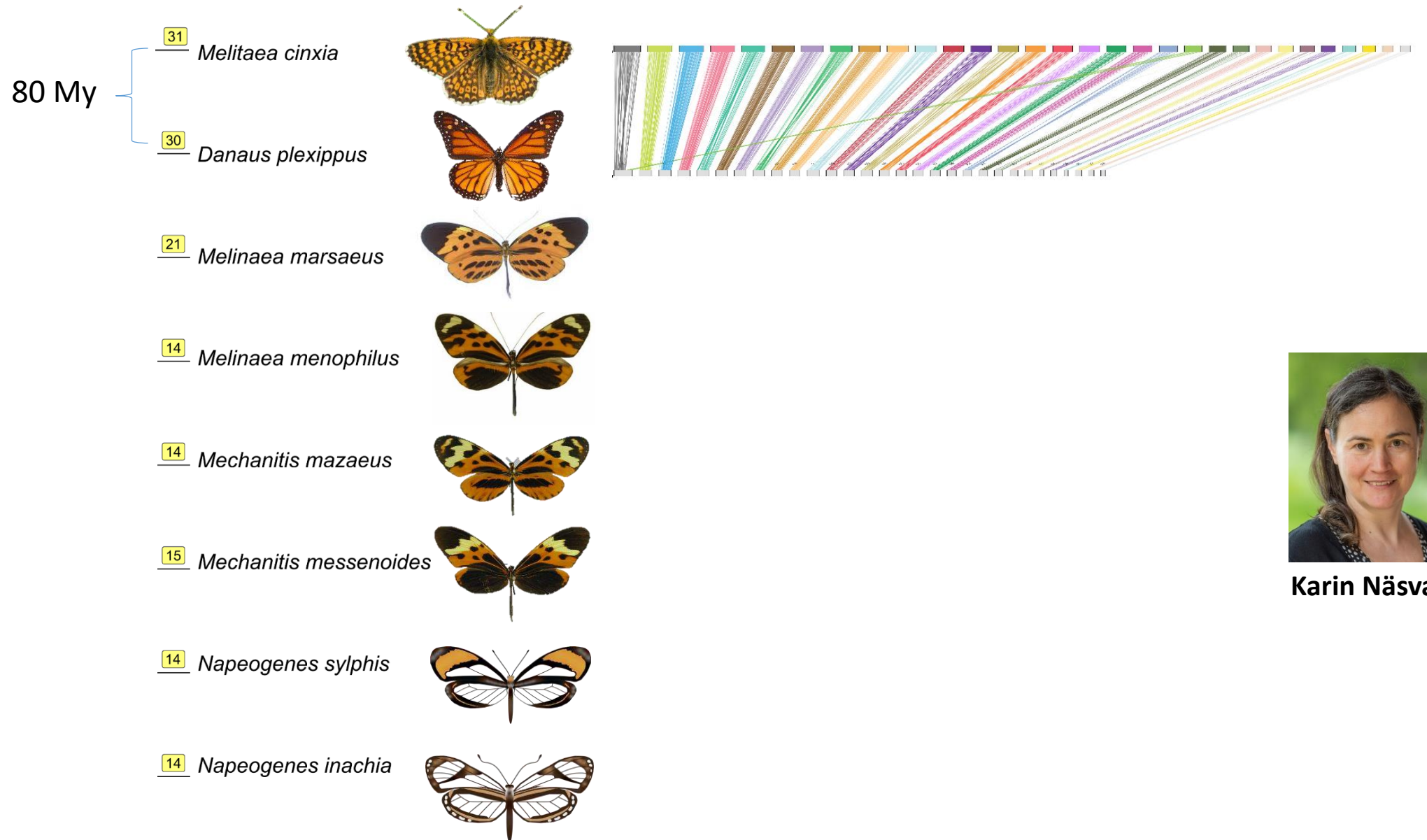
13-30 chromosomes



Keith Brown et al., 2004, Hereditas

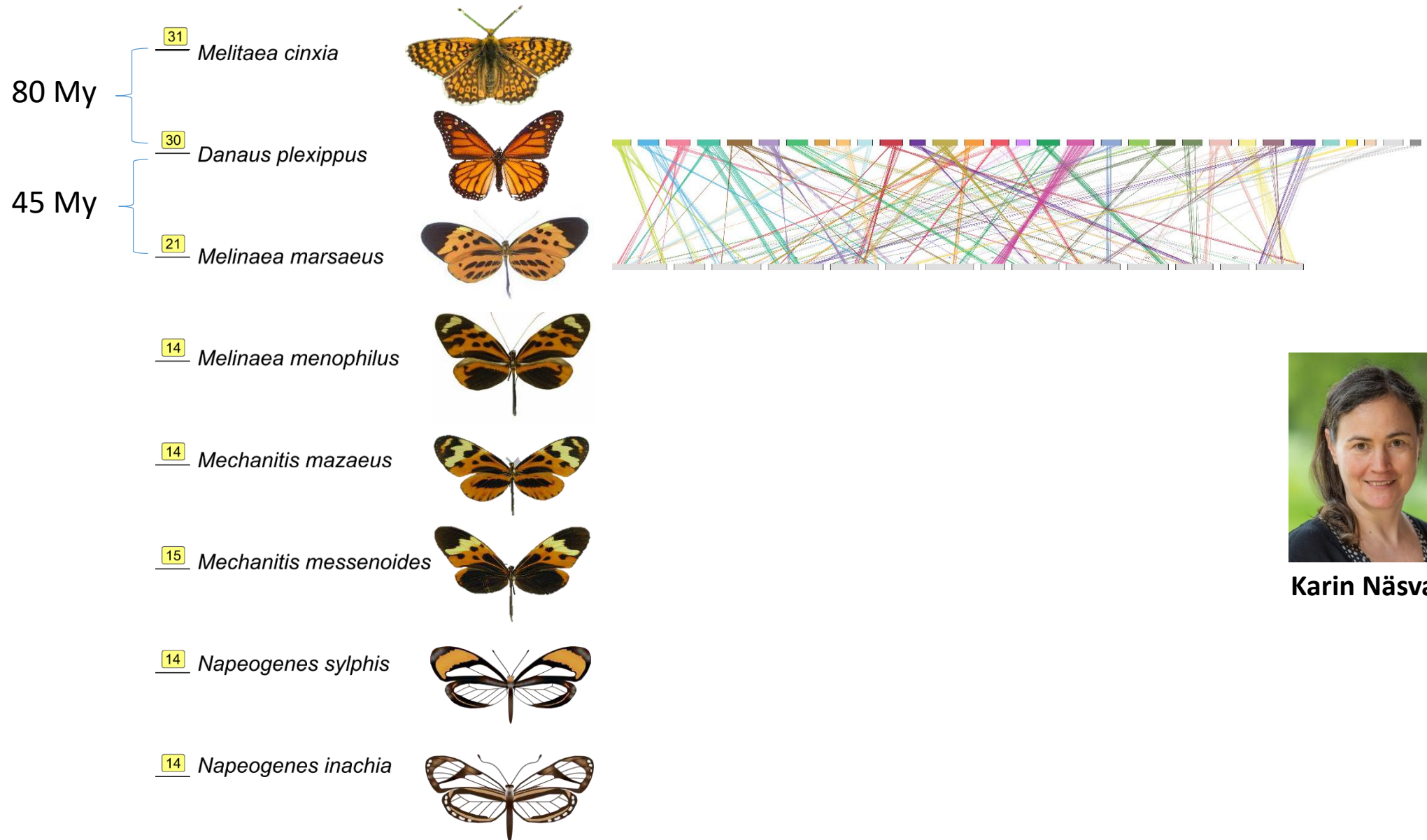


# Large-scale chromosomal rearrangements



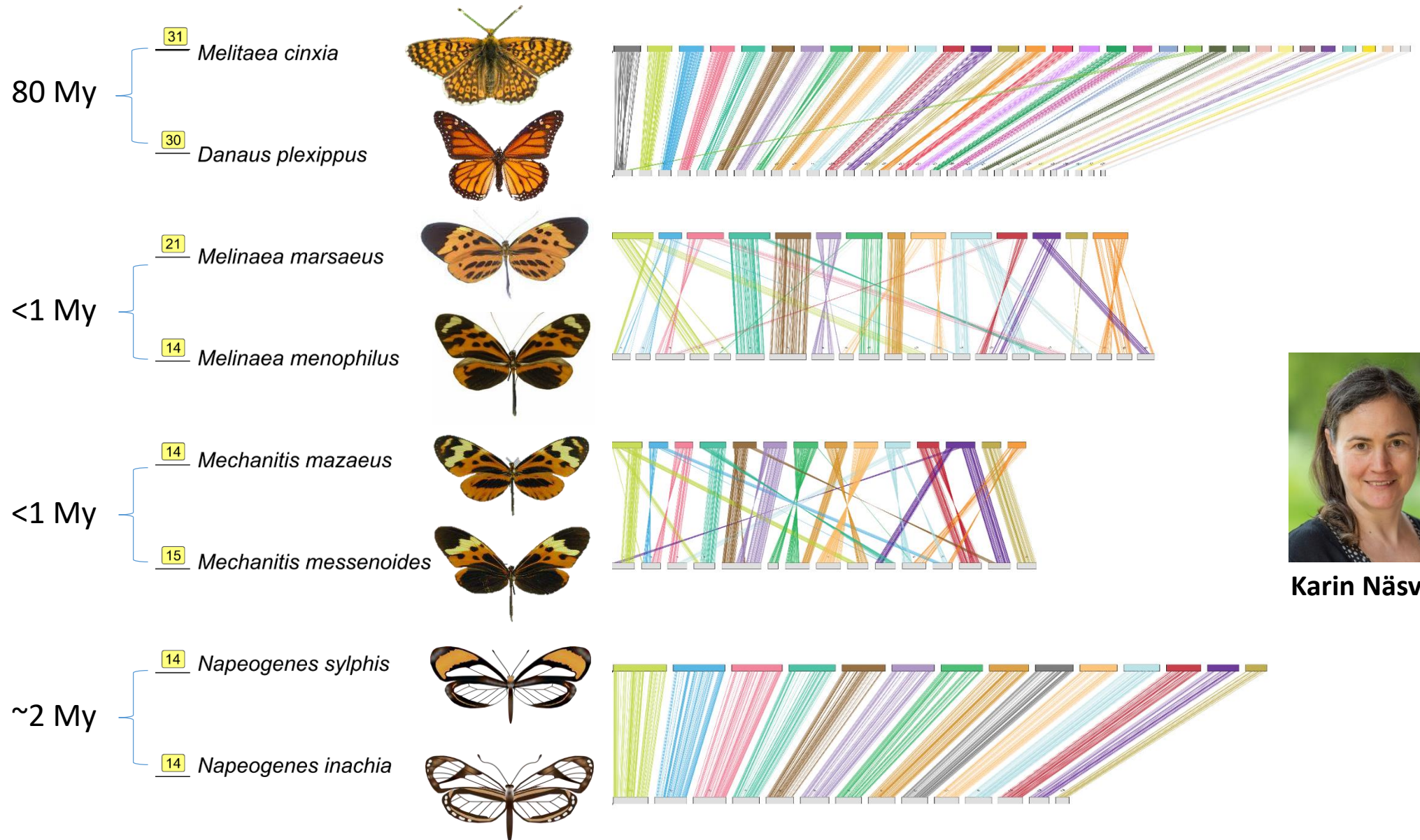
Karin Näsvali

# Large-scale chromosomal rearrangements



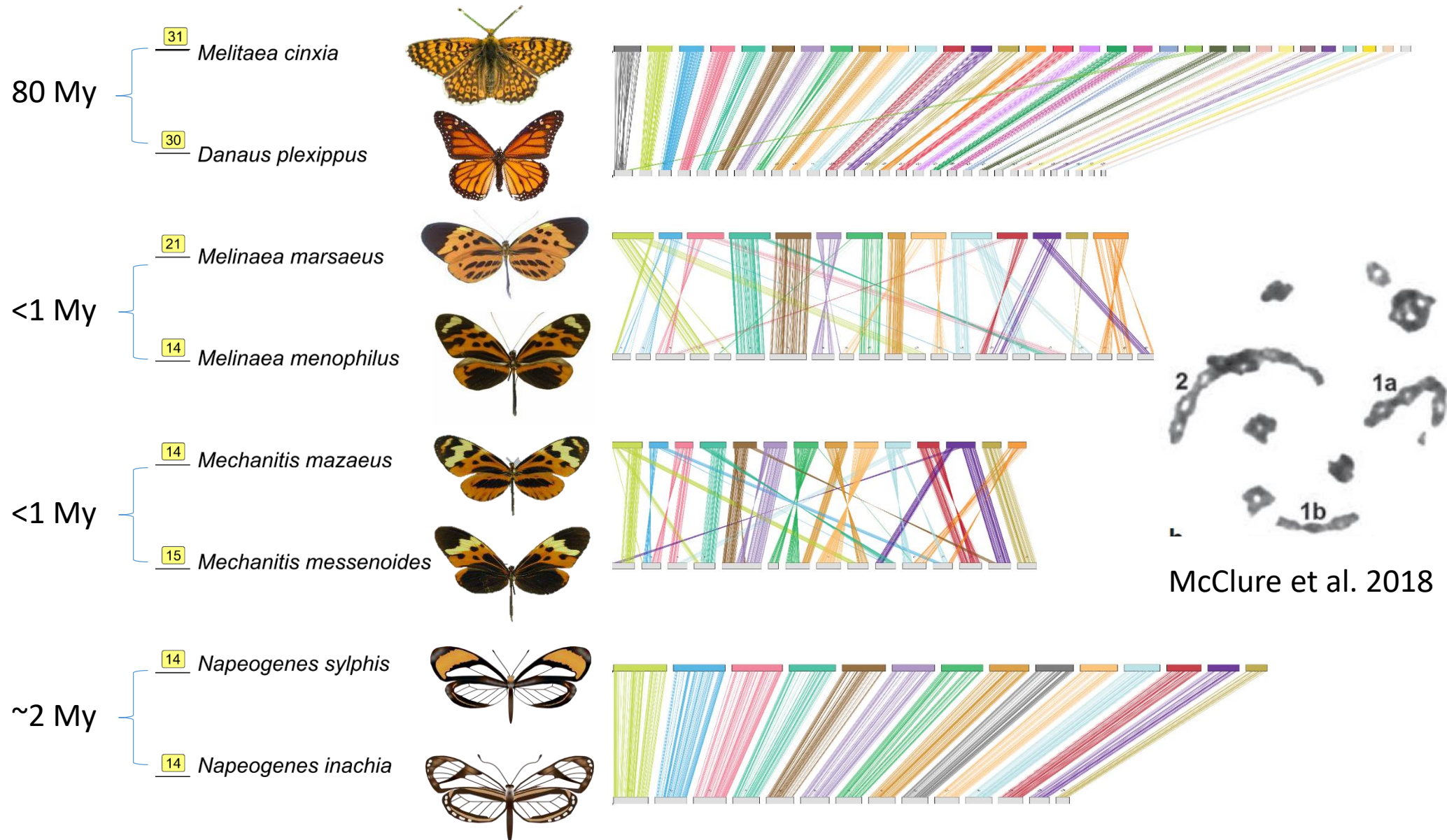
Karin Näsvali

# Large-scale chromosomal rearrangements

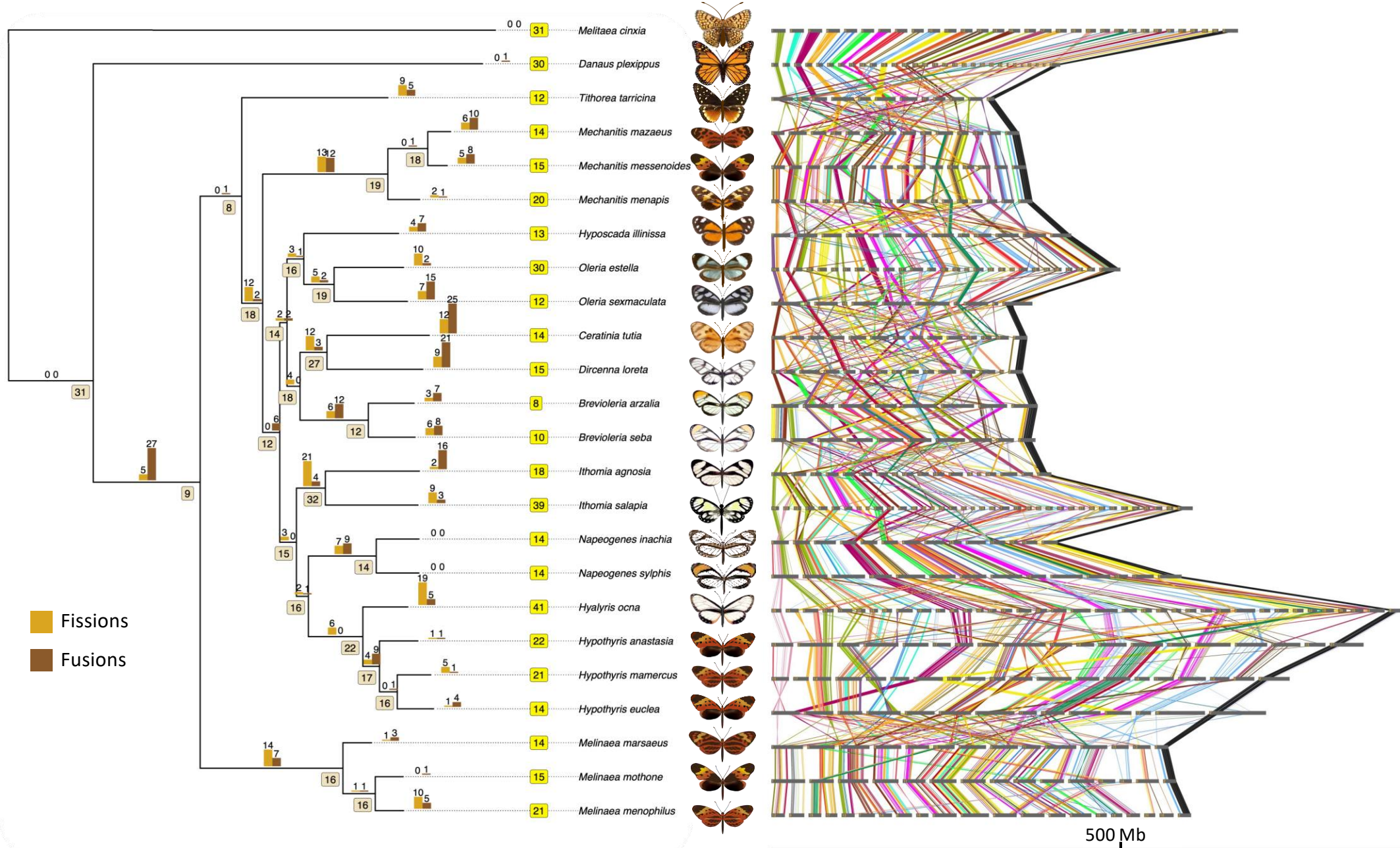


Karin Näsvali

# Large-scale chromosomal rearrangements



# Large-scale chromosomal rearrangements



Karin Näsvall

# Species of recent radiations (<1 Mya) show highly rearranged chromosomes

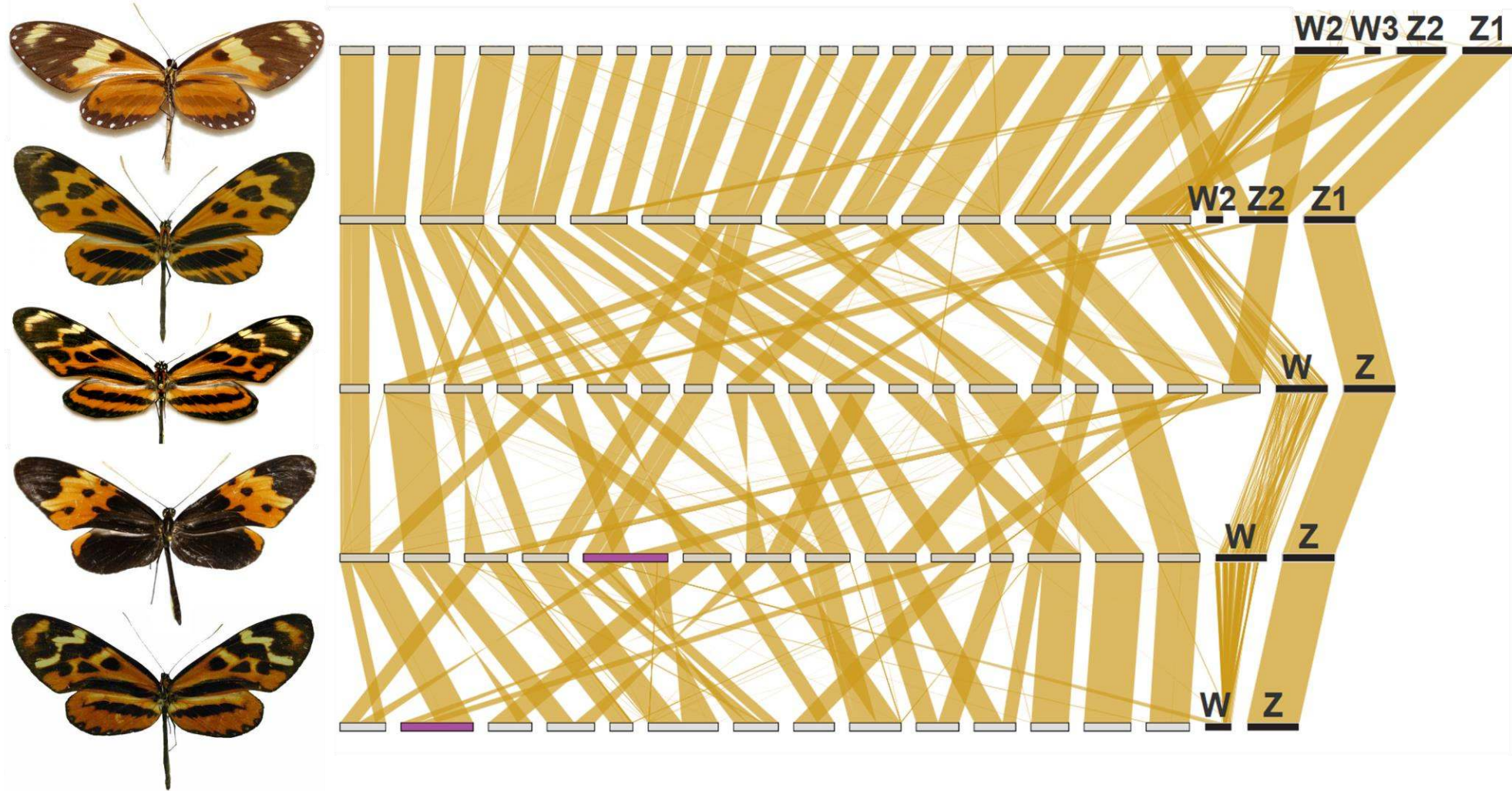
*M. macrinus* (23)  
↗  
n chromosomes

*M. mazaesus* (14)

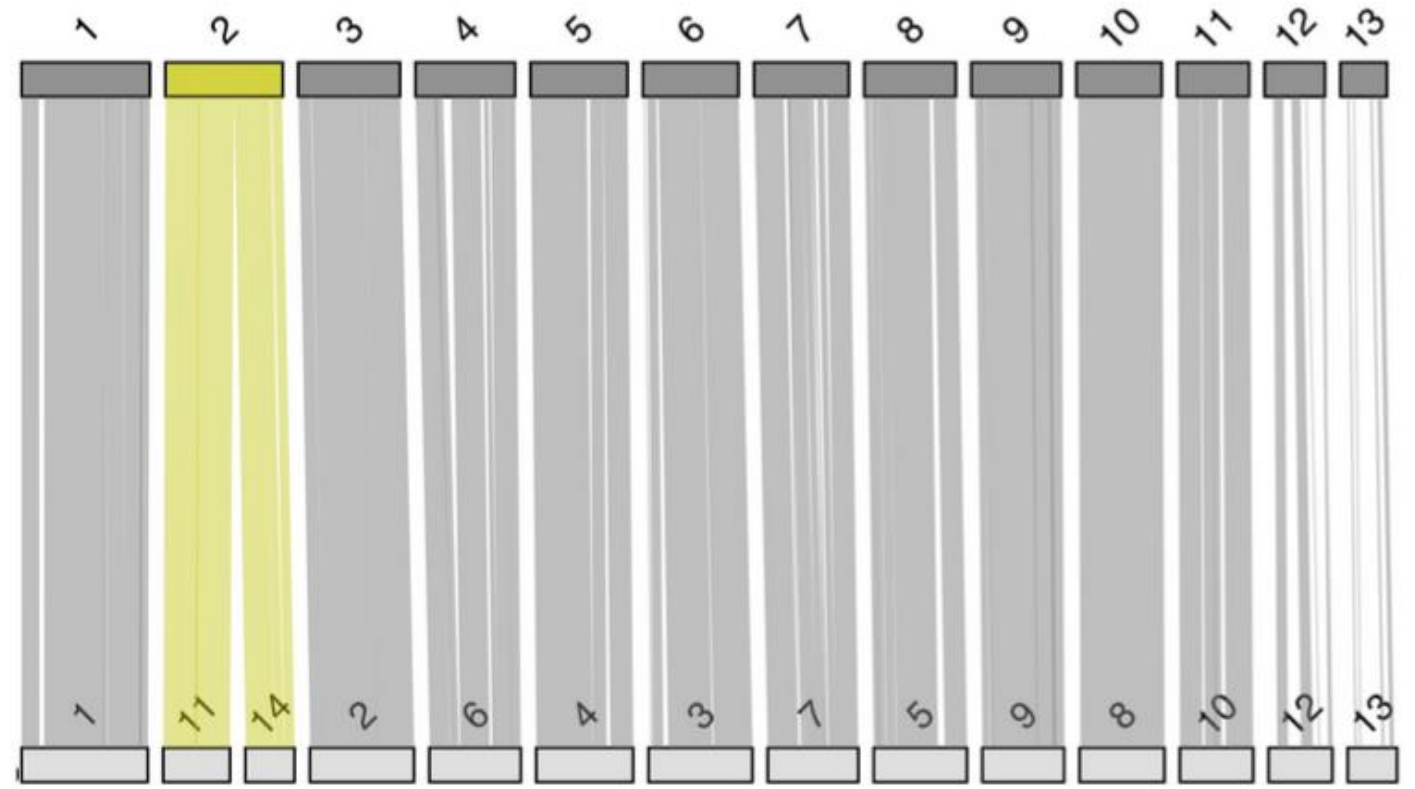
*M. menapis* (20)

*M. messenoides* (15)

*M. polymnia* (15)

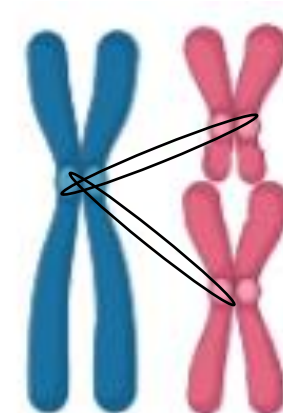
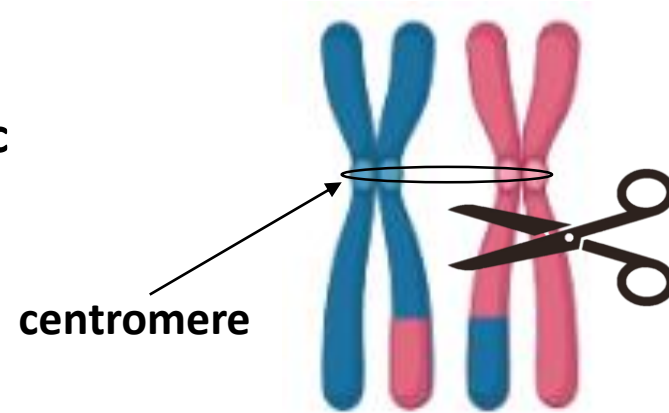


# Some individuals are heterozygous for fission-fusion rearrangements

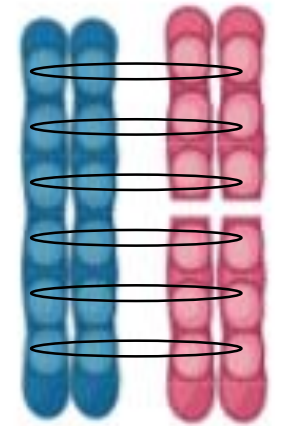
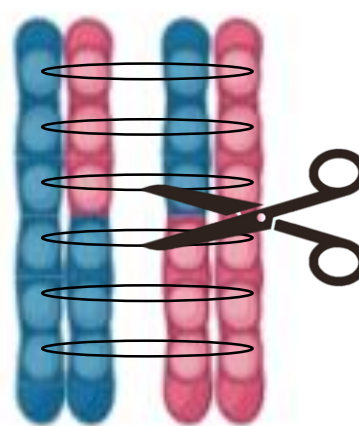


# Butterflies have holocentric chromosomes

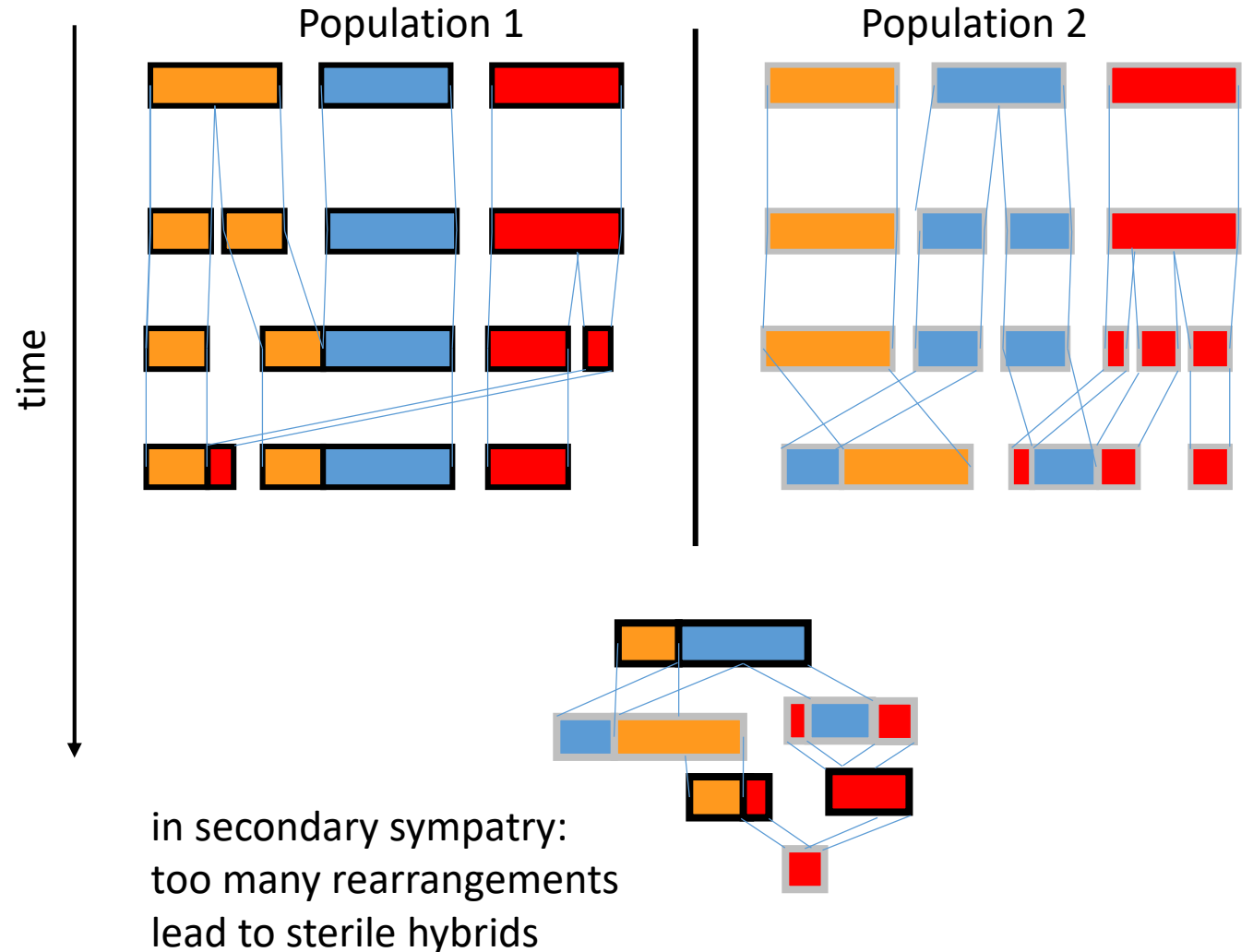
**Monocentric**  
(like us)



**Holocentric**  
(all butterflies)



# Hypothesis: Chromosomal rearrangements during periods of geographic isolation drive speciation







# Many thanks to my wonderful team and key collaborator

@Sanger



Arif Maulana  
PhD Student



Dr Karin Näsvall  
Postdoctoral Fellow



Dr Nicol Rueda  
Postdoctoral Fellow



Dr Patricio A.  
Salazar-Carrión  
Postdoctoral Fellow



Jonah Walker  
PhD Student



Eva van der  
Heijden  
PhD student

@IKIAM  
en Ecuador



Franz Chandi



Kimberly  
Gavilanes



Alex Arias



María José  
Sánchez



Prof Caroline Bacquet