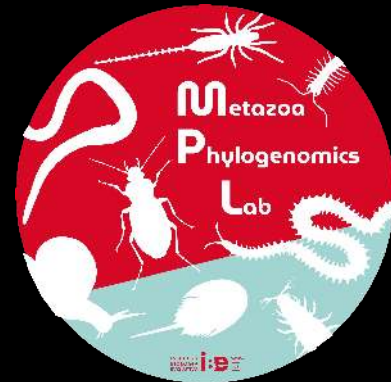


# INTRODUCTION TO PHYLOGENOMICS

Rosa Fernández  
Institute of Evolutionary Biology (CSIC-UPF)

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[www.metazomics.com](http://www.metazomics.com)



# Content of the lecture

1

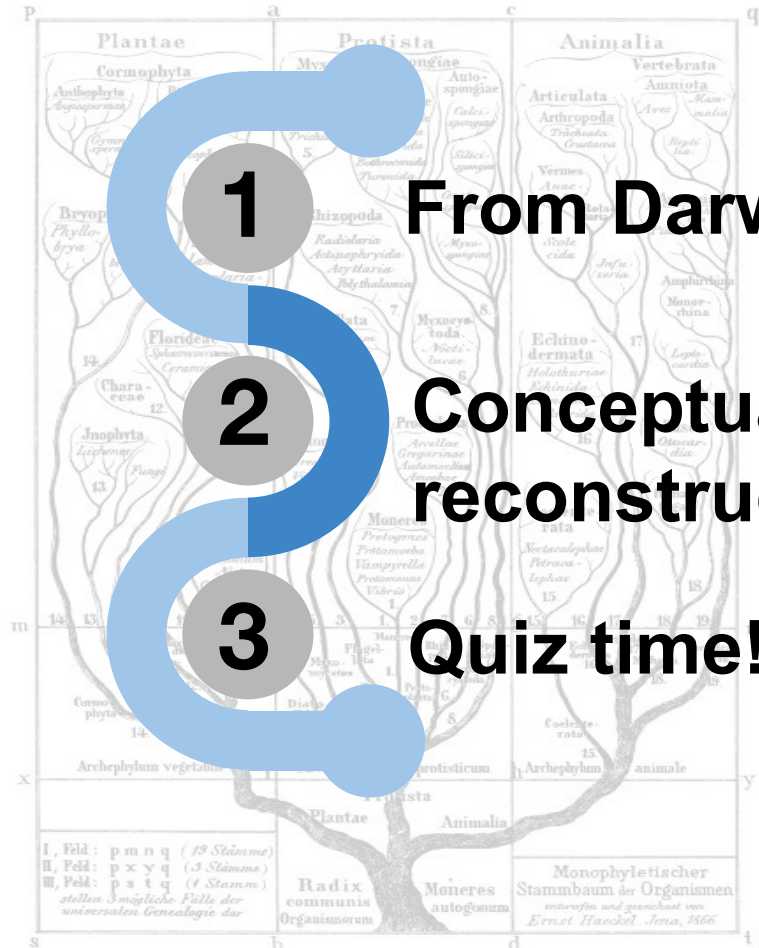
From Darwin to phylogenomics

2

Conceptual framework for phylogenomic reconstruction

3

Quiz time!



# Content of the lecture

1

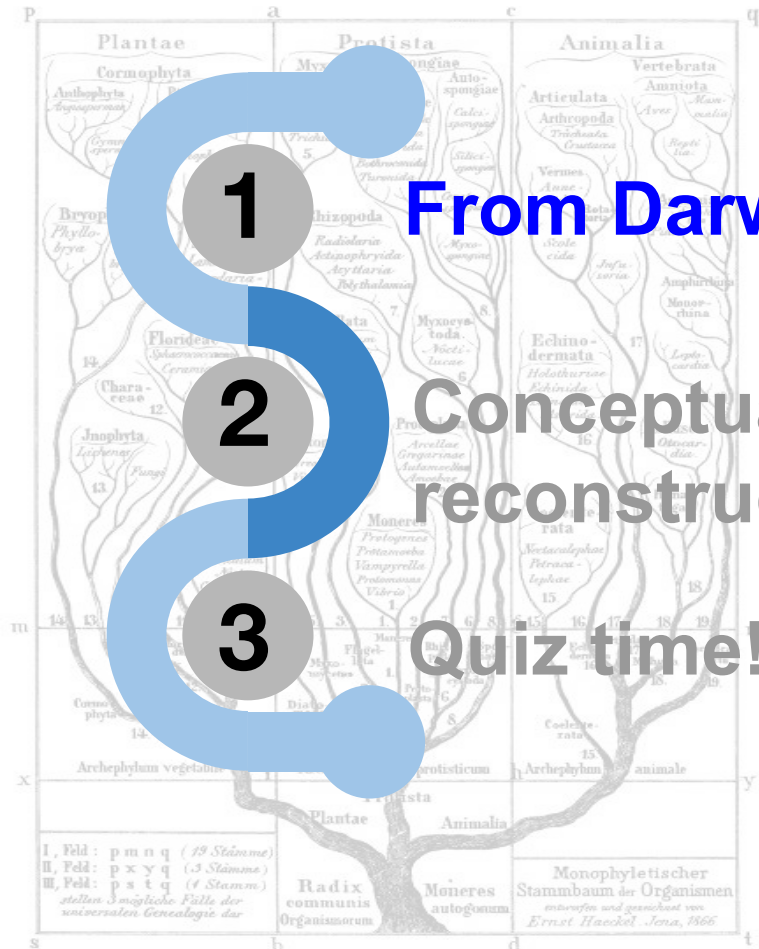
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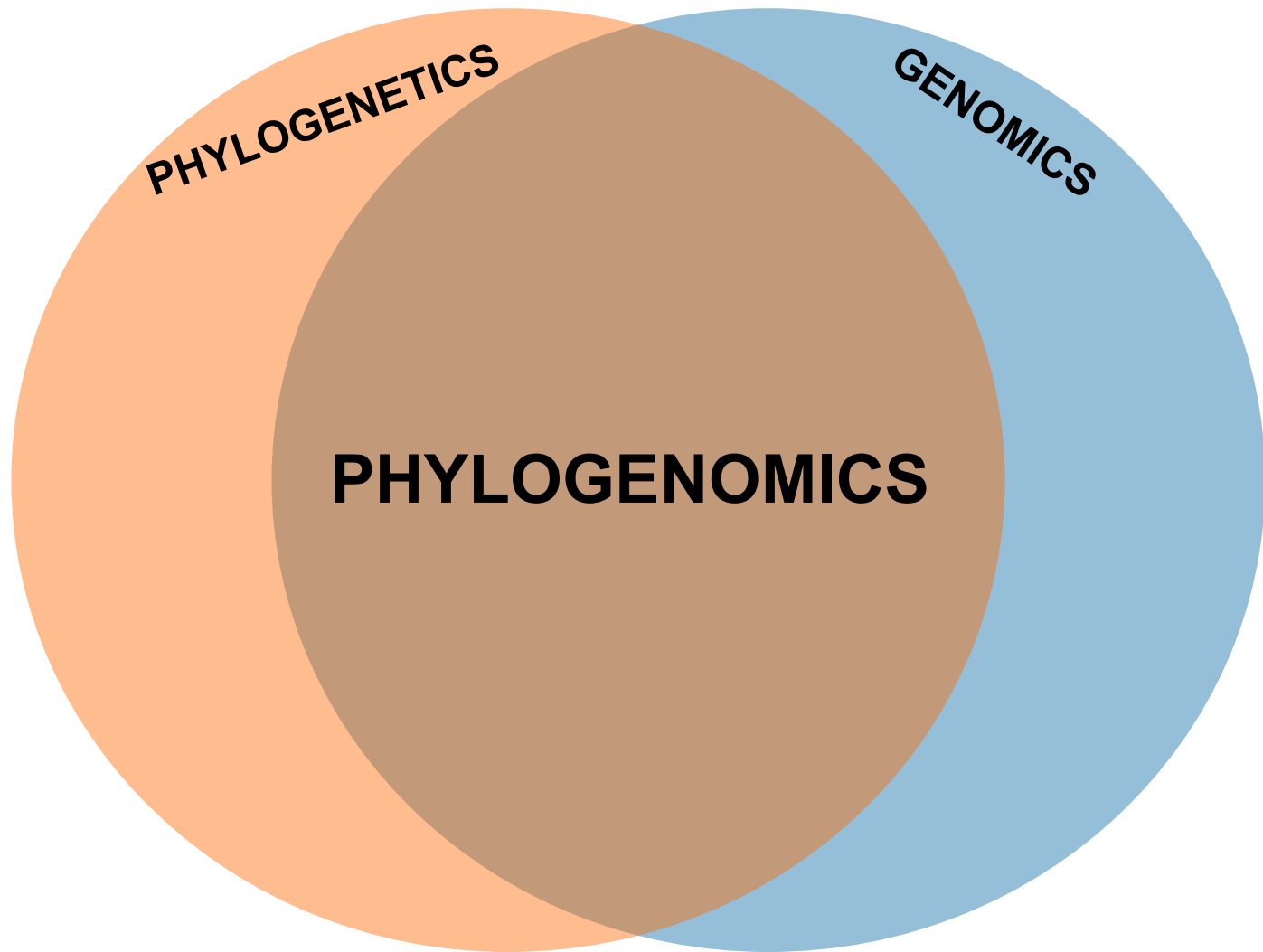
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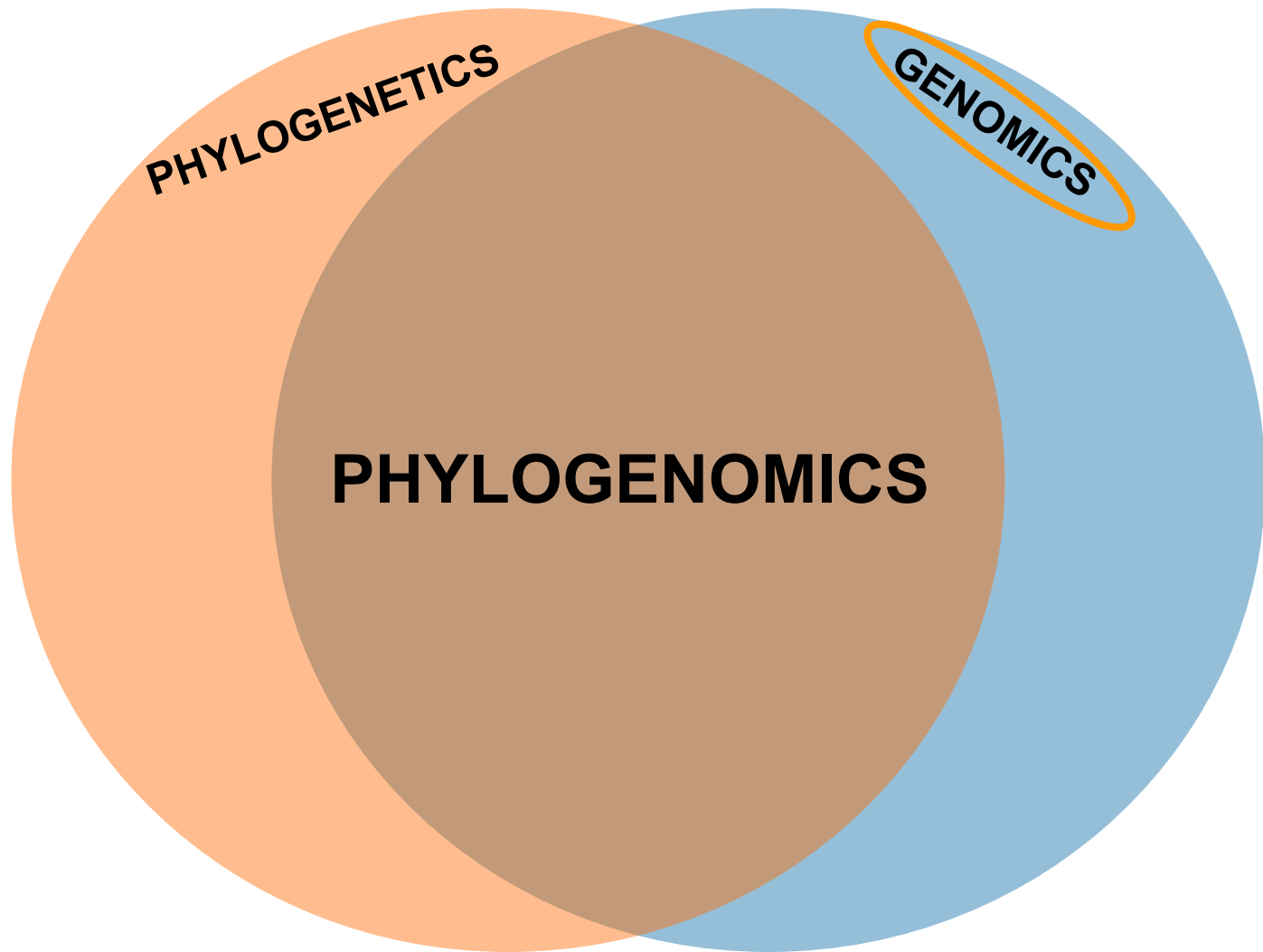
Conceptual framework for phylogenomic reconstruction

3

Quiz time!









## Genomics

---

- The study of an organism's complete set of genetic information.
- The genome includes both genes (coding) and non-coding DNA.
- 'Genome': the complete genetic information of an organism.

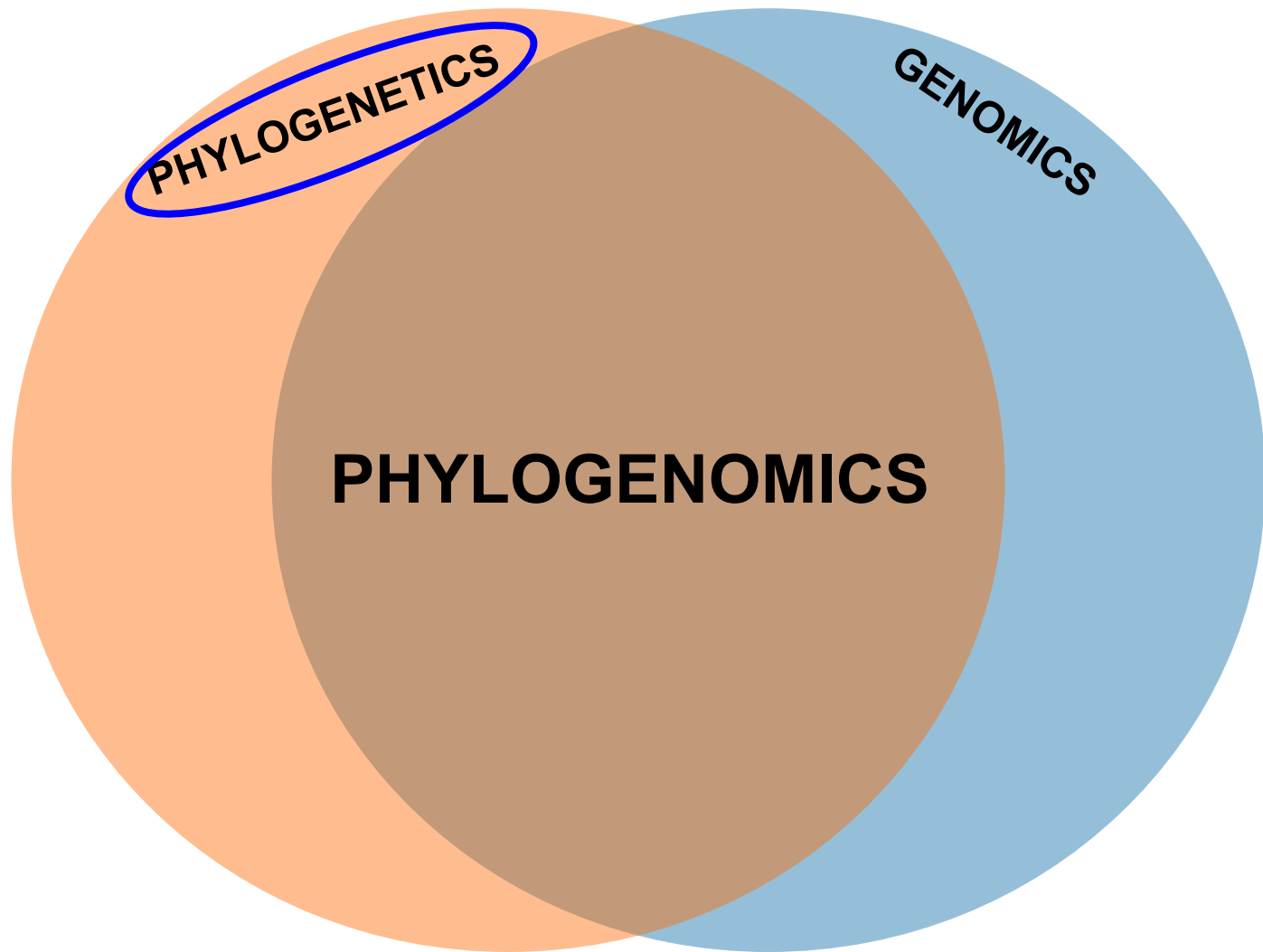
VS



## Genetics

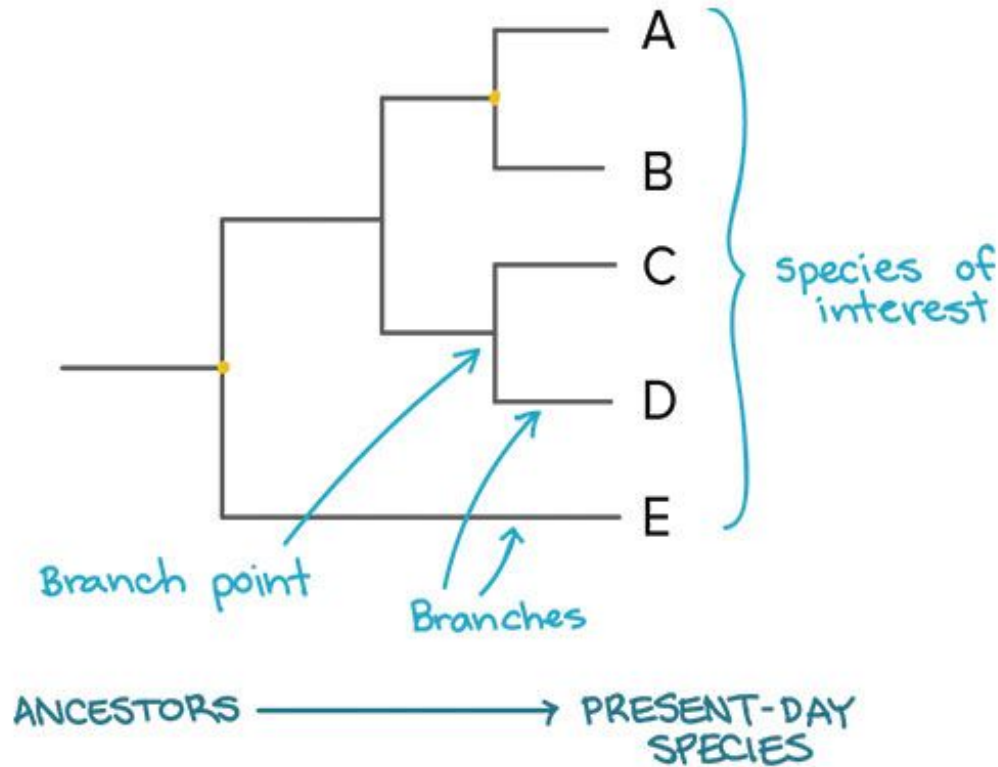
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- The study of heredity
- The study of the function and composition of single genes.
- 'Gene': specific sequence of DNA that codes for a functional molecule.



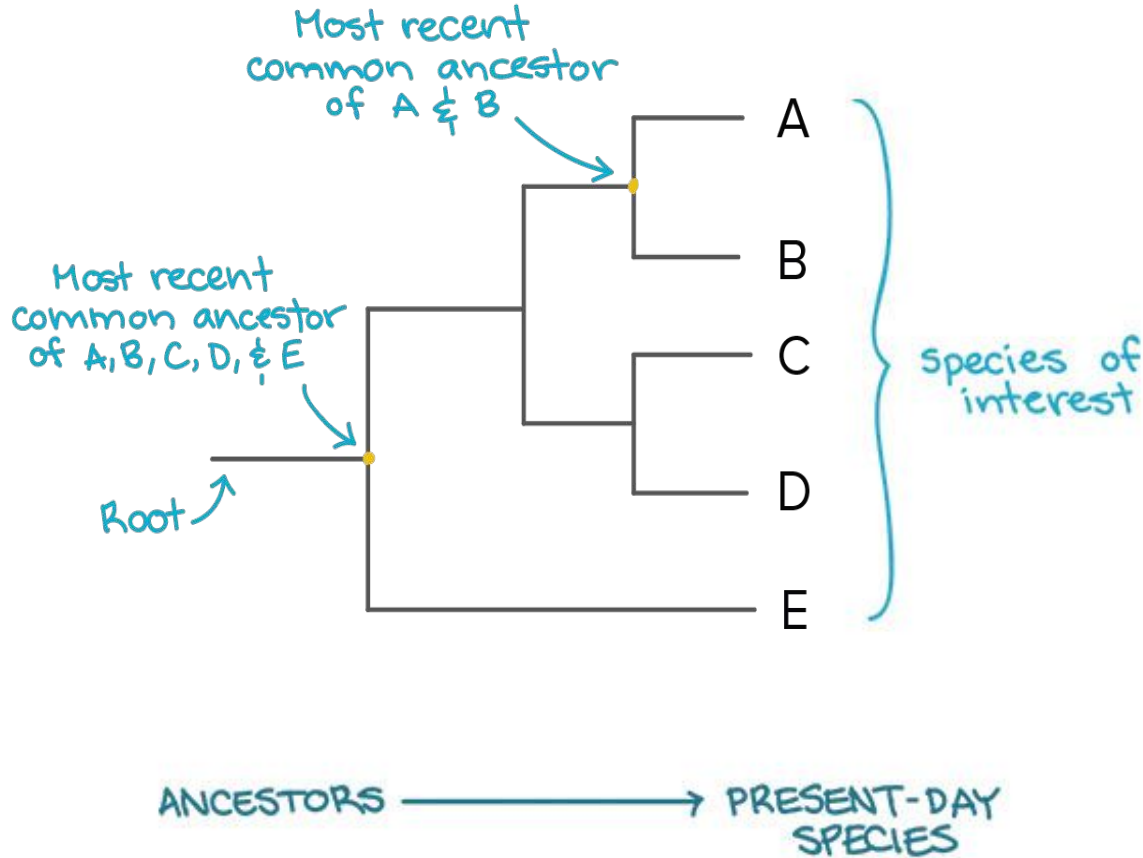
**What is a phylogeny...?**

# What is a phylogeny...?



A **phylogenetic tree** is a hypothesis of how species or genes are related through evolution

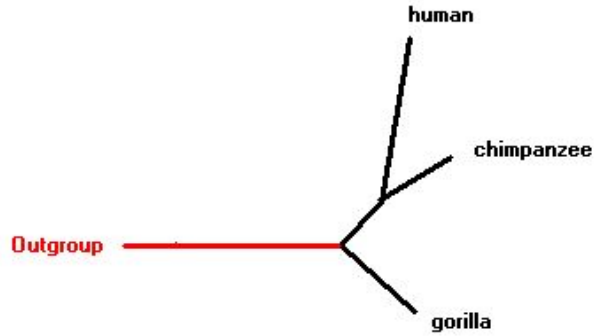
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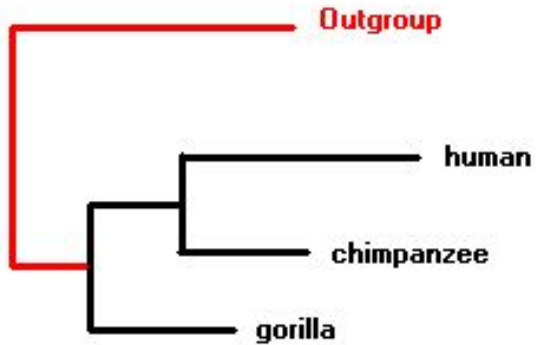
A **phylogenetic tree** is a hypothesis of how species or genes are related through evolution

# What is a phylogeny...?

Unrooted tree

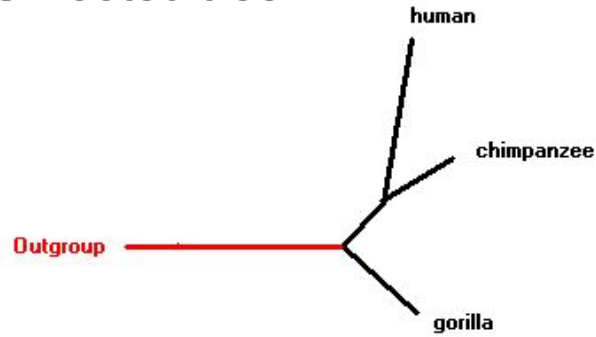


Rooted tree

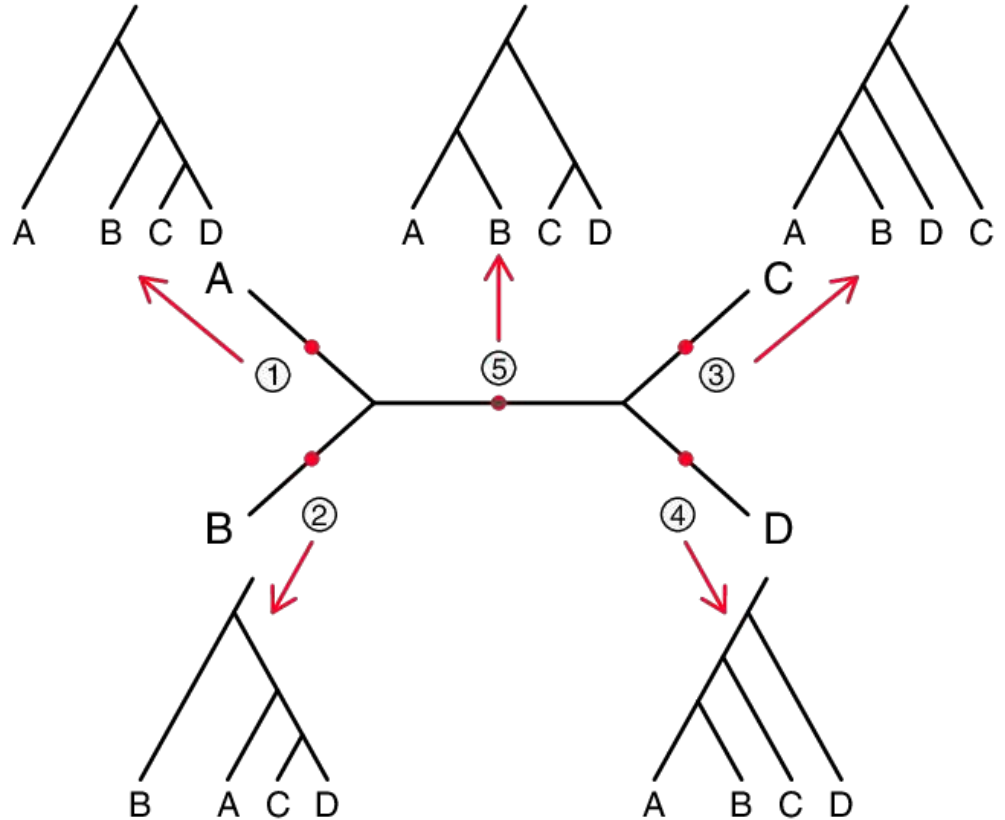
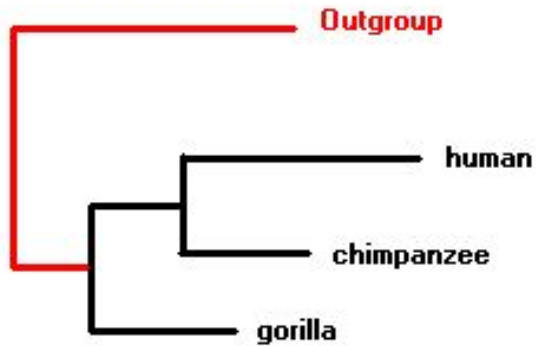


# What is a phylogeny...?

Unrooted tree



Rooted tree



**What is a phylogeny, why is it important...?**

**What is a phylogeny, why is it important...?**

**Which came first, the chicken or the egg?**



Birds  
**(Chickens)**

What is a phylogeny, why is it important...?

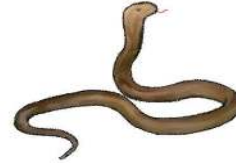
Which came first, the chicken or the egg?



Turtles



Lizards



Snakes



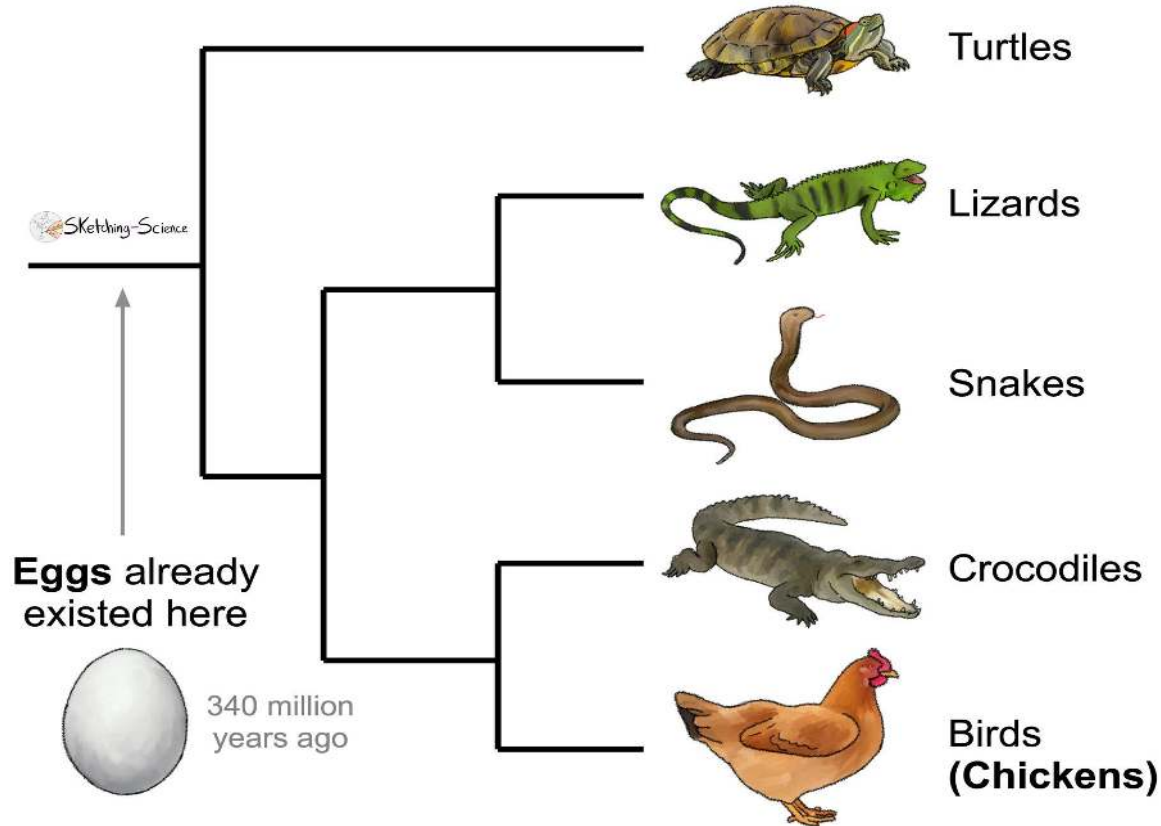
Crocodiles



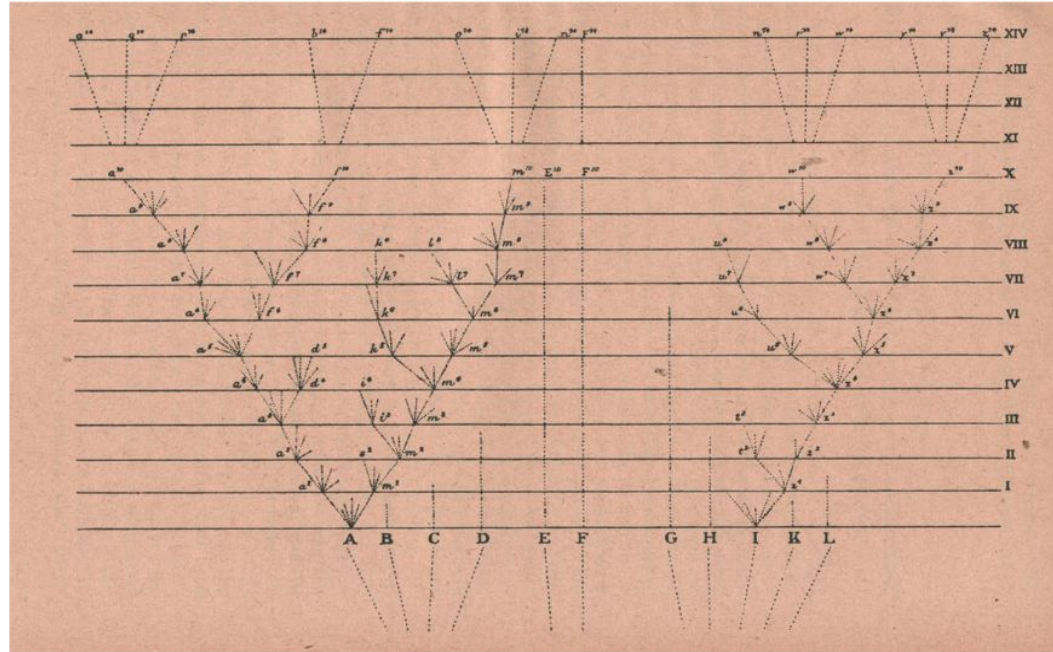
Birds  
**(Chickens)**

# What is a phylogeny, why is it important...?

## Which came first, the chicken or the egg?



# The first phylogenies



(Darwin 1859)

“As buds give rise by growth to fresh buds, and these, if vigorous, branch out and overtop on all sides many a feebler branch, so by generation I believe it has been with the great Tree of Life, which fills with its dead and broken branches the crust of the earth, and covers the surface with its ever branching and beautiful ramifications”

# The first phylogenies

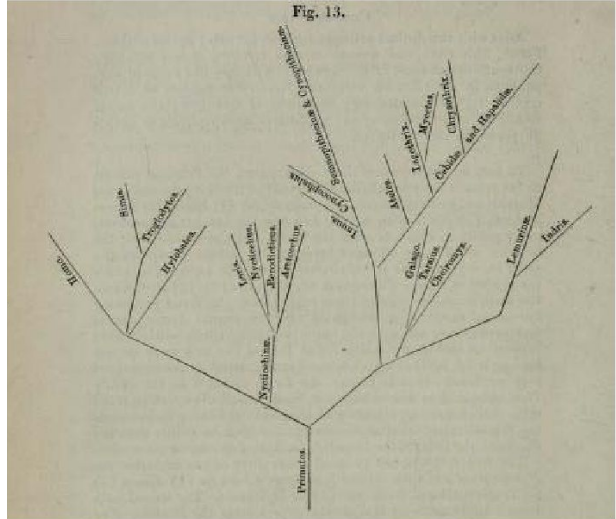
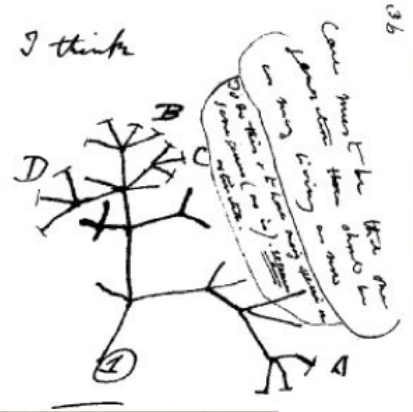
and instinct as the summing up of many contrivances, each useful to the possessor, nearly in the same way as when we look at any great mechanical invention as the summing up of the labour, the experience, the reason, and even the blunders of numerous workmen; when we thus view each organic being, how far more interesting, I speak from experience, will the study of natural history become!

A grand and almost untrodden field of inquiry will be opened, on the causes and laws of variation, on correlation of growth, on the effects of use and disuse, on the direct action of external conditions, and so forth. The study of domestic productions will rise immensely in value. A new variety raised by man will be a far more important and interesting subject for study than one more species added to the infinitude of already recorded species. Our classifications will come to be, as far as they can be so made, genealogies; and will then truly give what may be called the plan of creation. The rules for classifying will no doubt become simpler when we have a definite object in view. We possess no pedigrees or armorial bearings; and we have to discover and trace the many diverging lines of descent in our natural genealogies, by characters of any kind which have long been inherited. Rudimentary organs will speak infallibly with respect to the nature of long-lost structures. Species and groups of species, which are called aberrant, and which may fancifully be called living fossils, will aid us in forming a picture of the ancient forms of life. Embryology will reveal to us the structure, in some degree obscured, of the prototypes of each great class.

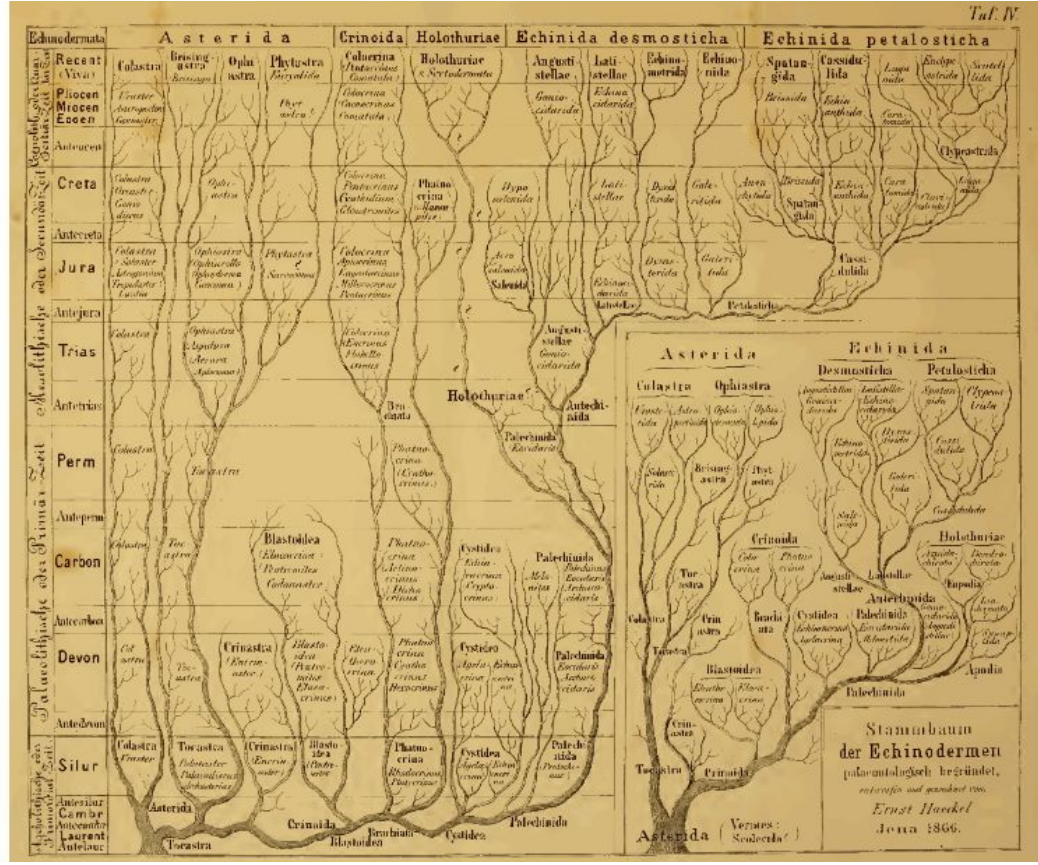
When we can feel assured that all the individuals of the same species, and all the closely allied species of most genera, have within a not very remote period de-

# The first phylogenies

The concept:  
Darwin's 'I think'  
(1837)



Mivart (1865) Proc. Zool. Soc. London

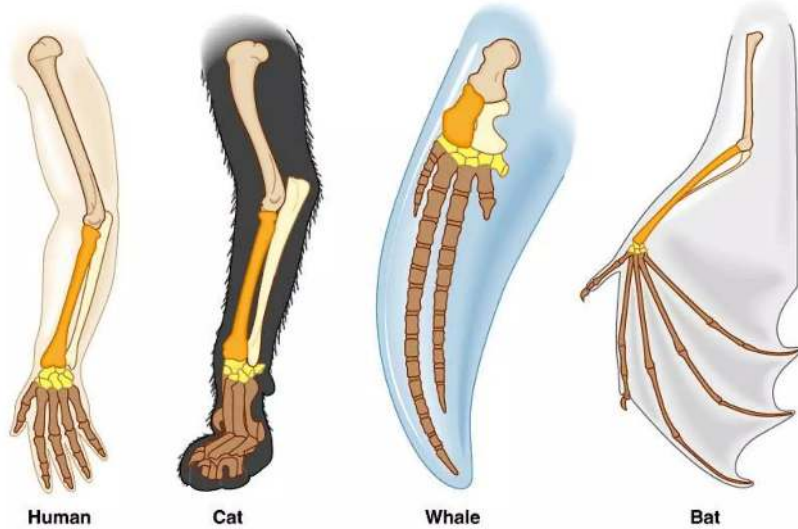


Haeckel (1866)

**What is a phylogeny, why is it important... and how do you build one?**

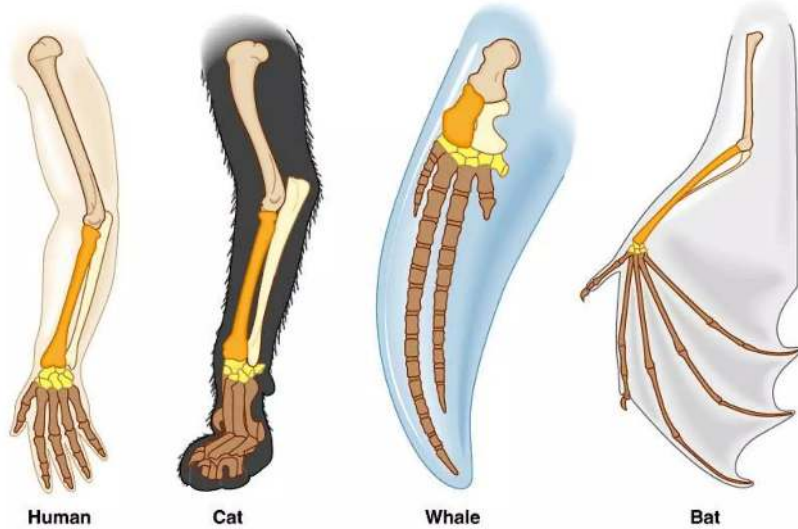
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## Homologous Structures



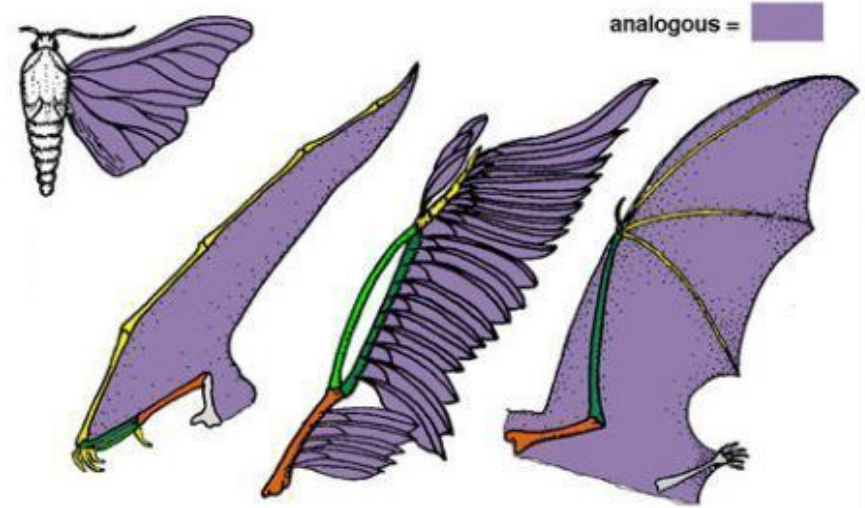
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## Homologous Structures



VS

## Analogous Structures

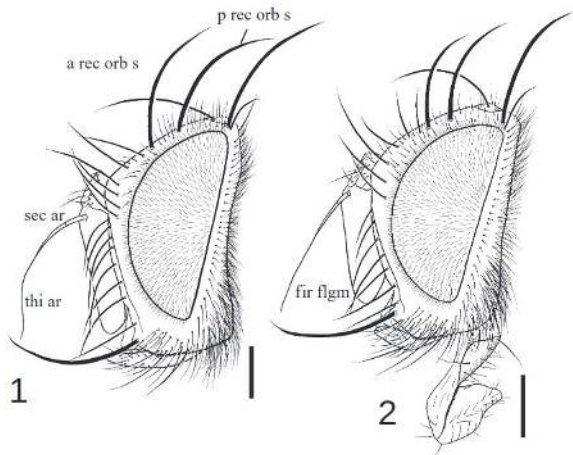


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Takuji Tachi<sup>A,C</sup> and Hiroshi Shima<sup>B</sup>

*Invertebrate Systematics*, 2006, **20**, 255–287



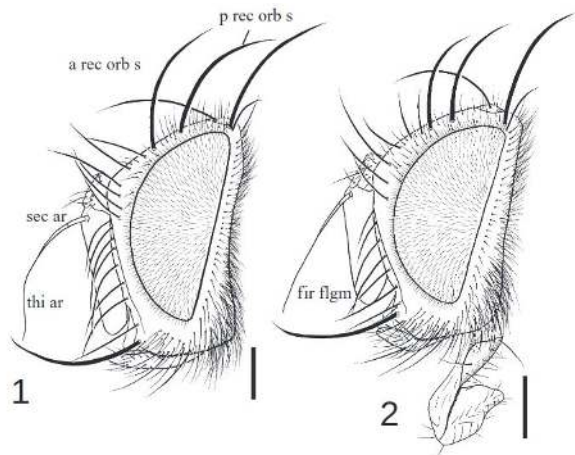
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Lengths (L), consistency indices (CI) and retention indices (RI) are described from the unweighted analysis.

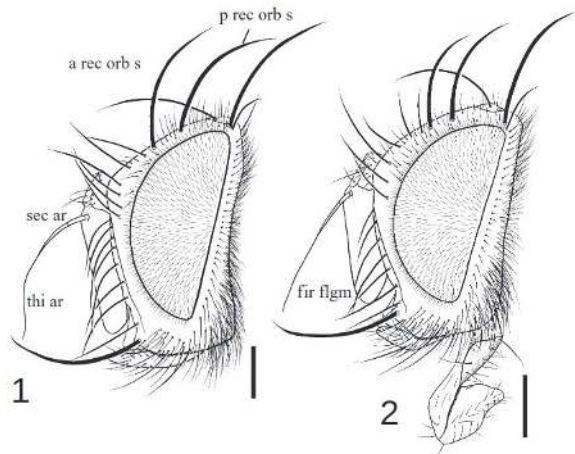
- |     |                                                                                                                                                            |
|-----|------------------------------------------------------------------------------------------------------------------------------------------------------------|
| (1) | Eye: 0, setulose (Figs 1–4); 1, bare or sparsely haired. L = 4; CI = 0.25; RI = 0.73.                                                                      |
| (2) | Ocellar setae: 0, present and strong (Figs 1–4); 1, absent or short and weak. L = 2; CI = 0.50; RI = 0.50.                                                 |
| (3) | Facial ridge: 0, bare; 1, with short setae; 2, with strong setae (Figs 1–4). L = 3; CI = 0.67; RI = 0.94.                                                  |
| (4) | Occiput: 0, without black setulae behind postocular row; 1, with black setulae behind postocular row. L = 2; CI = 0.50; RI = 0.86.                         |
| (5) | First supra-alar setae (sa): 0, longer than first intra-alar seta (ia); 1, shorter than first intra-alar seta. L = 1; CI = 1; RI = 0.                      |
| (6) | Apical scutellar setae: 0, horizontal or absent; 1, directed upwards. L = 4; CI = 0.25; RI = 0.81.                                                         |
| (7) | Setae on vein $R_{4+5}$ : 0, only base (at most to halfway to crossvein r-m); 1, from base nearly to crossvein r-m or beyond. L = 3; CI = 0.33; RI = 0.89. |

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**Table 3.** Morphological data matrix used for phylogenetic analysis

| Taxa                           | Characters               |                          |                          |
|--------------------------------|--------------------------|--------------------------|--------------------------|
|                                | 0000000001<br>1234567890 | 1111111112<br>1234567890 | 2222222223<br>1234567890 |
| <i>Winthemia venusta</i>       | 0000000000               | 0000000000               | –000001000               |
| <i>Drinomyia hokkaidensis</i>  | 1000100001               | 0100000000               | –000002000               |
| <i>Phorocerosoma vicarium</i>  | 0000100000               | 0010000000               | –000001000               |
| <i>Austrophorocera grandis</i> | 0120100000               | 0101010001               | 0000003000               |
| <i>A. hirsuta</i>              | 0020100000               | 0001010001               | 0000003000               |
| <i>Bessa parallela</i>         | 1021101000               | 0001010001               | 1000003000               |
| <i>B. ...</i>                  | 1001101000               | 0001010001               | 1000003000               |

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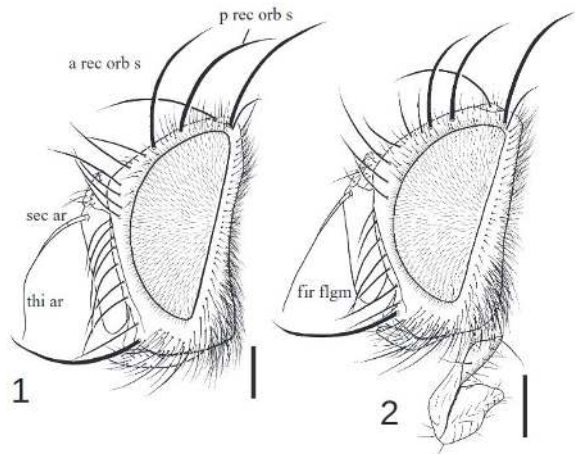
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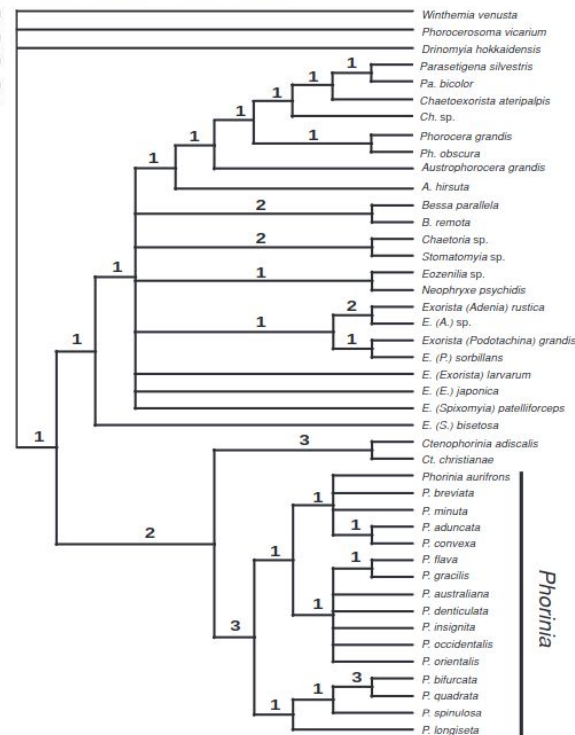
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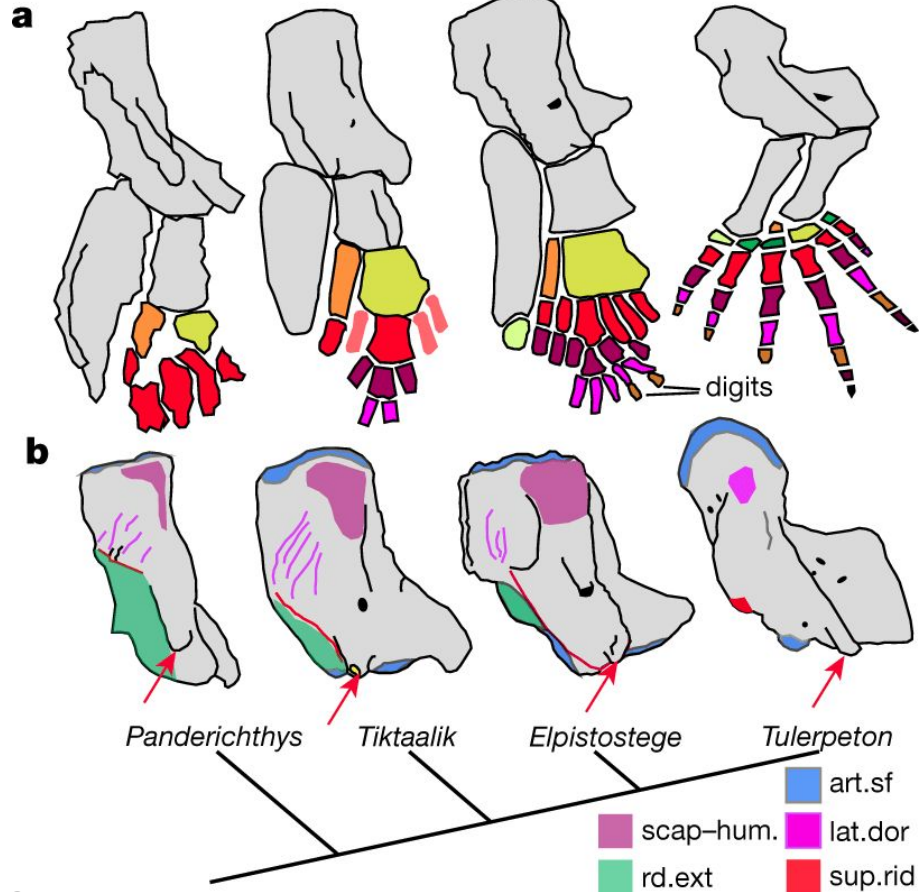
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| <i>B. remota</i>               | 1021101000 |            |            |

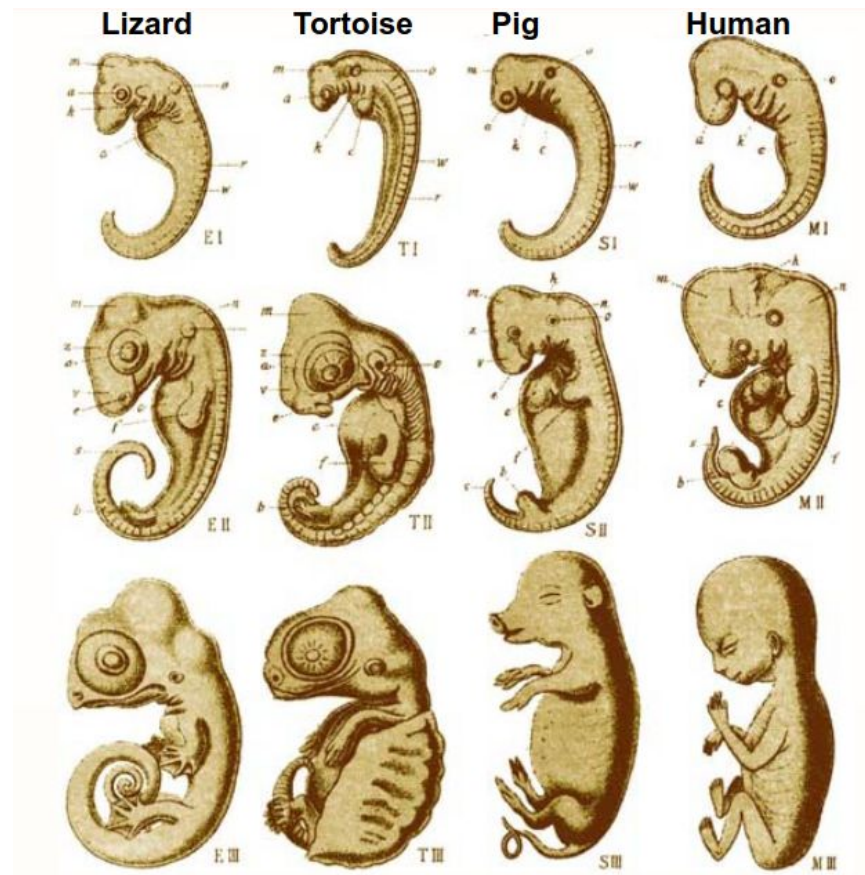
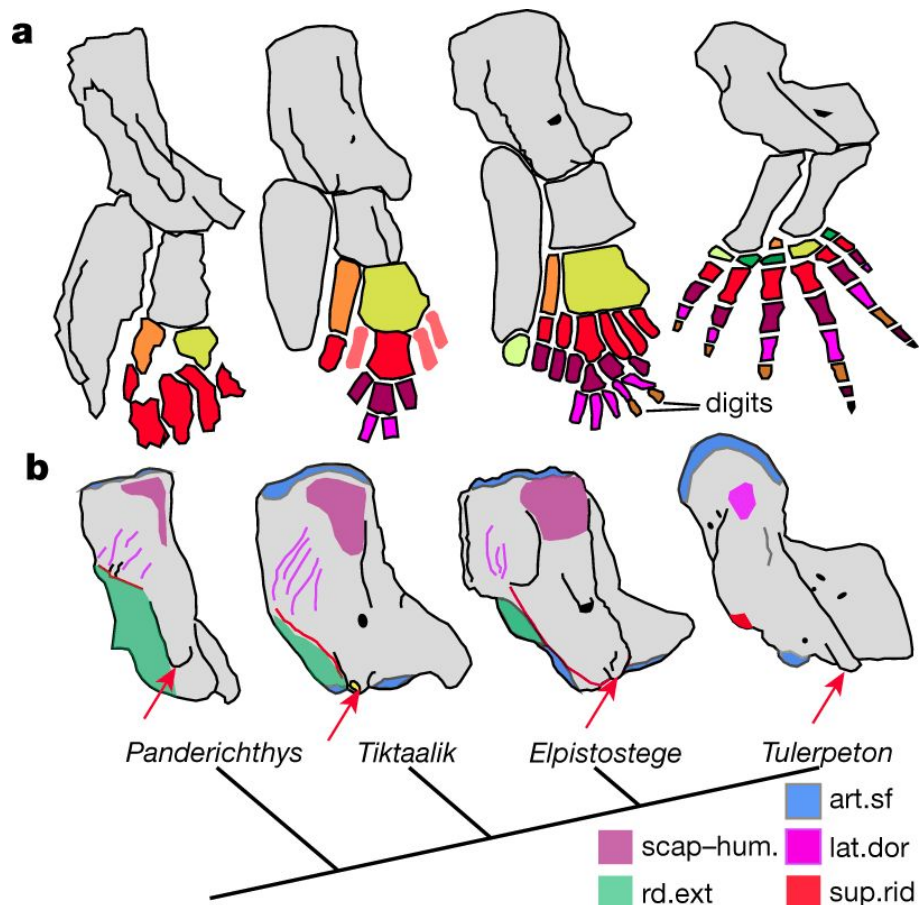


**Fig. 79.** Strict consensus of 186 equally most parsimonious cladograms (length = 66, consistency index (CI) = 0.530, rescaled consistency index (RCI) = 0.462) generated from an analysis of thirty-one morphological characters. Bremer support values are given on the branches.

# What is a phylogeny, why is it important... and how do you build one?



# What is a phylogeny, why is it important... and how do you build one?



# **The origin of molecular phylogenetics**

# The origin of molecular phylogenetics

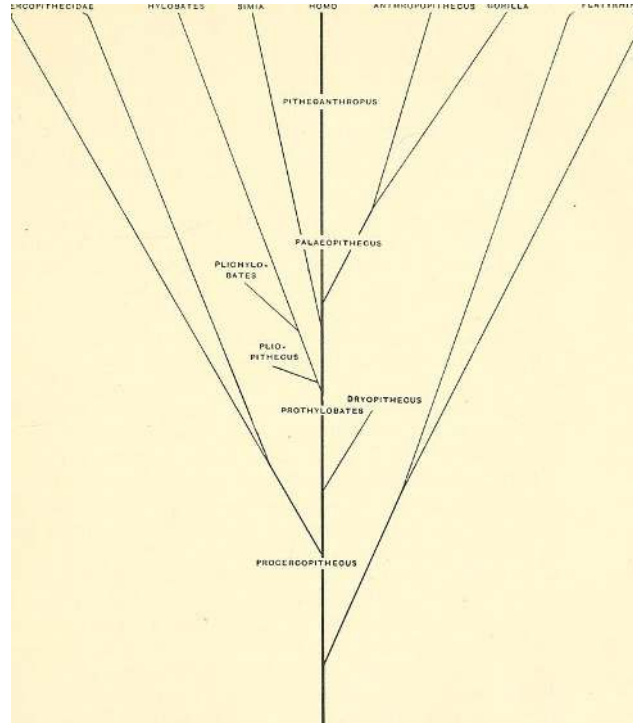
Nuttall (1904) - serological cross-reactions were stronger for more closely related organisms -> phylogeny of apes

BLOOD IMMUNITY  
AND  
BLOOD RELATIONSHIP  
A DEMONSTRATION OF CERTAIN BLOOD-RELATIONSHIPS  
AMONGST ANIMALS BY MEANS OF  
THE PRECIPITIN TEST FOR BLOOD

by  
GEORGE H. F. NUTTALL, M.A., M.D., PH.D.  
University Lecturer in Bacteriology and Preventive Medicine, Cambridge.

Including  
Original Researches by  
G. S. GRAHAM-SMITH, M.A., M.B., D.P.H. (Camb.)  
and  
T. S. P. STRANGEWAYS, M.A., M.R.C.S.

CAMBRIDGE :  
at the University Press  
1904



# The origin of molecular phylogenetics

Nuttall (1904) - serological cross-reactions were stronger for more closely related organisms -> phylogeny of apes

Dobzhansky & Sturtevant (1938) - genomic rearrangements in *Drosophila* as phylogenetic markers

## BLOOD IMMUNITY AND BLOOD RELATIONSHIP

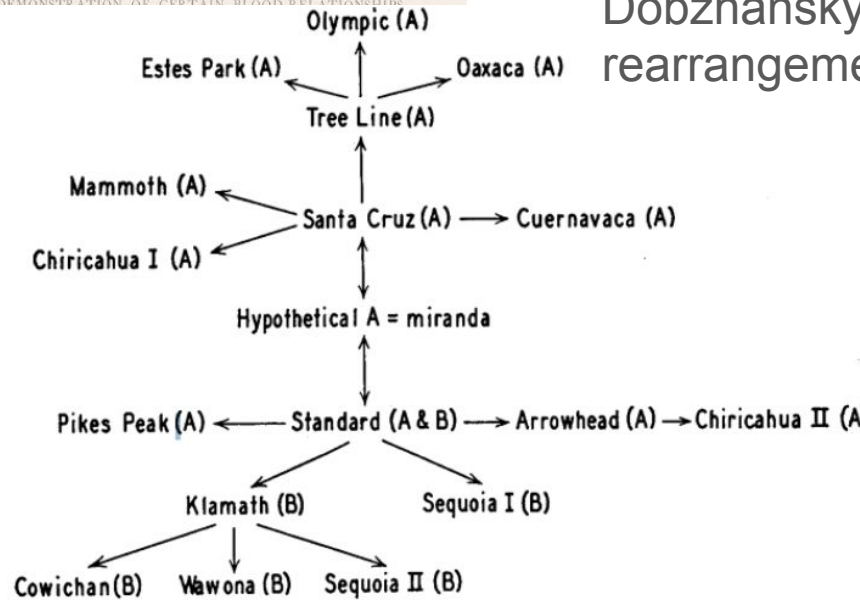
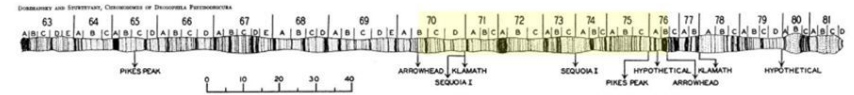
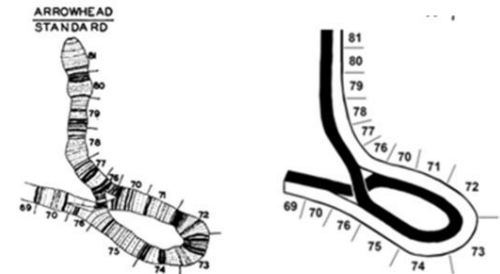


FIGURE 3.—Phylogeny of the gene arrangements in the third chromosome of *Drosophila pseudoobscura*. Any two arrangements connected by an arrow in the diagram differ by a single inversion. Further explanation in text.



Chromosome 3 of *Drosophila pseudoobscura*



Standard and Arrowhead arrangements differ by an inversion from segments 70 to 76

# The origin of molecular phylogenetics

Nuttall (1904) - serological cross-reactions were stronger for more closely related organisms -> phylogeny of apes

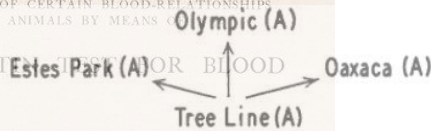
Dobzhansky & Sturtevant (1938) - genomic rearrangements in *Drosophila* as phylogenetic markers

Zuckerkandl & Pauling (1965) -

## BLOOD IMMUNITY AND BLOOD RELATIONSHIP

A DEMONSTRATION OF CERTAIN BLOOD-RELATIONSHIPS  
AMONGST ANIMALS BY MEANS OF

THE PRECIPITATION TEST FOR BLOOD



Olympic (A)  
Estes Park (A)      Oaxaca (A)  
Tree Line (A)



ELSEVIER

Journal of Theoretical Biology

Volume 8, Issue 2, March 1965, Pages 357-366



## Molecules as documents of evolutionary history ☆

Emile Zuckerkandl, Linus Pauling

version. Further explanation in text.

# The origin of molecular phylogenetics

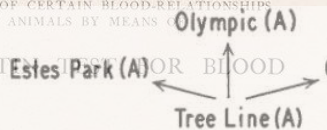
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ELSEVIER

## Journal of Theoretical Biology

Volume 8, Issue 2, March 1965, Pages 357-366



Zuckerlandl &  
Pauling (1965) -

### Abstract

## Molecule history ☆

Emile Zuckerlandl, I

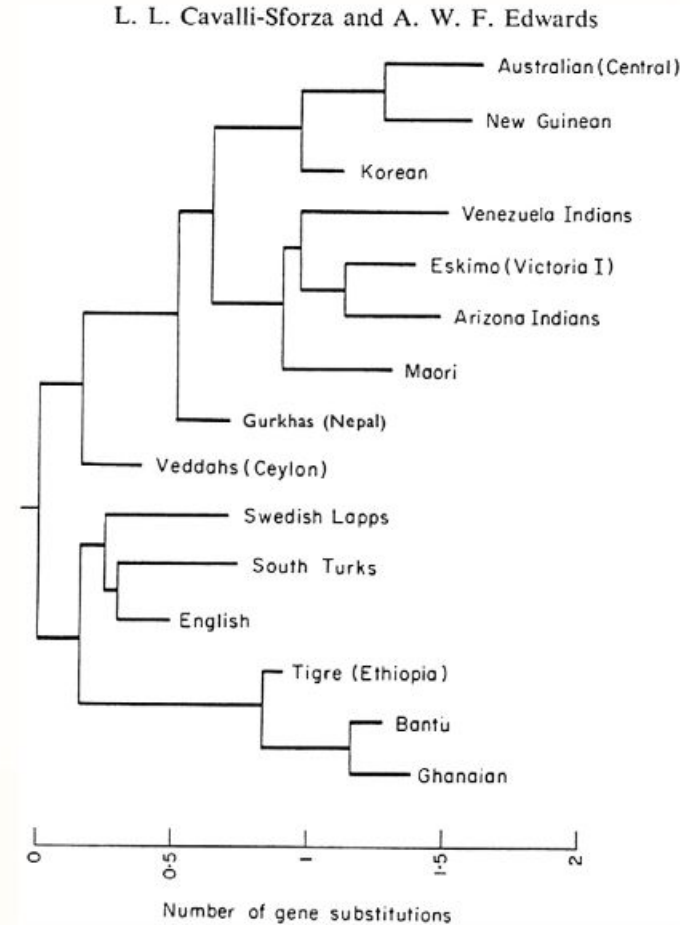
Different types of molecules are discussed in relation to their fitness for providing the basis for a molecular phylogeny. Best fit are the “semantides”, i.e. the different types of macromolecules that carry the genetic information or a very extensive translation thereof. The fact that more than one coding triplet may code for a given amino acid

version. Further explanation in

# Molecular phylogenetics: the new wave



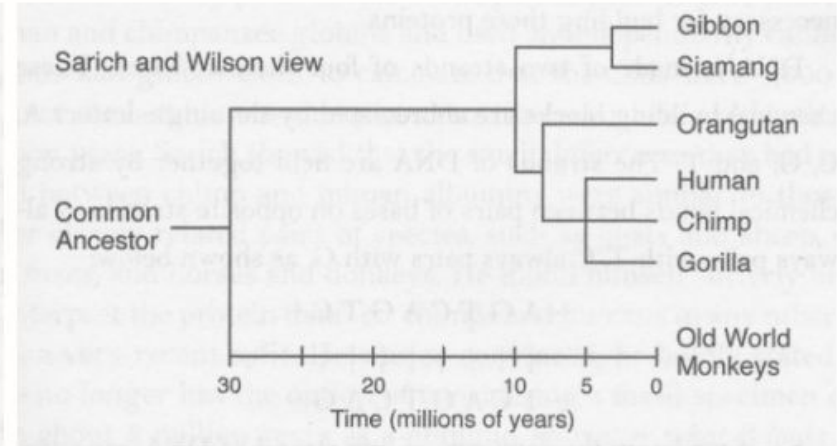
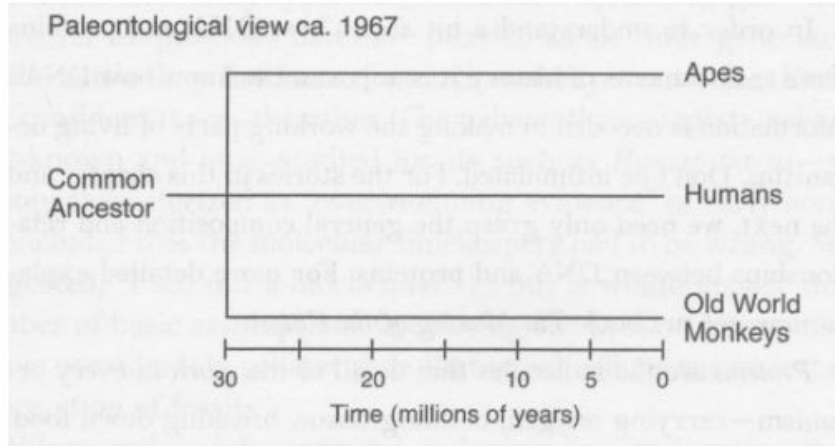
**Phylogeny inferred from blood group allele frequencies from 15 populations**



Cavalli-Sforza & Edwards (1965) in *Genetics Today*

# Molecular phylogenetics: the new wave

**Divergence times were estimated by measuring the immunological cross-reaction of blood serum albumin between pairs of primates**



**“no fuss, no muss, no dishpan hands. Just throw some proteins into a laboratory apparatus, shake them up, and bingo! – we have an answer to questions that have puzzled us for three generations.”**

Sarich & Wilson (1967) Science

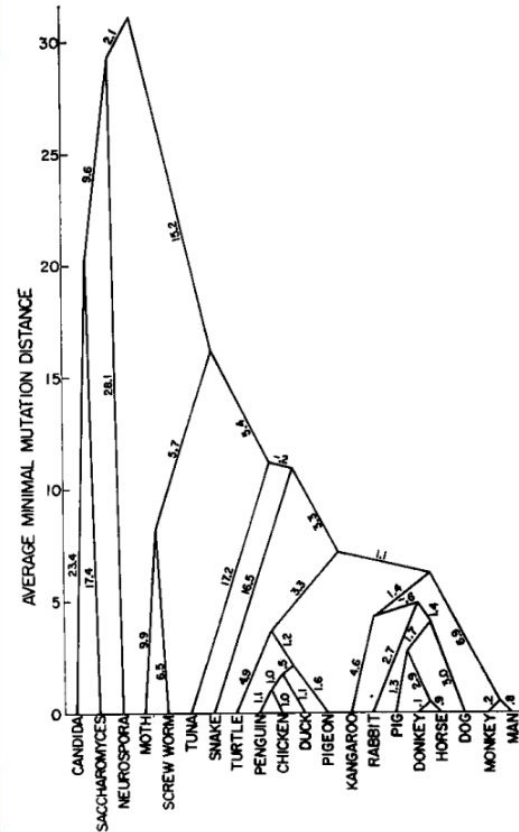
## Construction of Phylogenetic Trees

A method based on mutation distances as estimated from cytochrome *c* sequences is of general applicability.

Walter M. Fitch and Emanuel Margoliash

Biochemists have attempted to use quantitative estimates of variance between substances obtained from different species to construct phylogenetic trees. Examples of this approach include studies of the degree of interspecific hybridization of DNA (1), the degree of cross reactivity of antisera to purified proteins (2), the number of differences in the peptides from enzymic digests of purified homol-

ogous proteins, both as estimated by paper electrophoresis-chromatography or column chromatography and as estimated from the amino acid compositions of the proteins (3), and the number of amino acid replacements between homologous proteins whose complete primary structures had been determined (4). These methods have not been completely satisfactory because (i) the portion of the genome examined



# Molecular phylogenetics: the new wave

*Proc. Natl. Acad. Sci. USA*  
Vol. 74, No. 11, pp. 5088-5090, November 1977  
Evolution

## Phylogenetic structure of the prokaryotic domain: The primary kingdoms

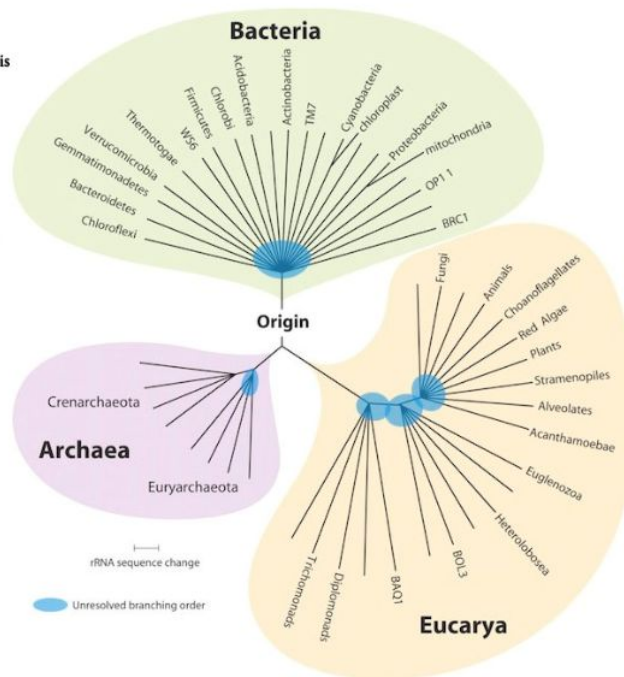
(archaeobacteria/eubacteria/urkaryote/16S ribosomal RNA/molecular phylogeny)

CARL R. WOESE AND GEORGE E. FOX\*

Department of Genetics and Development, University of Illinois, Urbana, Illinois

Communicated by T. M. Sonneborn, August 18, 1977

**ABSTRACT** A phylogenetic analysis based upon ribosomal RNA sequence characterization reveals that living systems represent one of three aboriginal lines of descent: (i) the eubacteria, comprising all typical bacteria; (ii) the archaeobacteria, containing methanogenic bacteria; and (iii) the urkaryotes, now represented in the cytoplasmic component of eukaryotic cells.



## The dawn of phylogenomics



## *Insight/Outlook*

### Phylogenomics: Improving Functional Predictions for Uncharacterized Genes by Evolutionary Analysis

Jonathan A. Eisen<sup>1</sup>

Department of Biological Sciences, Stanford University, Stanford, California 94305-5020 USA

**T**he ability to accurately predict gene function based on gene sequence is an important tool in many areas of biological research. Such predictions have become particularly important in the genomics age in which numerous gene sequences are generated with little or no accompanying experimentally determined functional information. Almost all functional prediction methods rely on the identification, characterization,

(e.g., Altschul et al. 1989; Goldman et al. 1996). In this commentary, I discuss the use of evolutionary information in the prediction of gene function. To appreciate the potential of a *phylogenomic approach* to the *prediction of gene function*, it is necessary to first discuss how gene sequence is commonly used to predict gene function and some general features about gene evolution.

convergence (the exact threshold for such an inference is not well established).

Improvements in database search programs have made the identification of likely homologs much faster, easier, and more reliable (Altschul et al. 1997; Henikoff et al. 1998). However, as discussed above, in many cases the identification of homologs is not sufficient to make specific functional predictions be-

*Phylogenomics: prediction of gene function and gene family evolution*

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## Sequence Similarity, Homology, and Functional Predictions

To make use of the identification of sequence similarity between genes, it is helpful to understand how such similarity arises. Genes can become similar in sequence either as a result of *convergence* (similarities that have arisen without a common evolutionary history) or *descent with modification* from a common ancestor (also known as *homology*). It is imperative to recognize that sequence similarity and homology are not interchangeable terms. Not all homologs are similar in sequence (i.e., homologous genes can diverge so much that similarities are difficult or impossible to detect) and not all similarities are due to homology (Reeck et al. 1987; Hillis 1994). Similarity due to convergence, which is likely limited to small regions of genes, can be useful for some functional predictions (Henikoff et al. 1997). However, most sequence-based functional predictions are based on the identification (and subsequent analysis) of similarities that are thought to be due to homology. Because homology is a statement about common ancestry, it cannot be proven directly from sequence similarity. In these cases, the inference of homology is made based on finding levels of sequence similarity that are thought to be too high to be due to

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# The dawn of phylogenomics

8:163-167 ©1998 by Cold Spring Harbor Laboratory Press ISSN 1054-9803/98 \$5.00; www.genome.org

GENOME RESEARCH 163

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*Phylogenomics: prediction of gene function and gene family evolution*

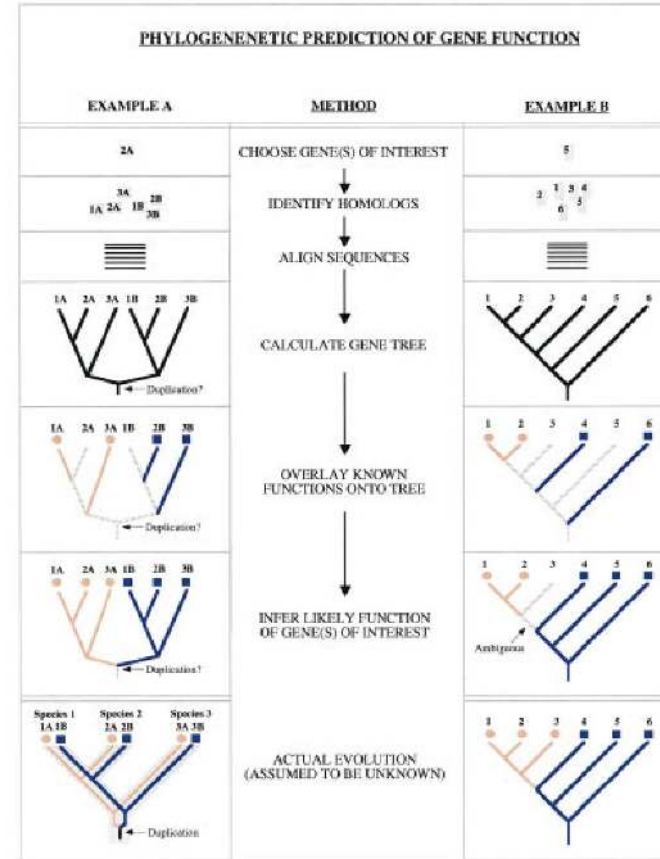


Figure 1 Outline of a phylogenomic methodology. In this method, information about the evolutionary relationships among genes is used to predict the functions of uncharacterized genes (see text for details). Two hypothetical scenarios are presented and the path of trying to

## The analysis of 100 genes supports the grouping of three highly divergent amoebae: *Dictyostelium*, *Entamoeba*, and *Mastigamoeba*

Eric Baptiste\*, Henner Brinkmann†, Jennifer A. Lee‡, Dorothy V. Moore‡, Christoph W. Sensen§, Paul Gordon¶, Laure Duruflé\*, Terry Gaasterland‡, Philippe Lopez\*, Miklós Müller‡, and Hervé Philippe\*||

The phylogenetic relationships of amoebae are poorly resolved. To address this difficult question, we have sequenced 1,280 expressed sequence tags from *Mastigamoeba* *balamuthi* and assembled a large data set containing 123 genes for representatives of three phenotypically highly divergent major amoeboid lineages: Pelobionta, Entamoebidae, and Mycetozoa. Phylogenetic reconstruction was performed on ~25,000 aa positions for 30 species by using maximum-likelihood approaches. All well-established eukaryotic groups were recovered with high statistical support, validating our approach. Interestingly, the three amoeboid lineages strongly clustered together in agreement with the Conosa hypothesis [as defined by T. Cavalier-Smith (1998) *Biol. Rev. Cambridge Philos. Soc.* 73, 203–266]. Two amitochondriate amoebae, the free-living *Mastigamoeba* and the human parasite *Entamoeba*, formed a significant sister group to the exclusion of the mycetozoan *Dictyostelium*. This result suggested that a part of the reductive process in the evolution of *Entamoeba* (e.g., loss of typical mitochondria) occurred in its free-living ancestors. Applying this inexpensive expressed sequence tag approach to many other lineages will surely improve our understanding of eukaryotic evolution.

*Phylogenomics*: species tree inference

# The dawn of phylogenomics

1414–1419 | PNAS | February 5, 2002 | vol. 99 | no. 3

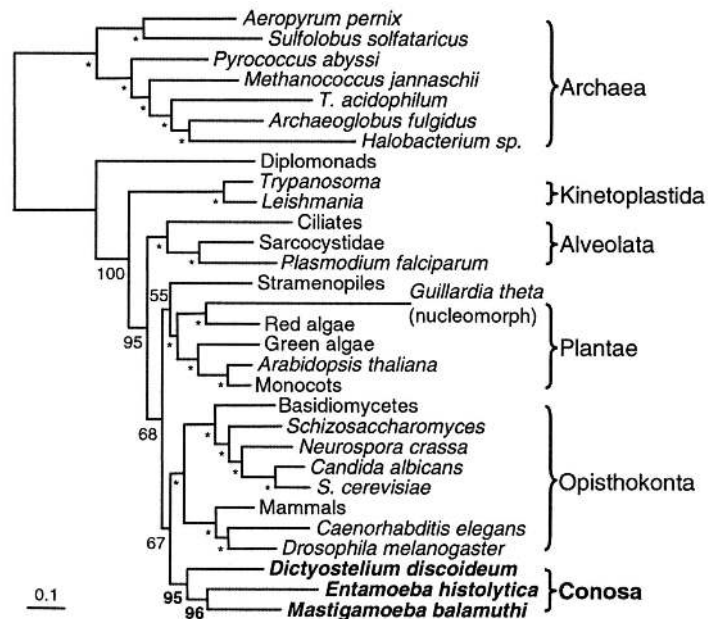
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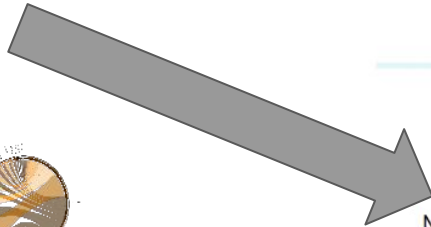
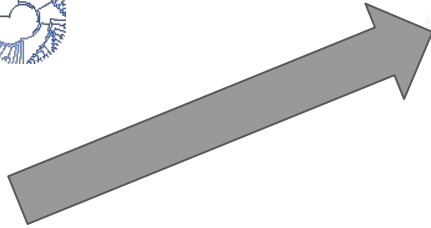
*Phylogenomics: species tree inference*



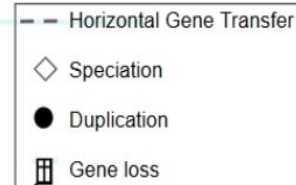
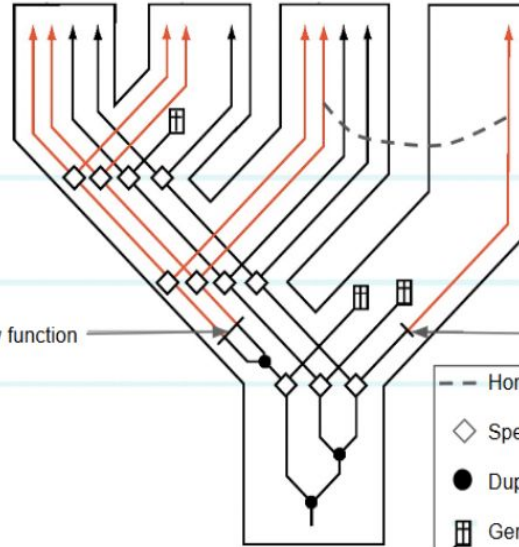
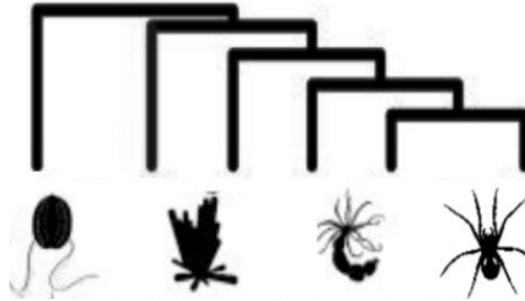
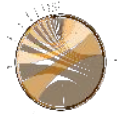
ML tree based on 25,032 aa positions. \* indicates a constrained node. We used the JTT model, without taking into account among-sites rate variation. The branch lengths have been computed on the concatenated sequences. BVs were obtained by bootstrapping the 123 genes.

# The dawn of phylogenomics

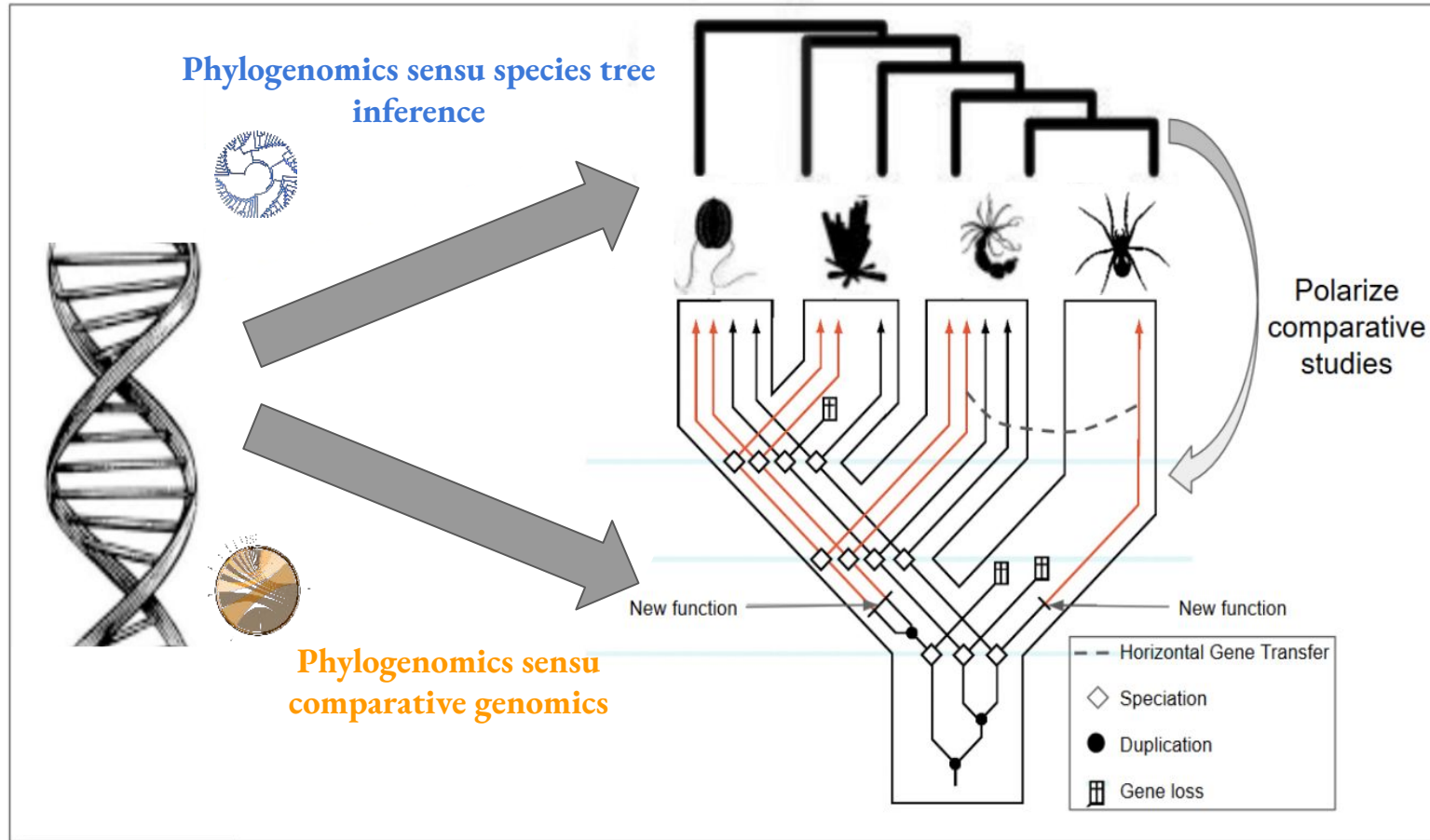
Phylogenomics sensu species tree inference



Phylogenomics sensu comparative genomics



# The dawn of phylogenomics



# Content of the lecture

1

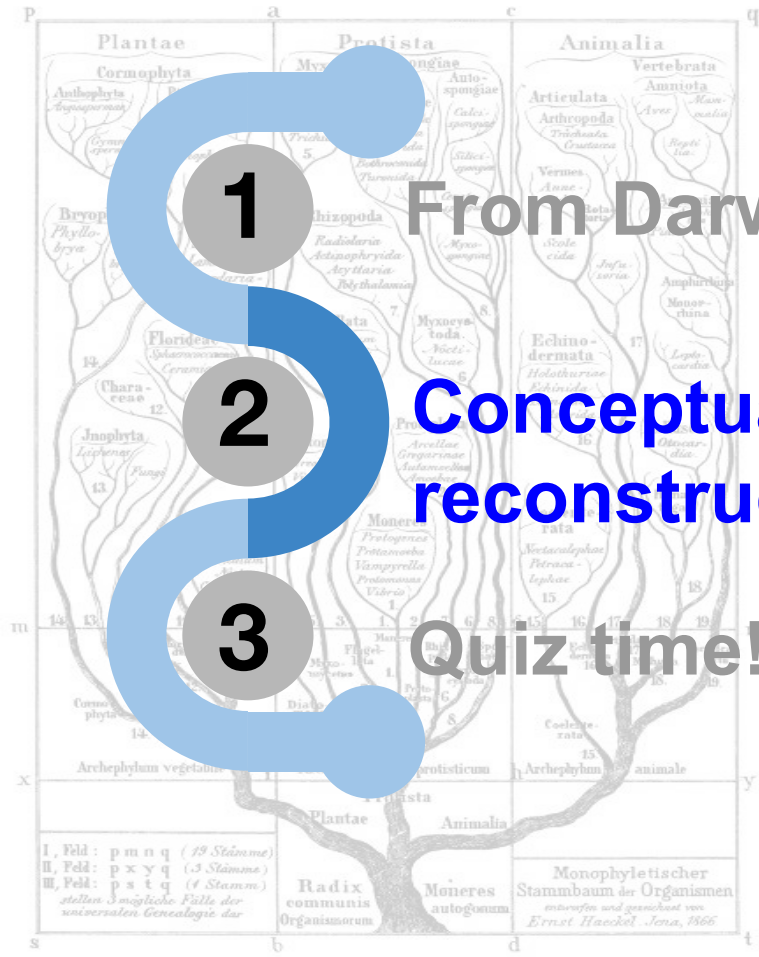
From Darwin to phylogenomics

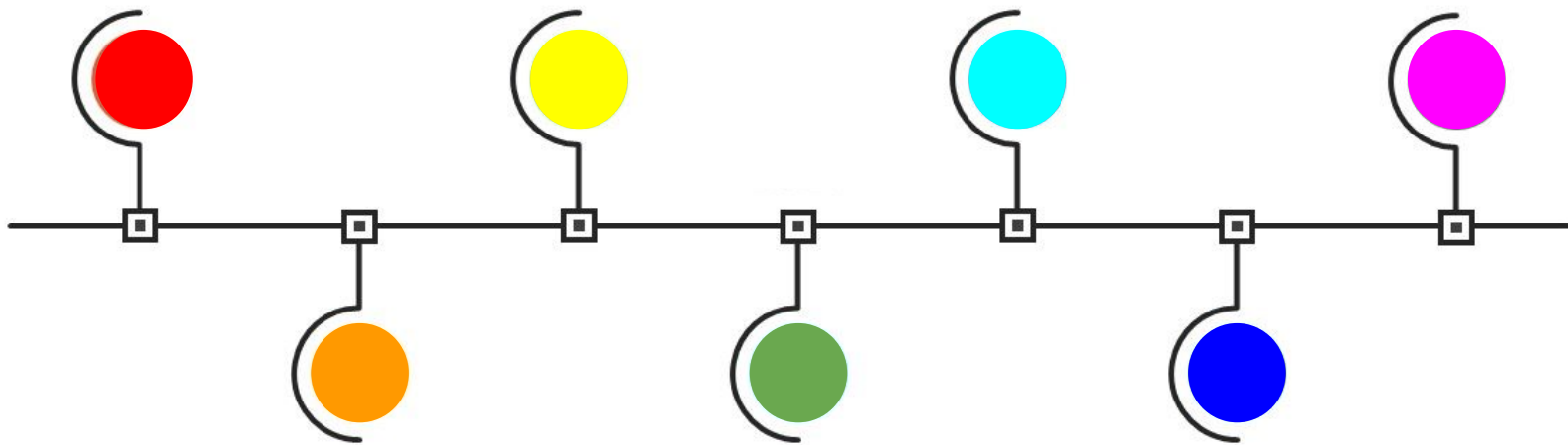
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Conceptual framework for phylogenomic reconstruction

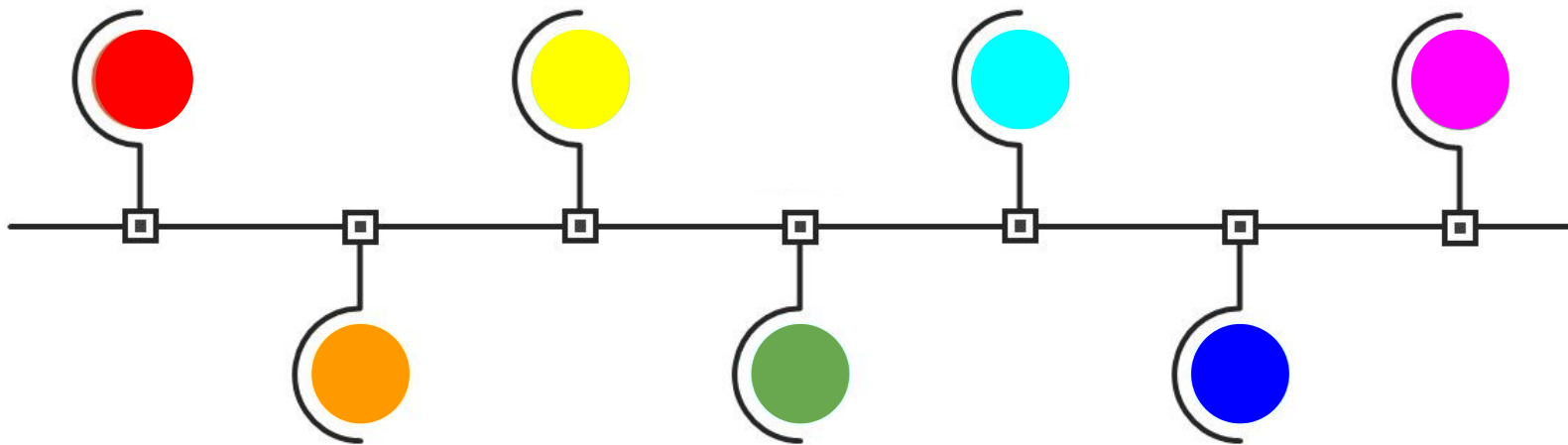
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Quiz time!

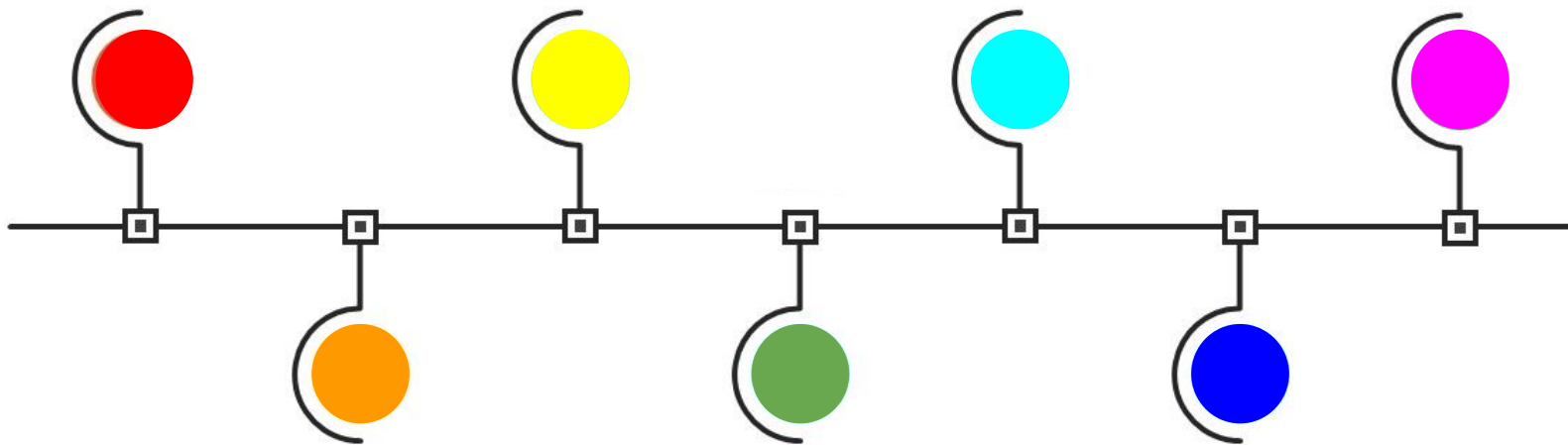




## 01 DATA



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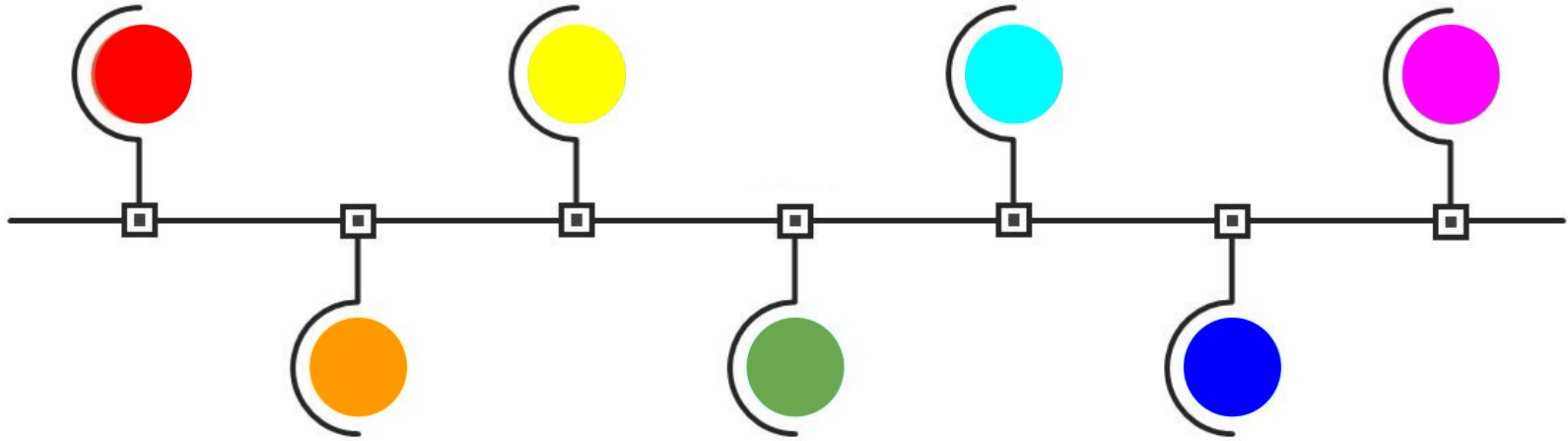


## 02 ORTHOLOGY INFERENCE

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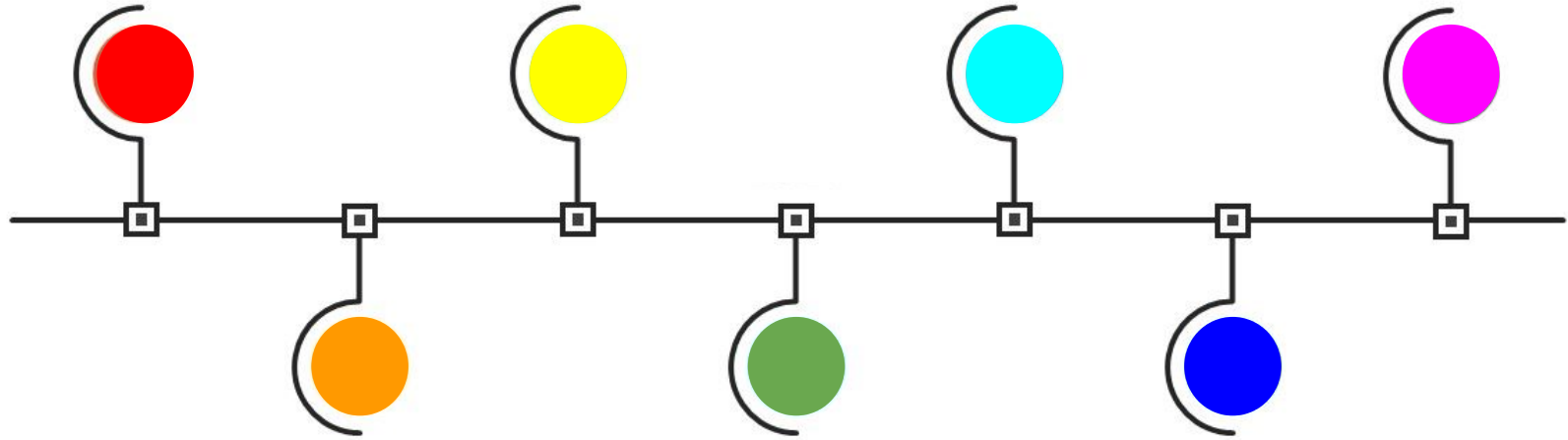
**03 ALIGNMENT  
& TRIMMING**

**02 ORTHOLOGY  
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**04 PHYLOGENOMIC  
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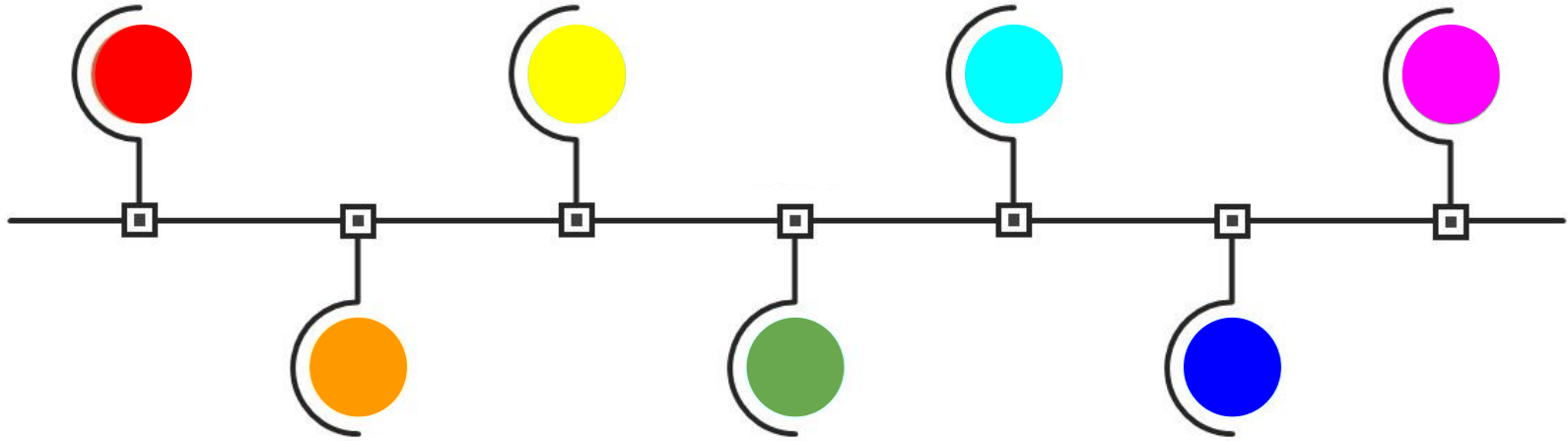
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**05 SUPERMATRIX  
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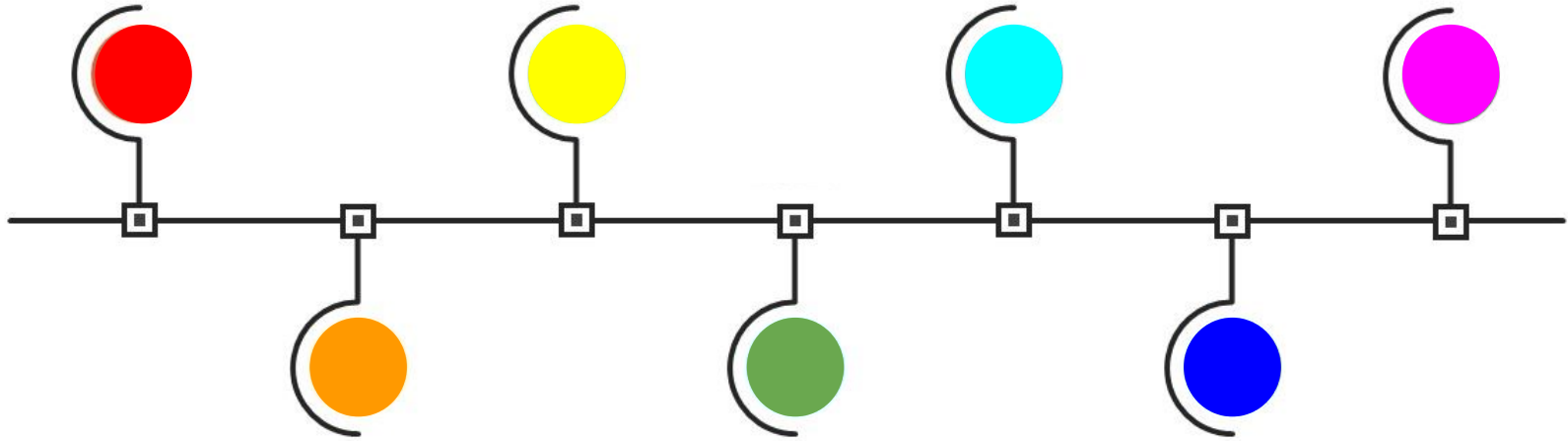
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**06 MODEL  
SELECTION &  
PHYLOGENETIC  
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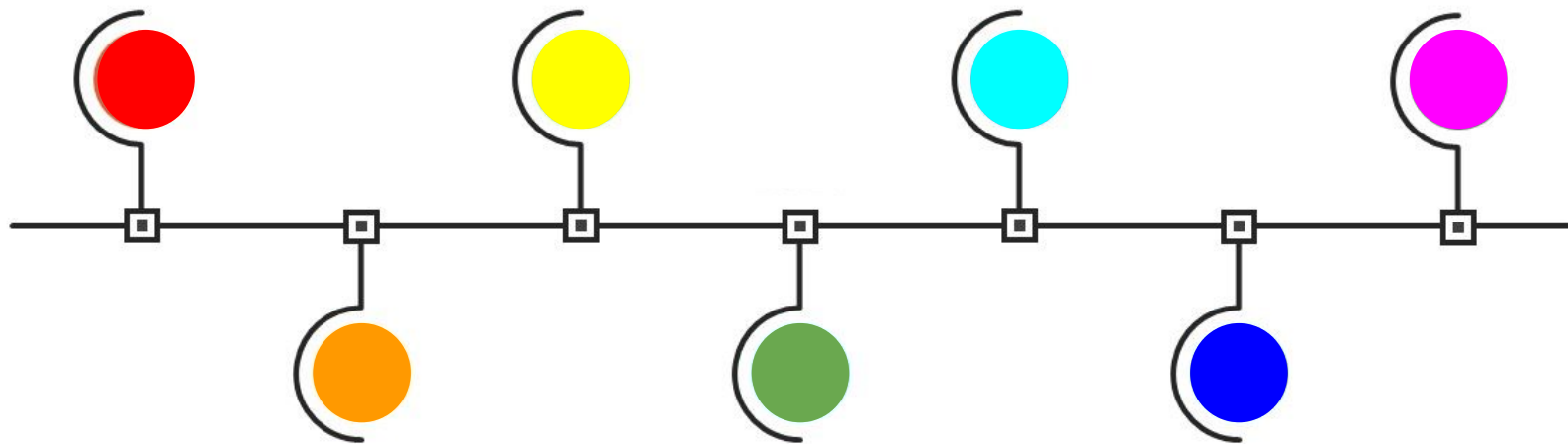
**05 SUPERMATRIX  
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**07 TESTING THE  
ROBUSTNESS  
OF YOUR TREE**

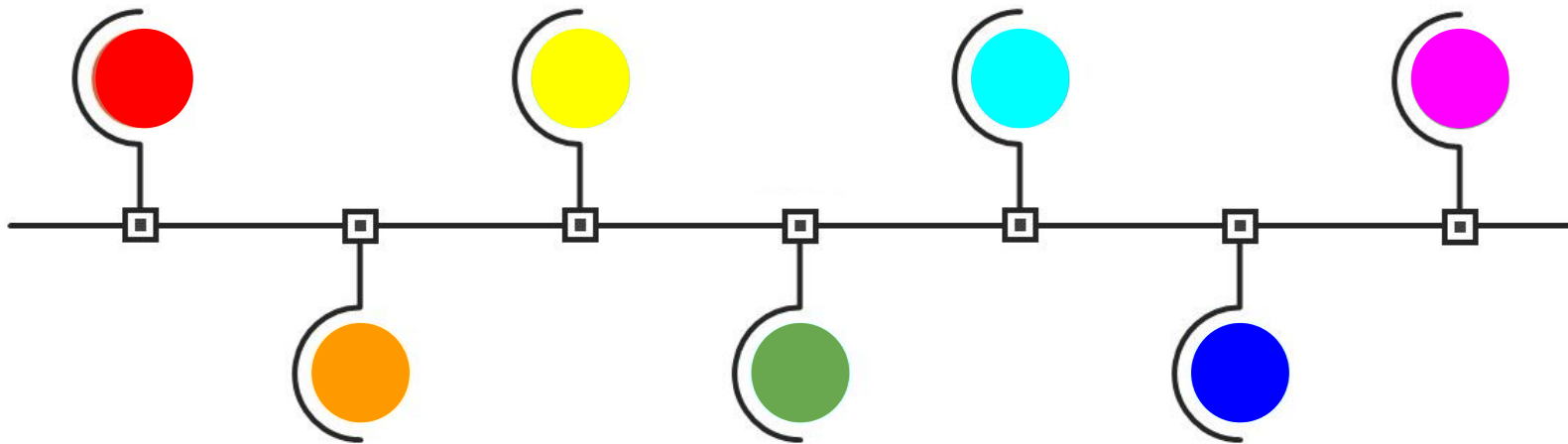
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## 01 DATA



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Incomplete, biased, or improper **taxon sampling** can lead to misleading results in reconstructing evolutionary relationships.

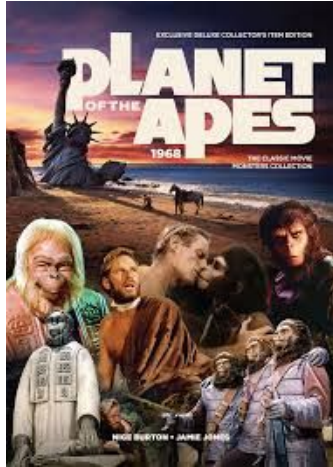
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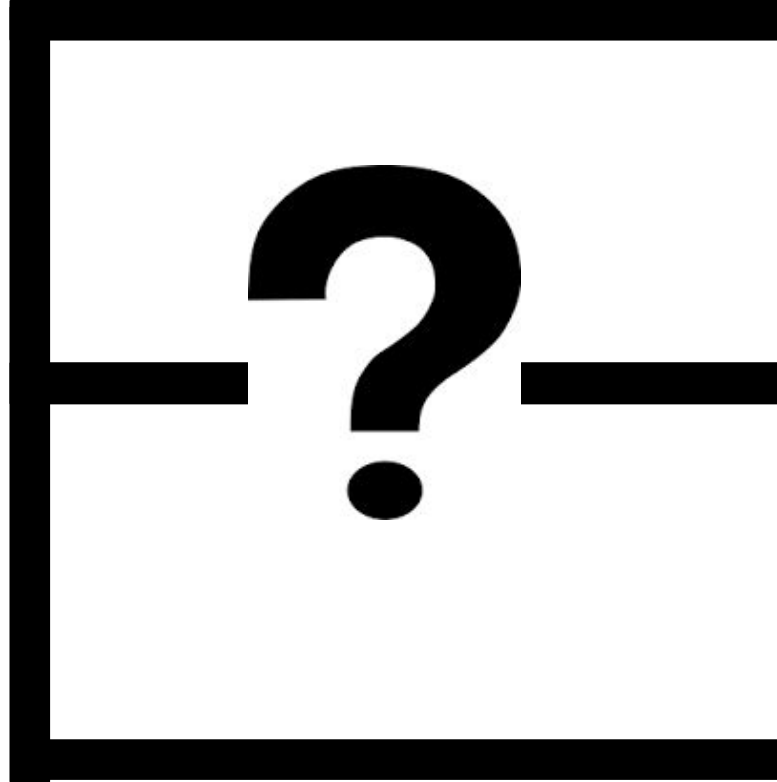
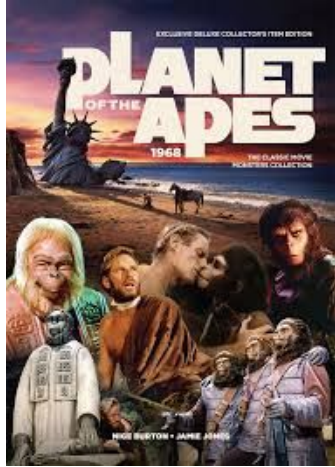
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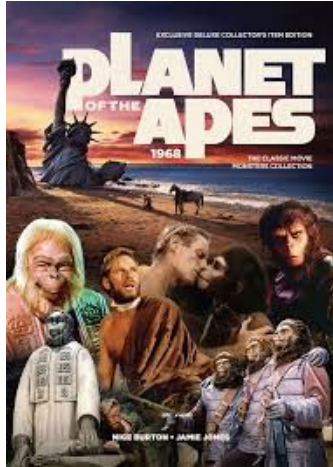
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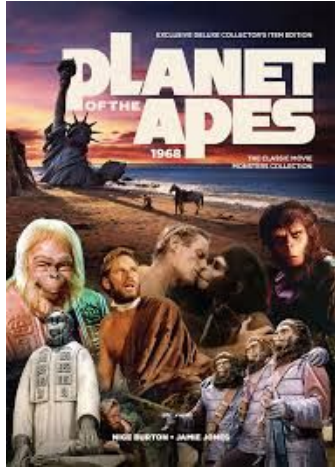
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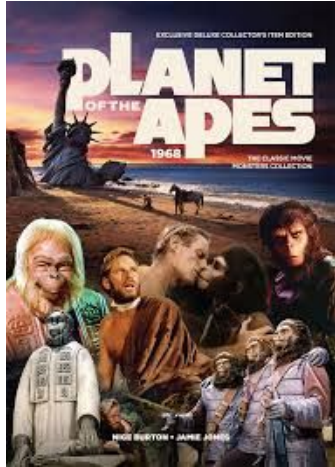
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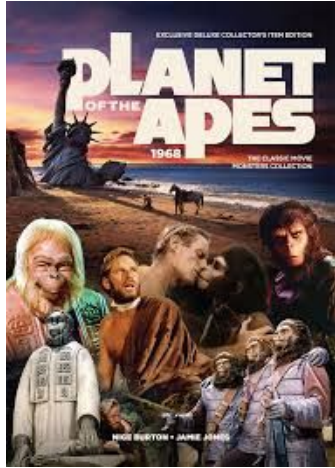
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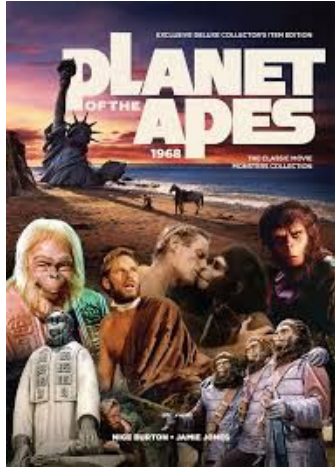
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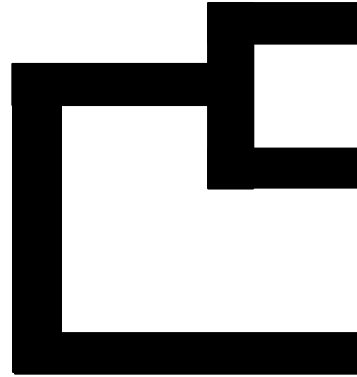
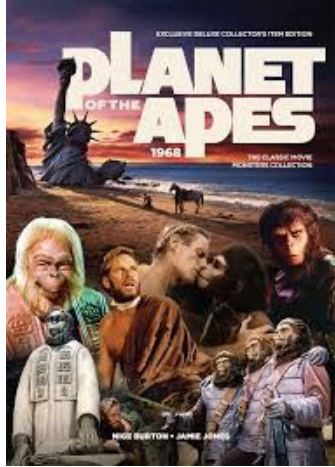
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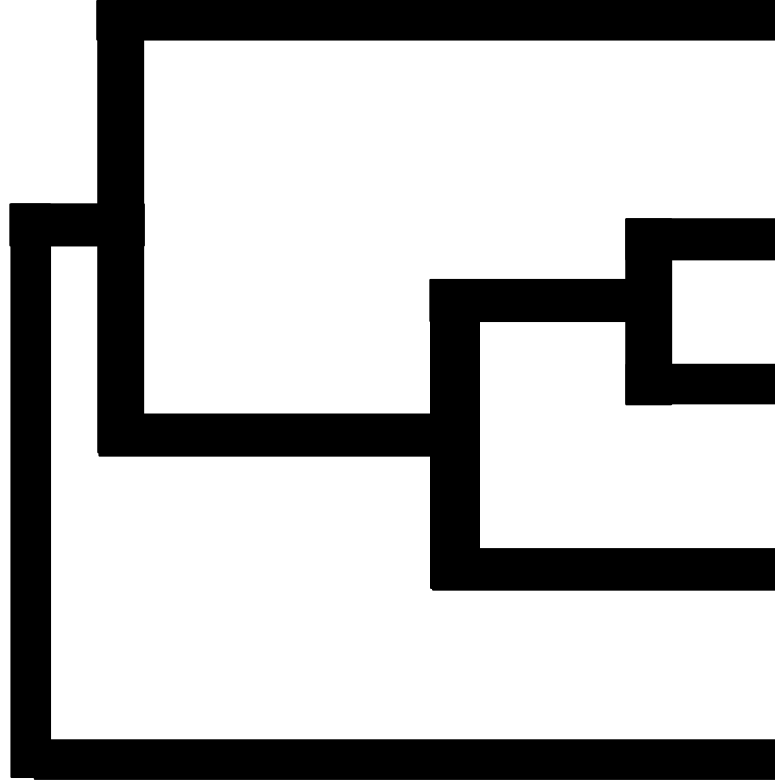
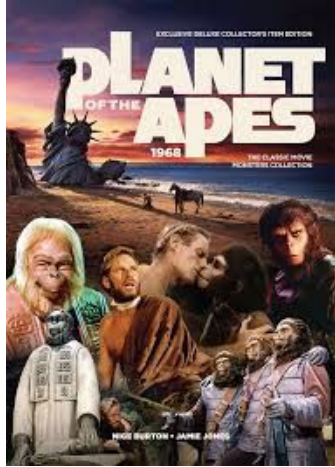
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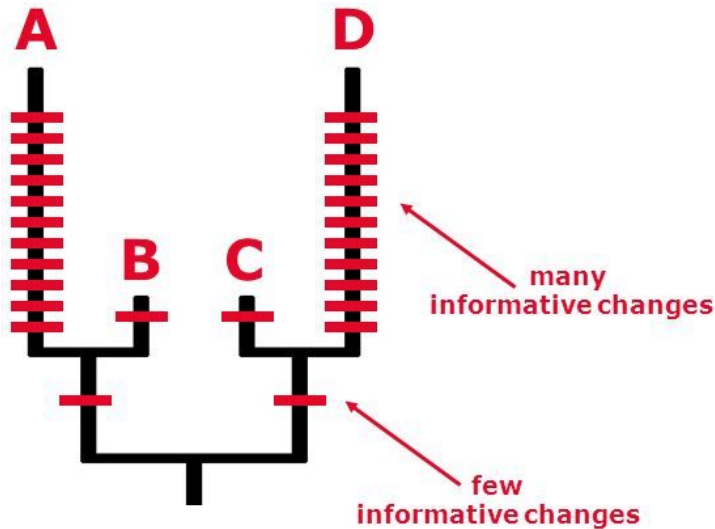


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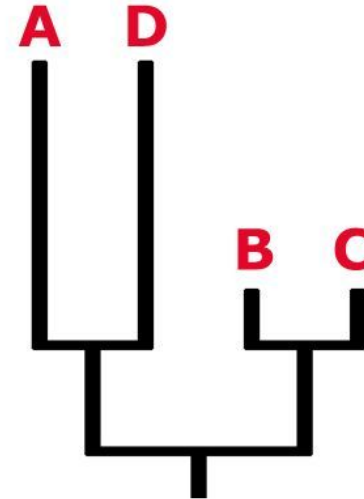
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## Long Branch Attraction

True Tree



Reconstructed Tree



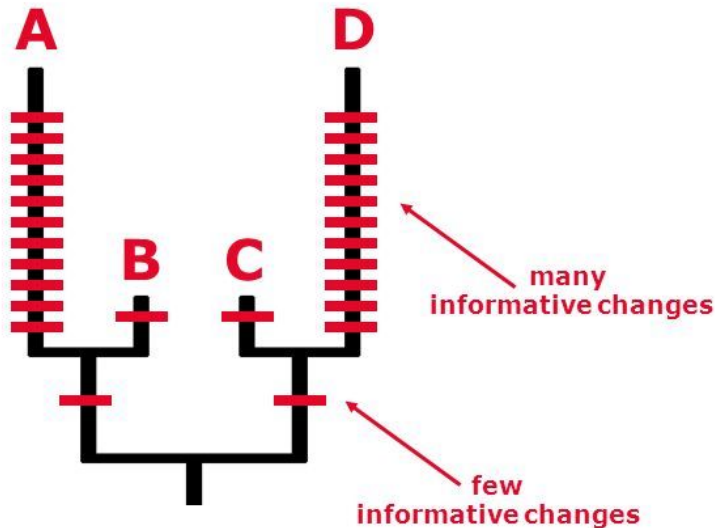
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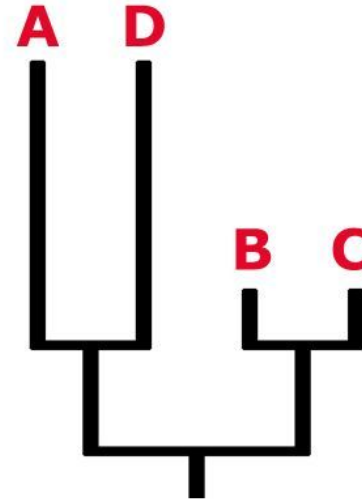
## Long Branch Attraction

Outgroups / Fast-evolving lineages / Missing data

**True Tree**



**Reconstructed Tree**



# 01 DATA

Source of your data

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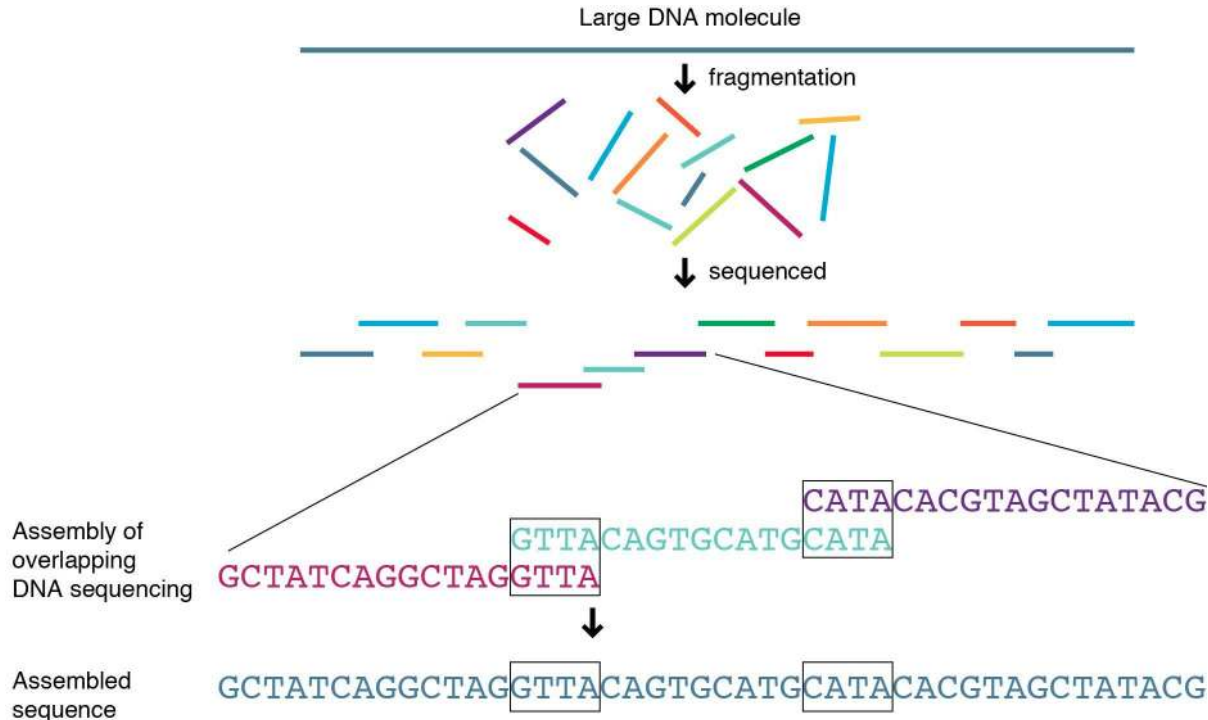
Source of your data

GENOMES

# 01 DATA

## Source of your data

### GENOMES



- Assembled and annotated.
- Coding genes are retrieved (longest isoform) -> this is your dataset!

### GENOMES

#### Pros:

- Very large set of genetic markers
- Good identification of full-length genes, less chimeras (if the assembly and annotation are of good quality)
- Good for shallow and deep evolutionary distances
- Ethanol-fixed tissue OK (for draft genomes)

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#### Cons:

- Annotation may vary quite a lot between species (source, software, etc), may not be comparable.
- Expensive (money and computing time)
- More difficult to have a high number of species
- Fresh tissue needed (for chromosome-level genomes)

# 01 DATA

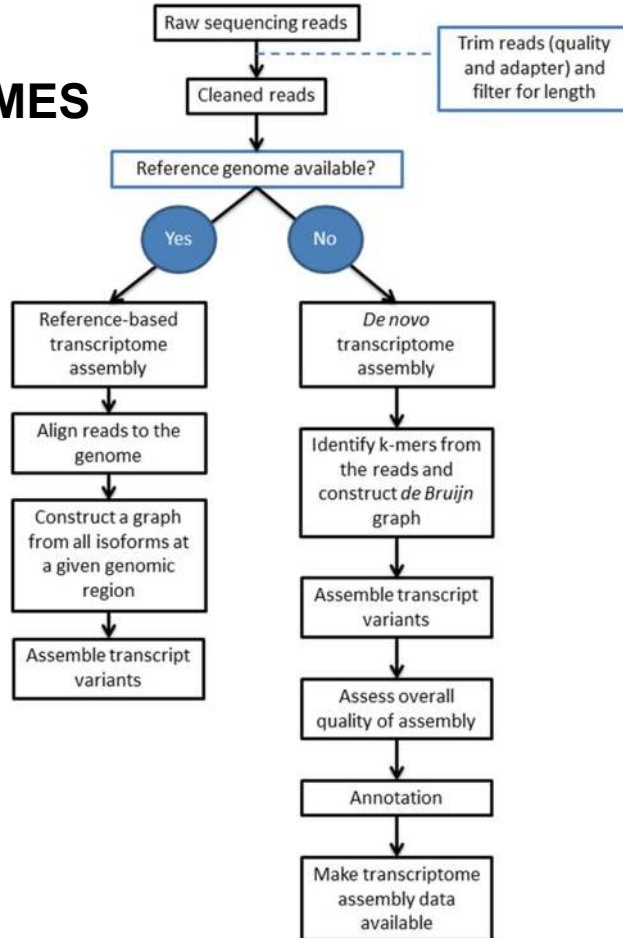
Source of your data

TRANSCRIPTOMES

# 01 DATA

## Source of your data

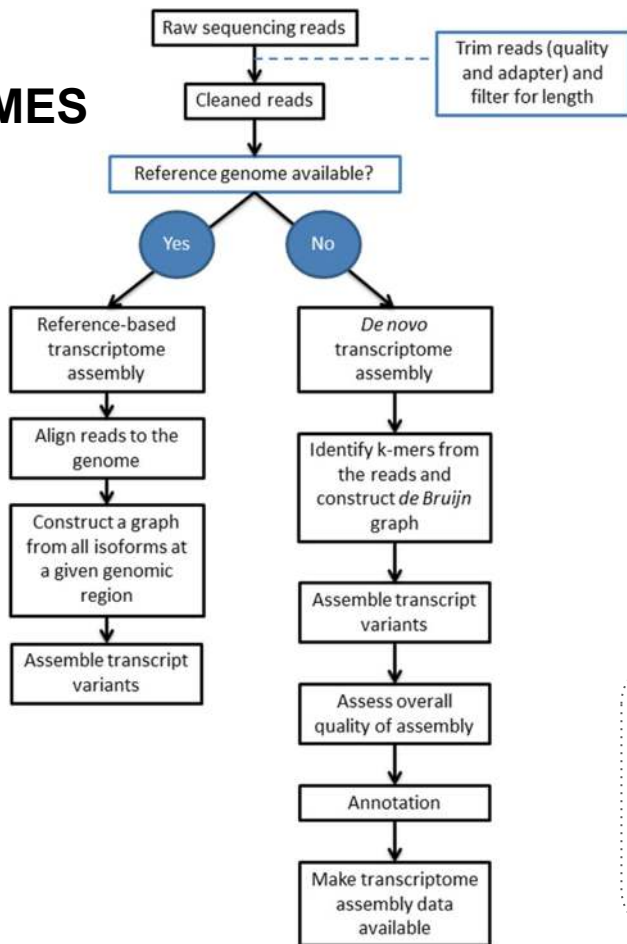
### TRANSCRIPTOMES



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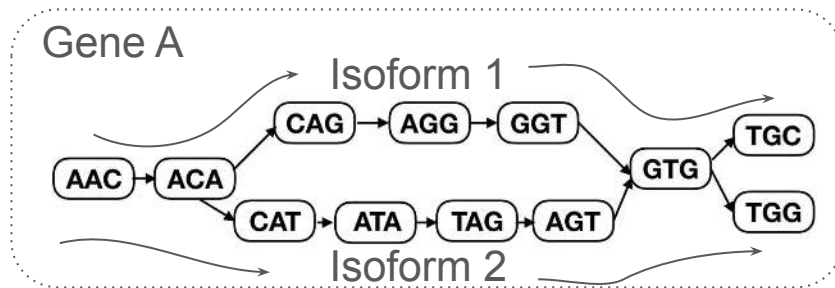
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### TRANSCRIPTOMES



- Assembled de novo

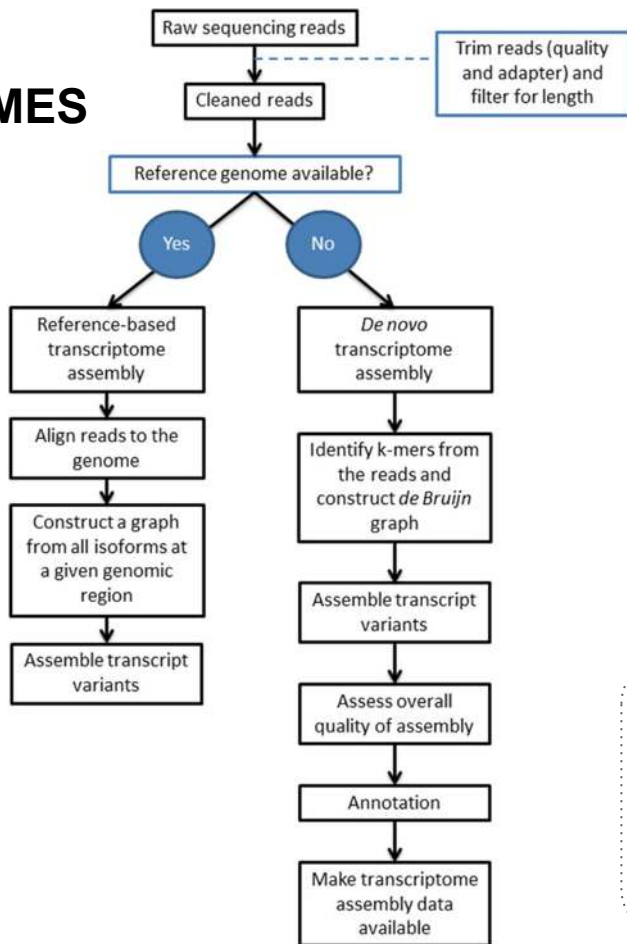
### De Bruijn Graph



# 01 DATA

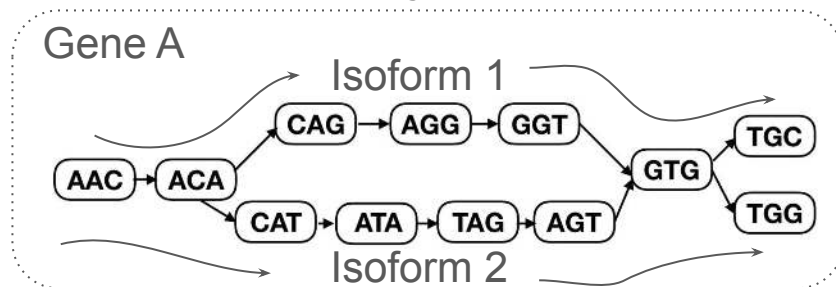
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- Coding genes are retrieved (after inferring ORFs; longest isoform) -> this is your dataset!

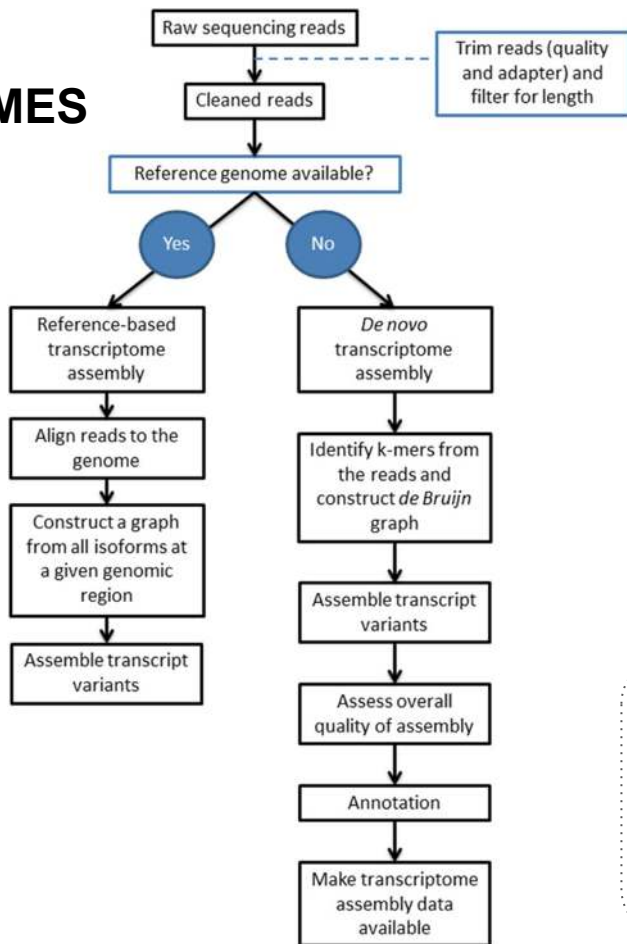
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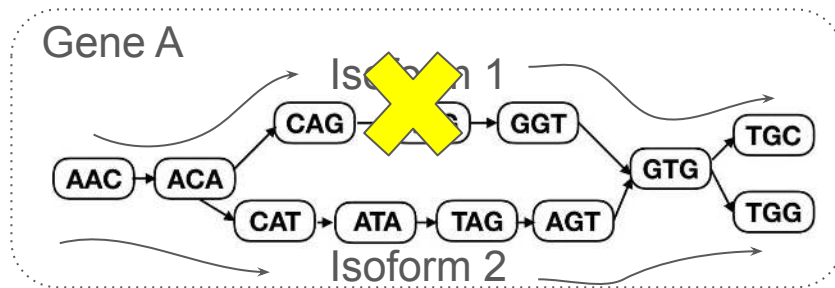
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- Very large set of genetic markers
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#### Cons:

- Incomplete identification of full-length genes and single-copy transcripts.
- Potential misassembly of transcripts (especially when duplicates are present)
- Missing data as a product of the transcriptome representing a snapshot of expression (but this could also affect genome annotation)
- Fresh tissue needed

# 01 DATA

Source of your data

**ULTRACONSERVED ELEMENTS (UCEs)**

## ULTRA-CONSERVED ELEMENTS (UCEs)

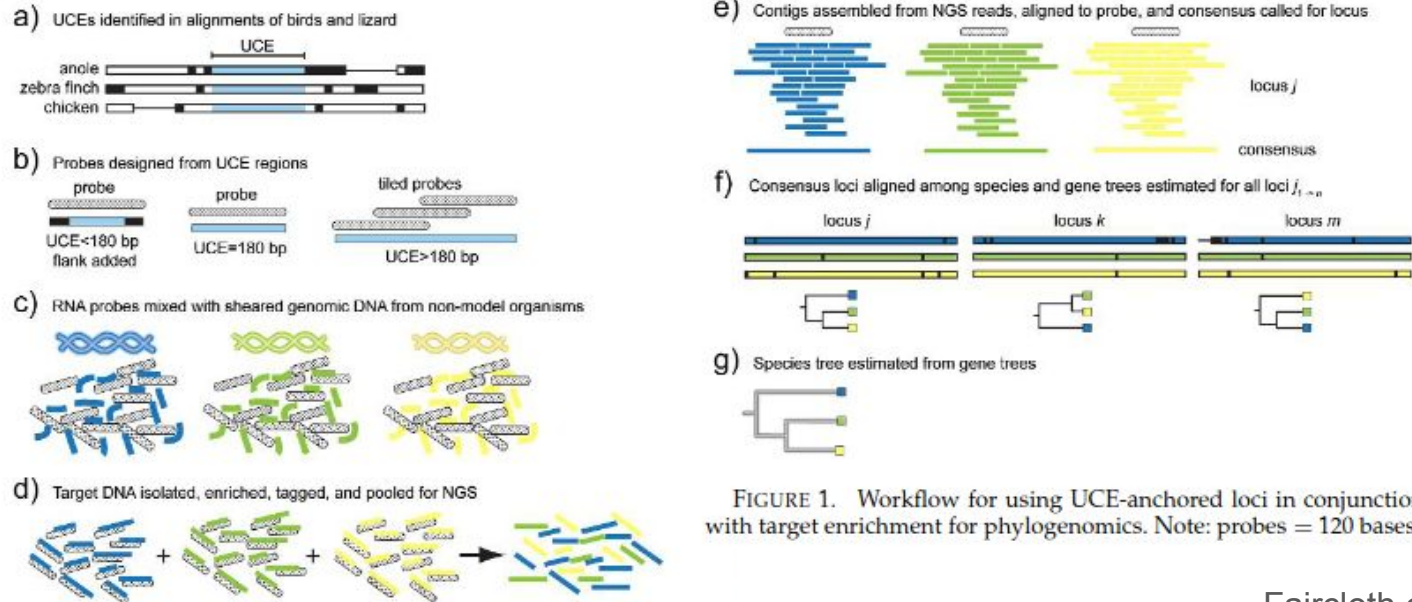


FIGURE 1. Workflow for using UCE-anchored loci in conjunction with target enrichment for phylogenomics. Note: probes = 120 bases.

Faircloth et al. 2012

The UCEs are designed a priori -> after hybridization, sequencing, assembly and mapping, this is your data!

### ULTRA-CONSERVED ELEMENTS (UCEs)

#### Pros:

- Medium-large set of genetic markers
- Much cheaper than sequencing genomes -> easier to have a high number of species
- Not dependent upon a reference genome
- Tissues fixed in EtOH or museum specimens are OK

(Lisa Pokorny's talk on 31st Jan)

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#### Pros:

- Medium-large set of genetic markers
- Much cheaper than sequencing genomes -> easier to have a high number of species
- Not dependent upon a reference genome
- Tissues fixed in EtOH or museum specimens are OK

#### Cons:

- Limited availability of marks outside the designed ones.
- Potential misassembly (if probes are designed with a limited amount of species)
- Retrieval success dependent on DNA quality
- Usefulness of markers known a posteriori
- No proper orthology inference



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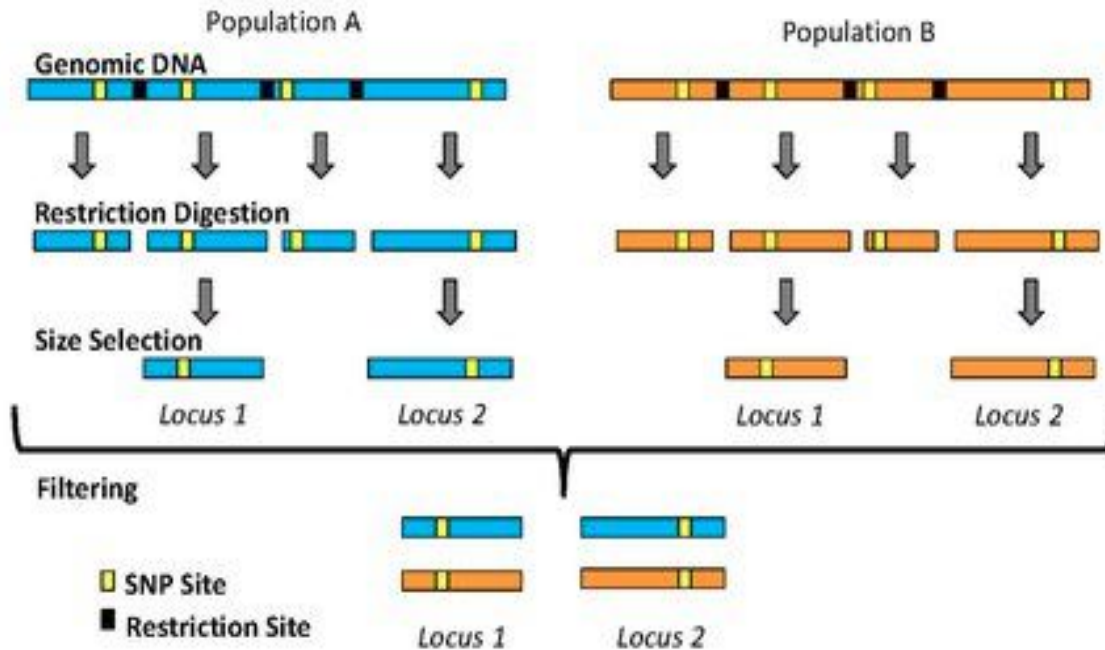
Source of your data

**REDUCED REPRESENTATION (RADseq, GBS)**

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### REDUCED REPRESENTATION (RADseq, GBS)



After digestion, sequencing and mapping, this is your data!

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#### Pros:

- The cheapest of the methods
- Not dependent upon a reference genome
- Samples fixed in ethanol OK
- Markers distributed evenly across the genome

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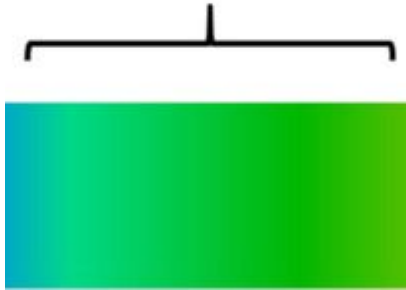
- No full genes, only SNPs
- Only for population genomics or phylogeny including closely-related species
- Missing data as a product of the transcriptome representing a snapshot of expression (but this could also affect genome annotation)
- No proper orthology inference

# 01 DATA

## Source of your data

### METAGENOMICS/METATRANSCRIPTOMICS

Isolate Genome  
(bulk)

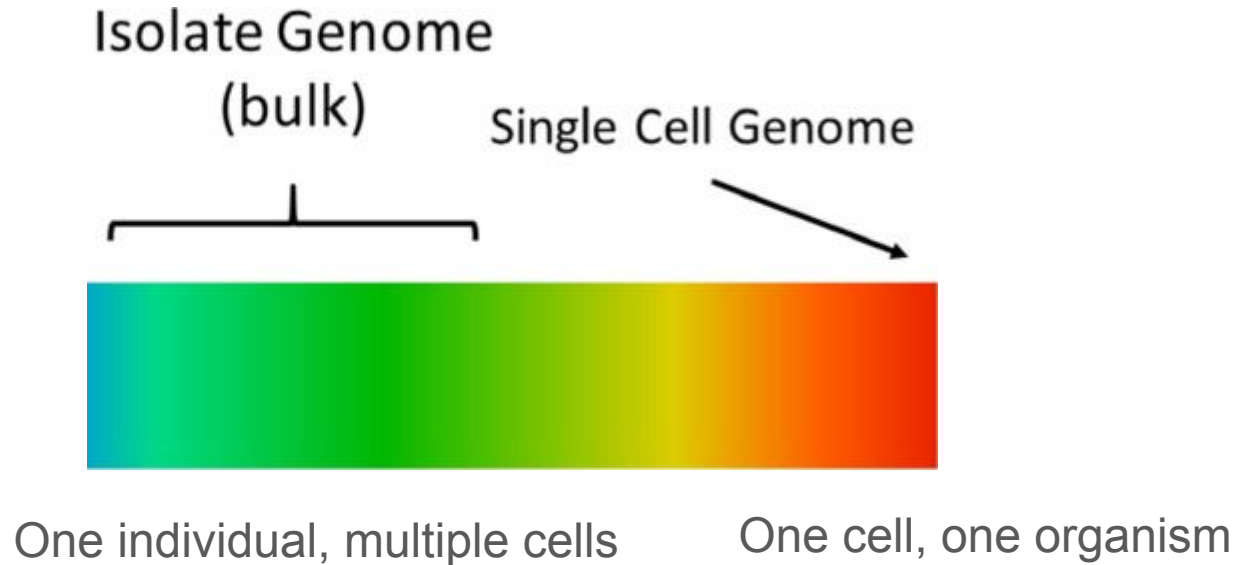


One individual, multiple cells

# 01 DATA

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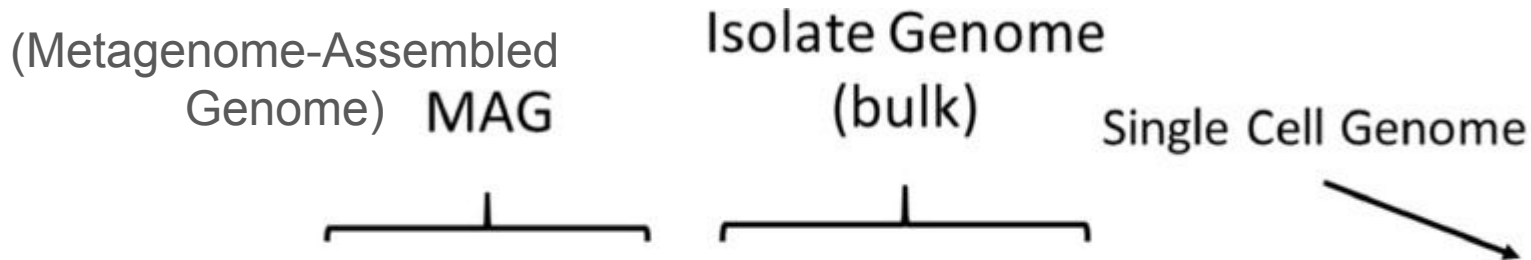
### METAGENOMICS/METATRANSCRIPTOMICS



# 01 DATA

## Source of your data

### METAGENOMICS/METATRANSCRIPTOMICS



One cell, multiple organisms

One individual, multiple cells

One cell, one organism

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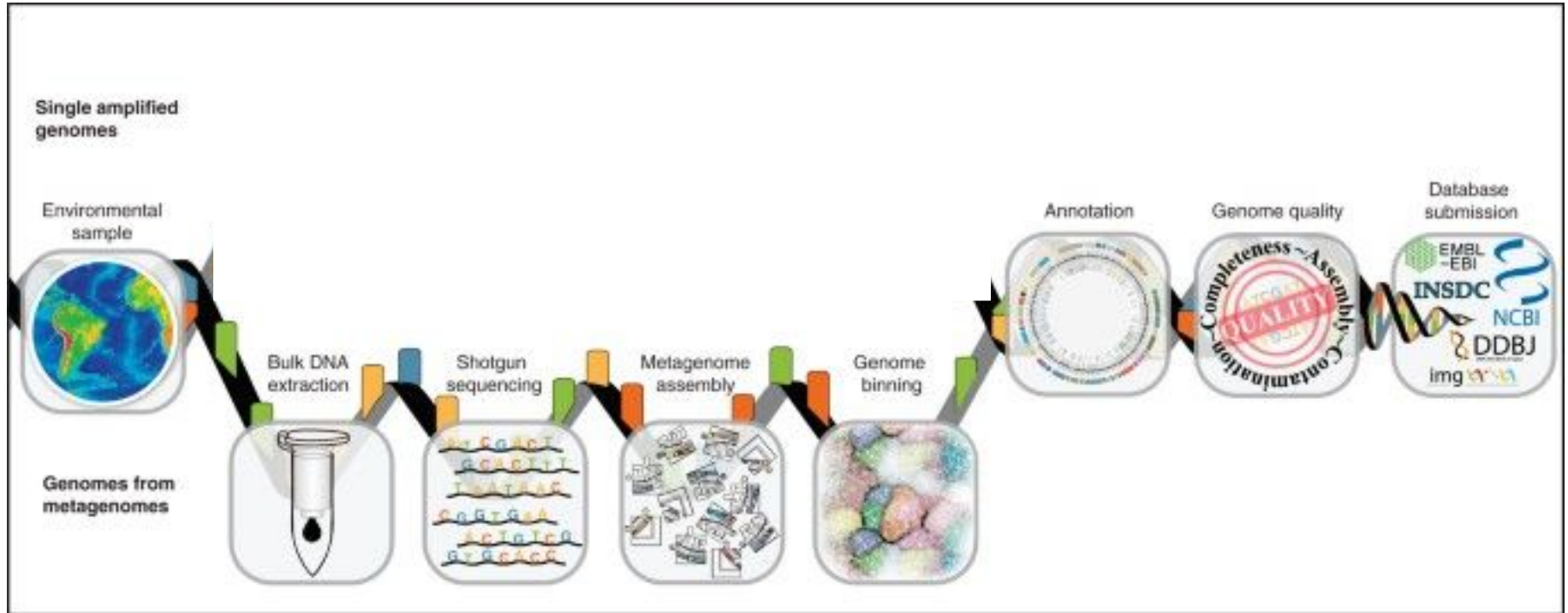
Source of your data

**METAGENOMICS - single cell**

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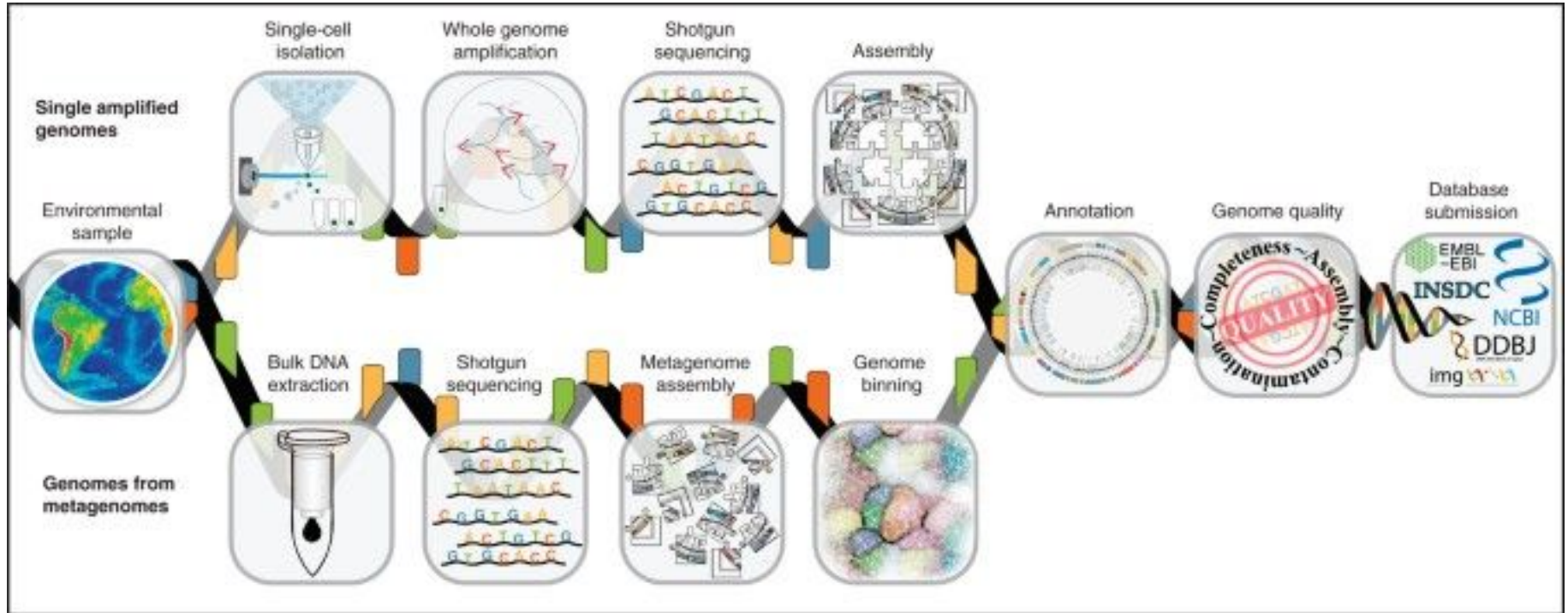
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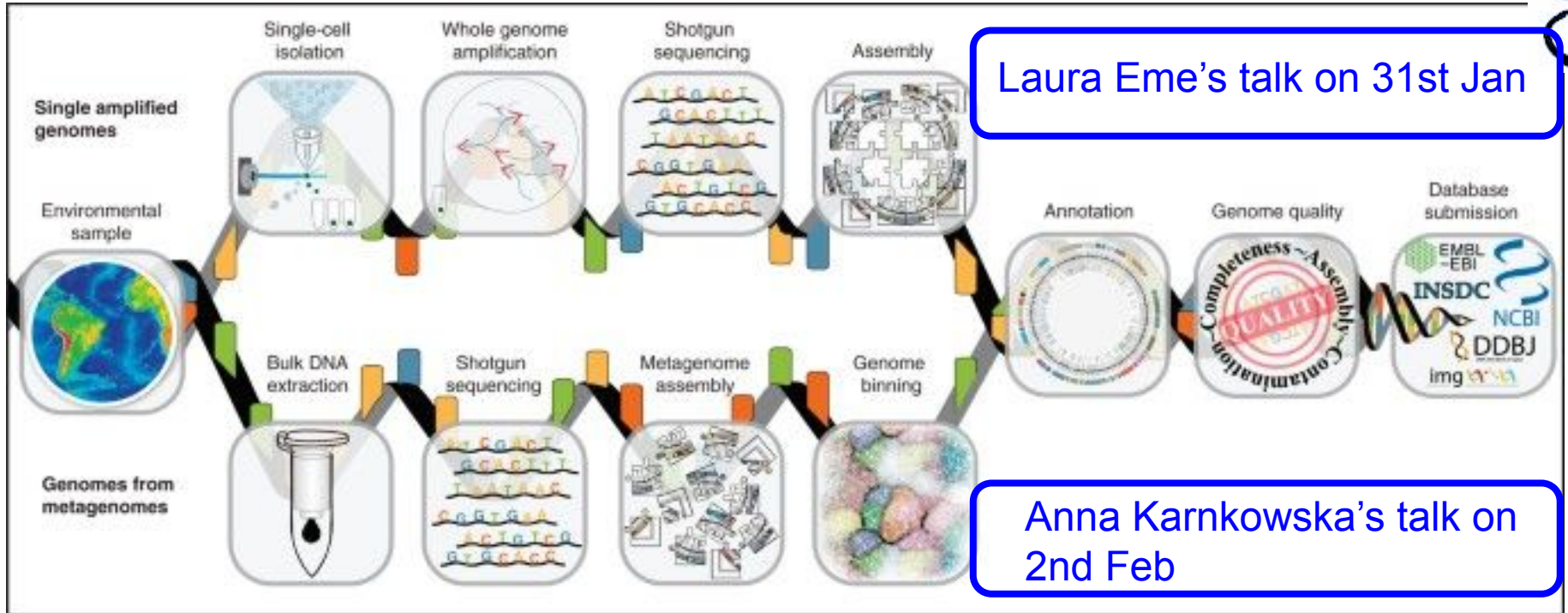
### METAGENOMICS - single cell vs MAGs



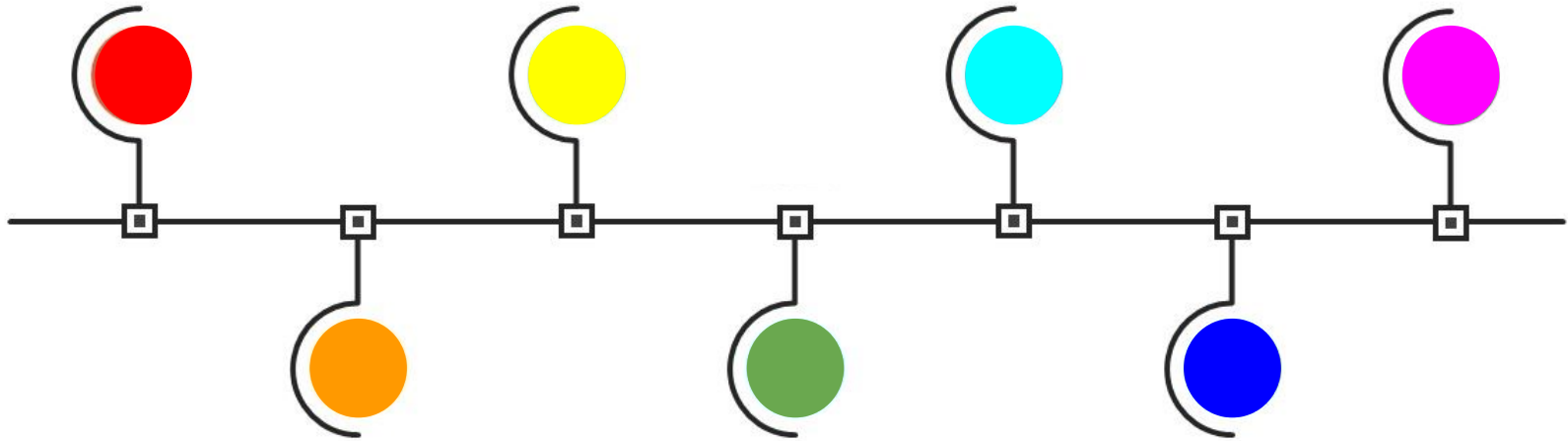
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## 01 DATA



**02 ORTHOLOGY  
INFERENCE**

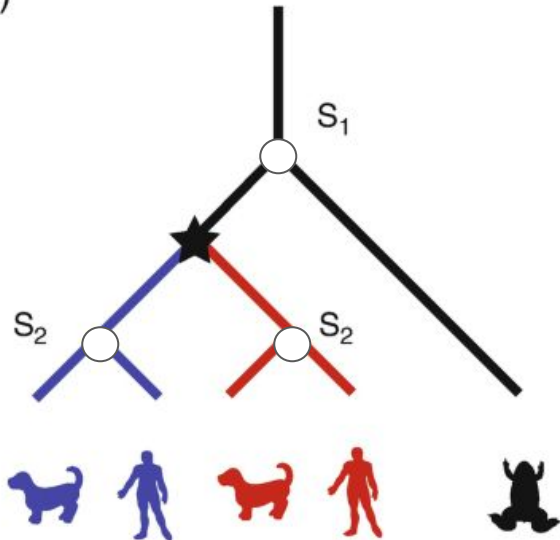
## 02 ORTHOLOGY INFERENCE

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## Definitions

- Two genes are **orthologs** if their MRCA is a **speciation**:  $\circ$

a)

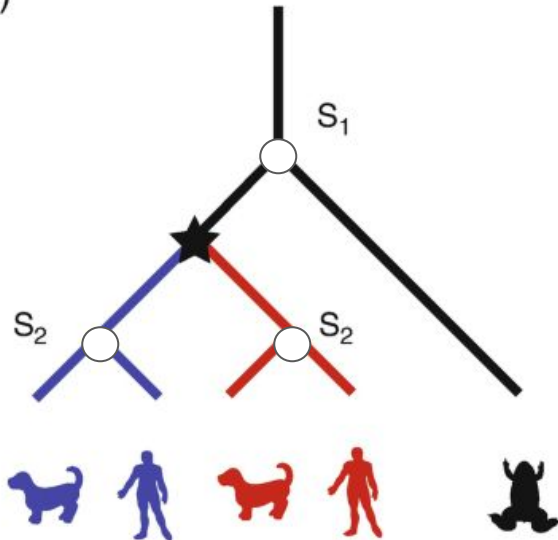


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- Two genes are **orthologs** if their MRCA is a *speciation*: ○
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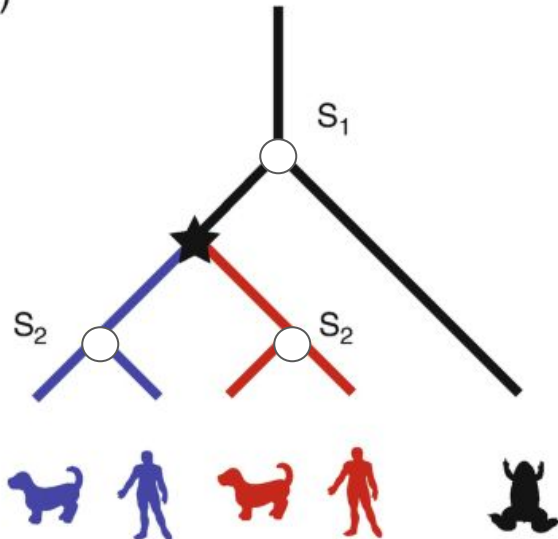
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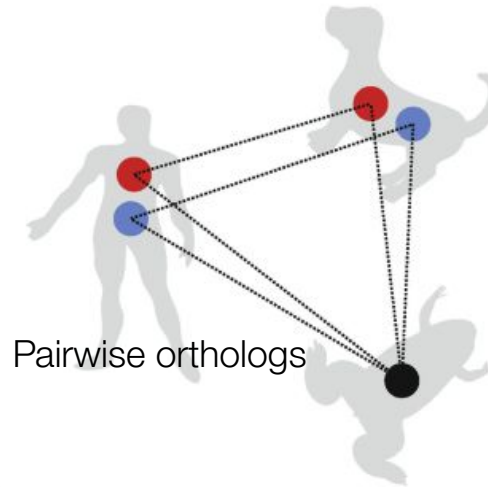
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Orthology relationships are inferred *pairwise*

a)



b)

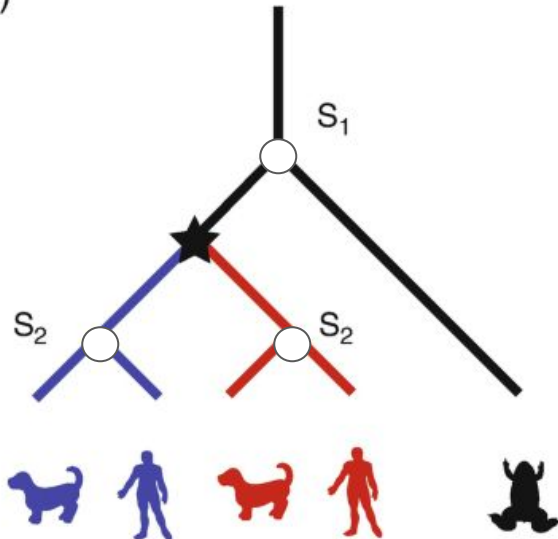


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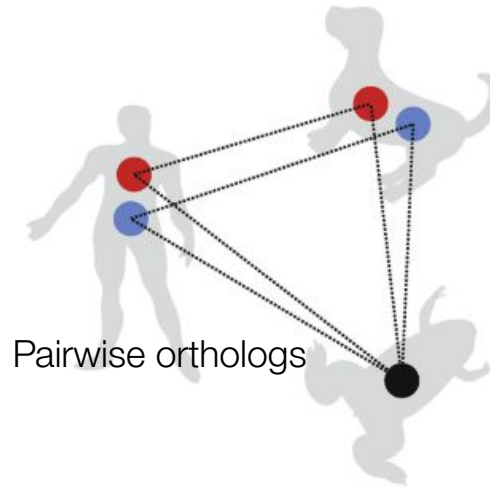
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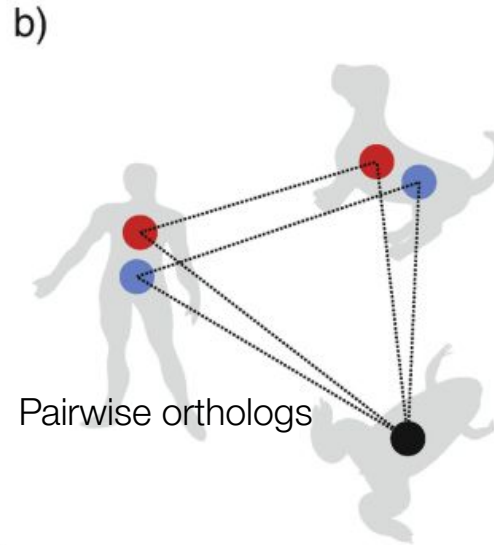
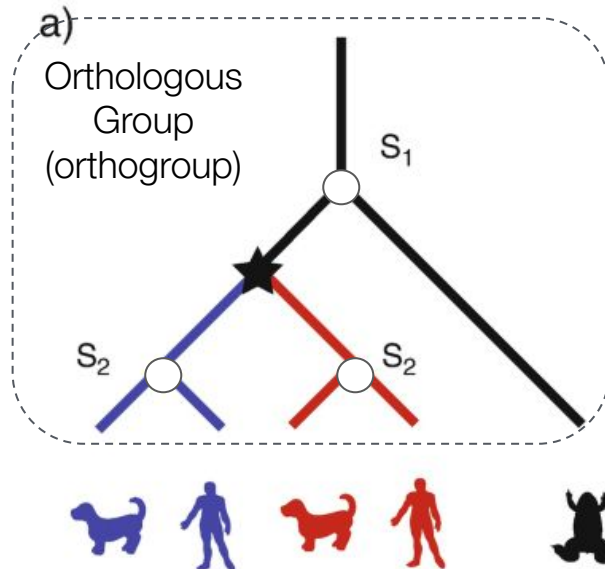
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When we have multiple species, we should consider the concept of *orthogroup*

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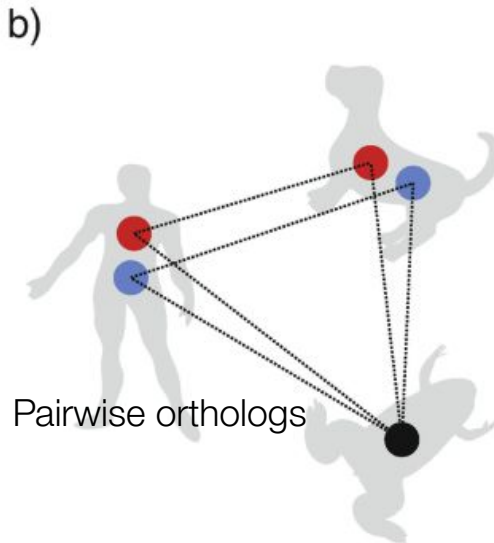
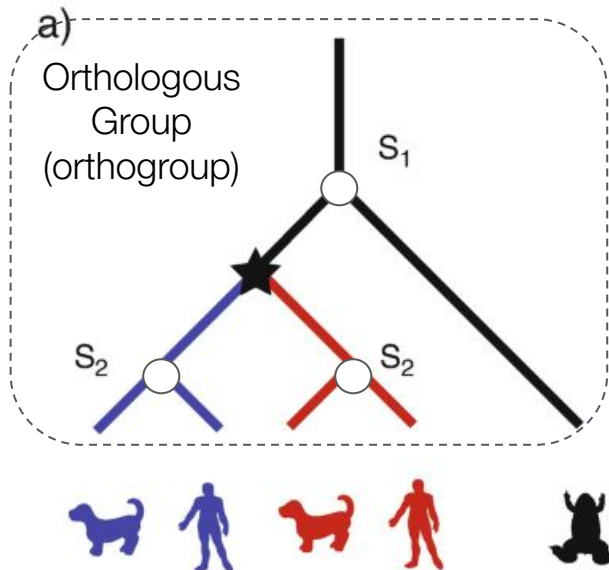
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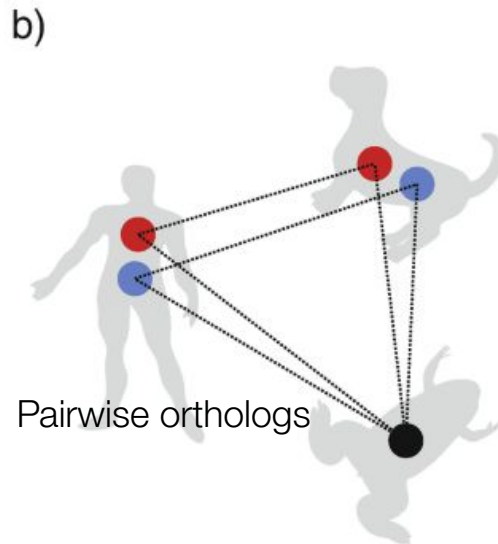
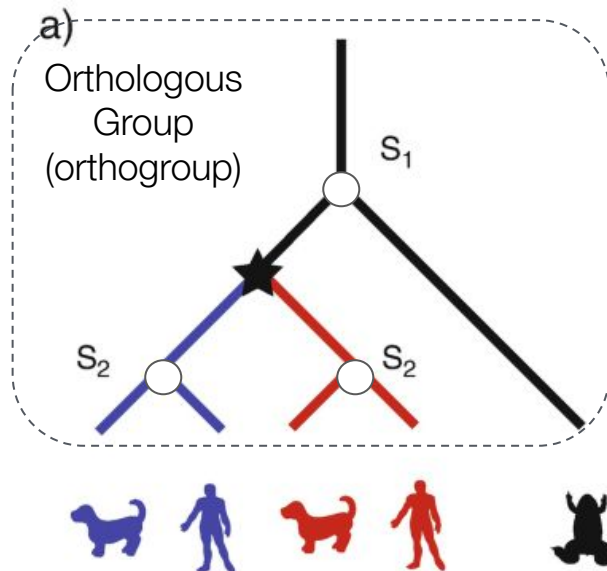
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Orthology inference is essential for phylogenomics, as you want to consider only genes that arise through speciation events

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For phylogenomic inference, we want either:

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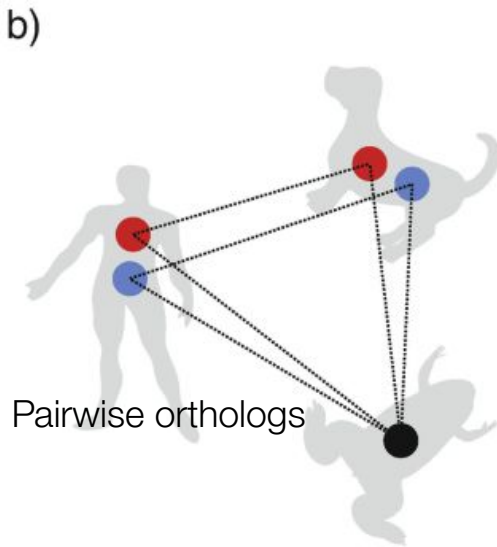
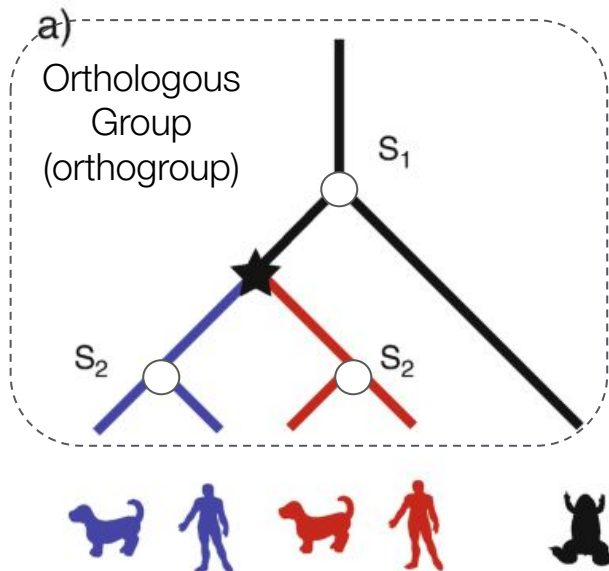
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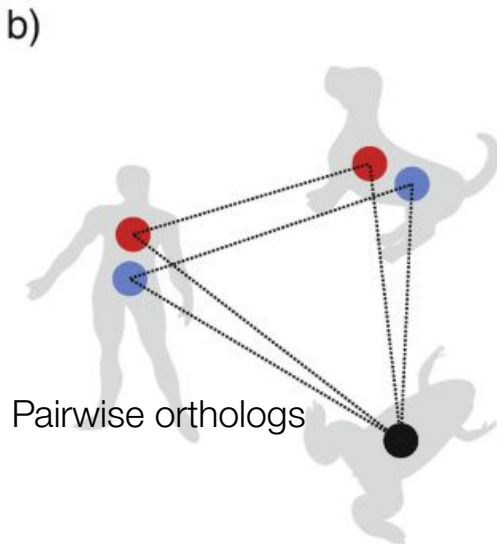
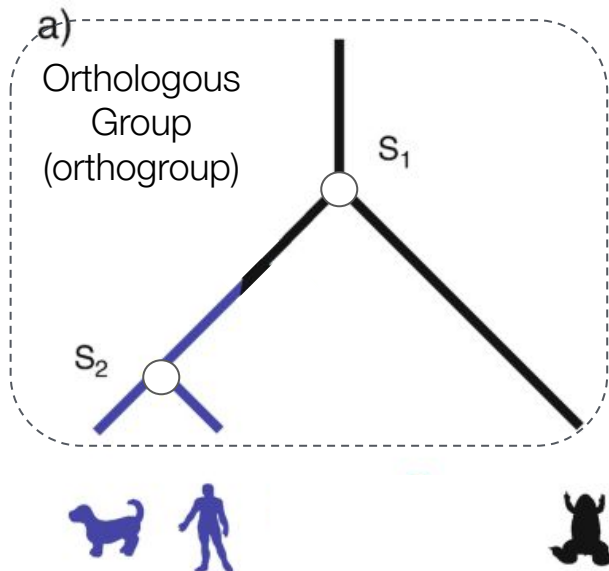
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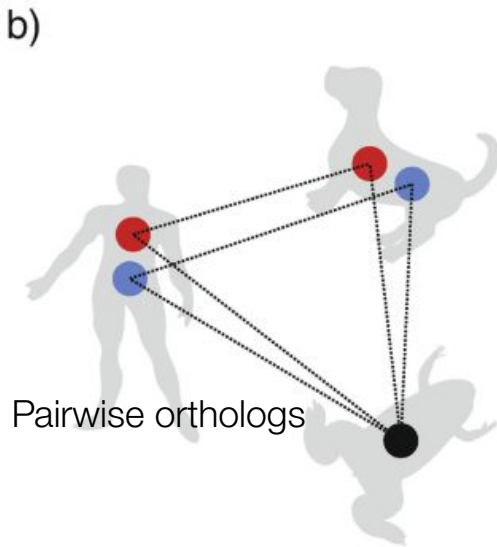
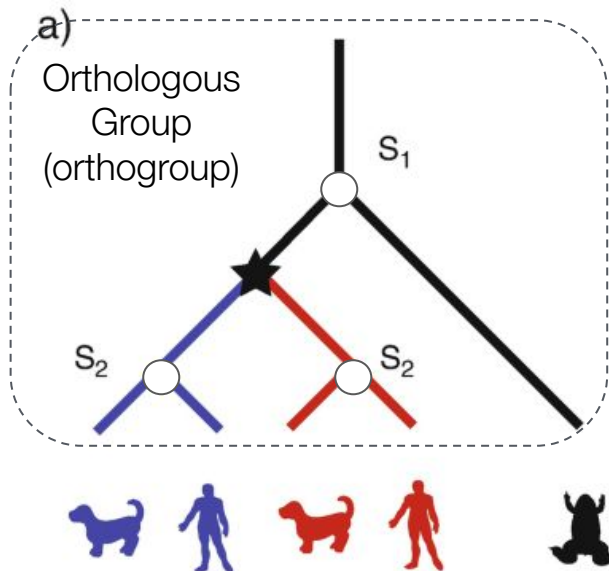
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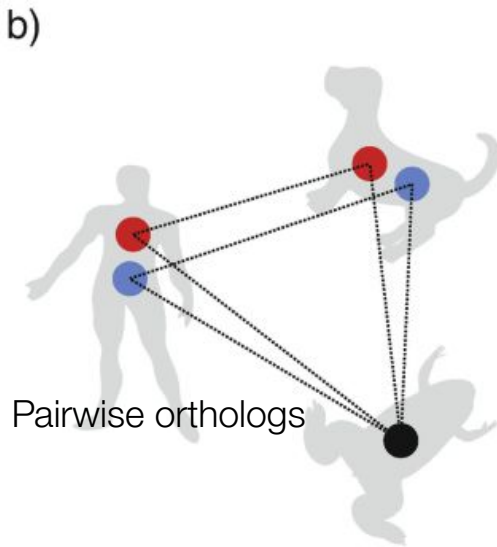
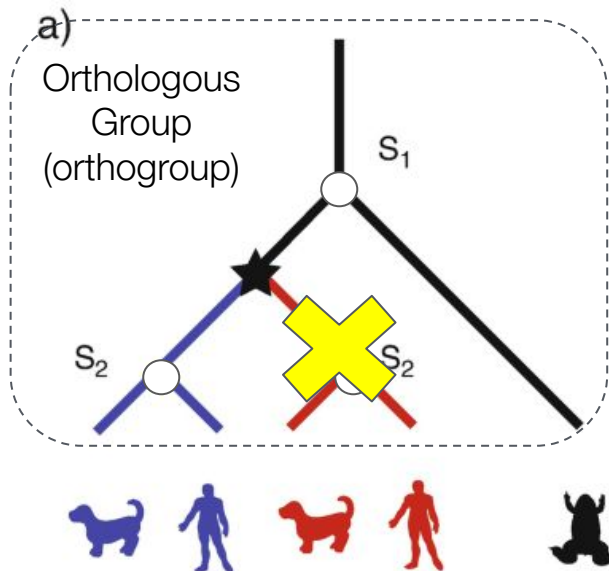
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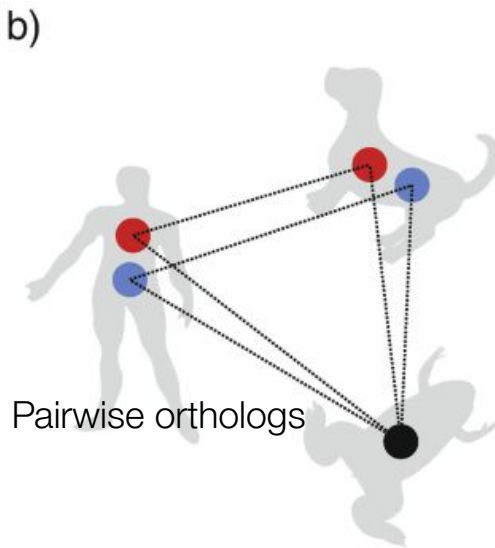
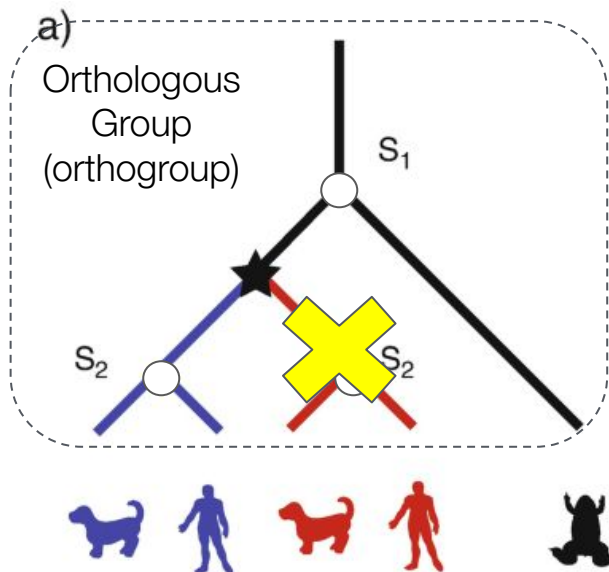
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Marina Marcet-Houben tomorrow

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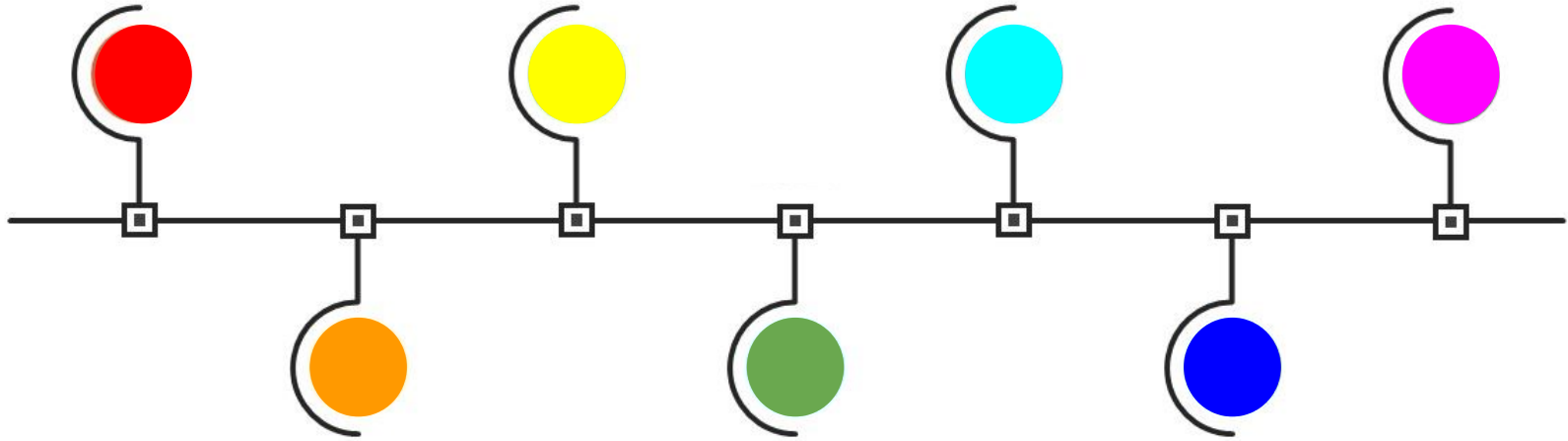
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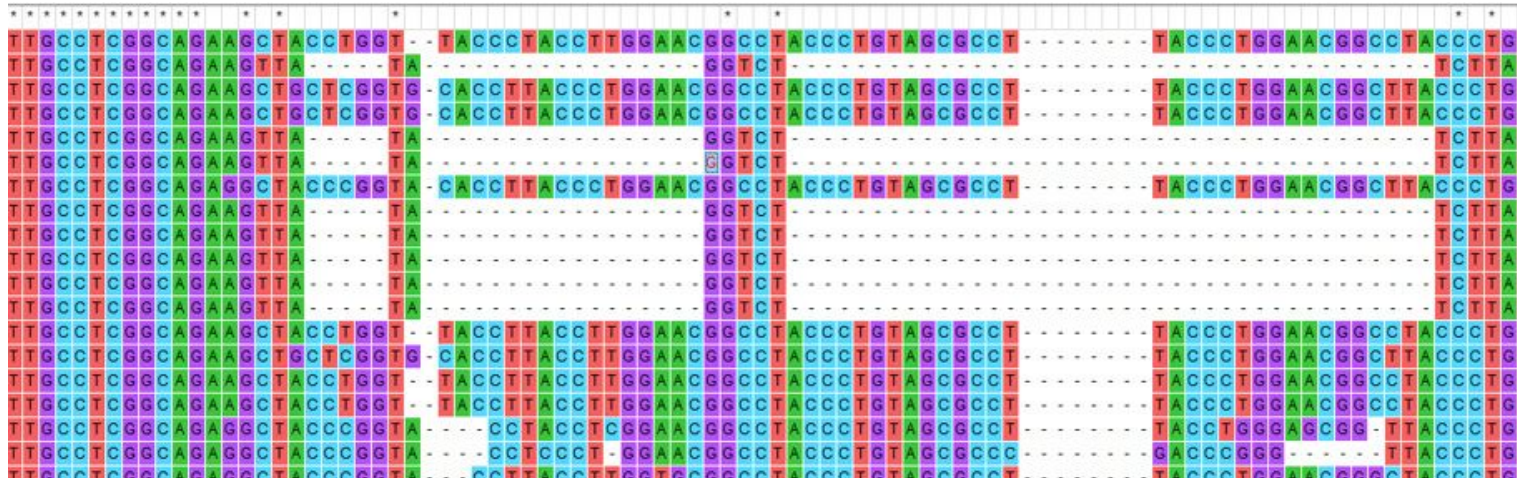
**03 ALIGNMENT  
& TRIMMING**

**02 ORTHOLOGY  
INFERENCE**



# 03 ALIGNMENT AND TRIMMING

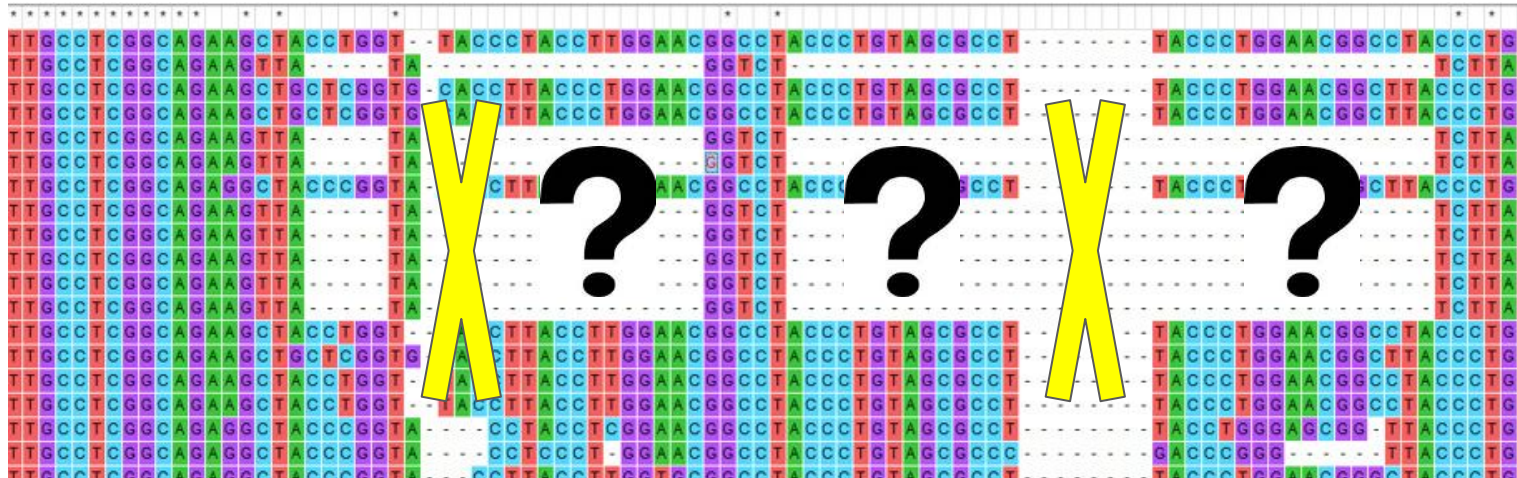
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If the sequences are poorly aligned, you may want to consider trimming the poorly aligned areas.



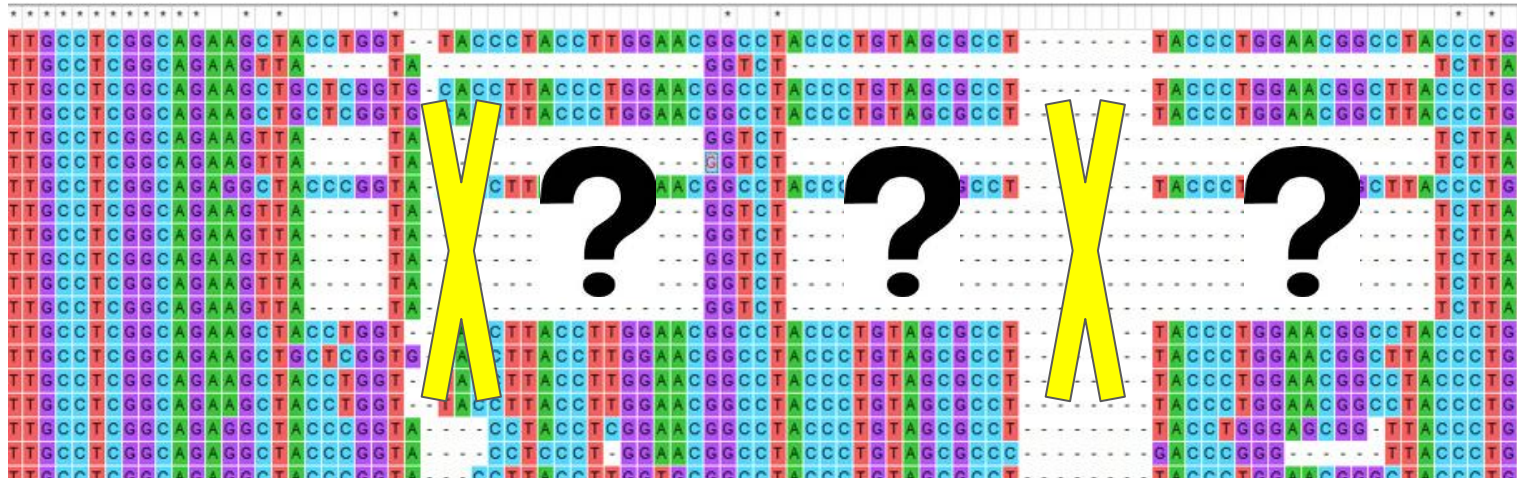
# 03 ALIGNMENT AND TRIMMING

Jacob and Marina today



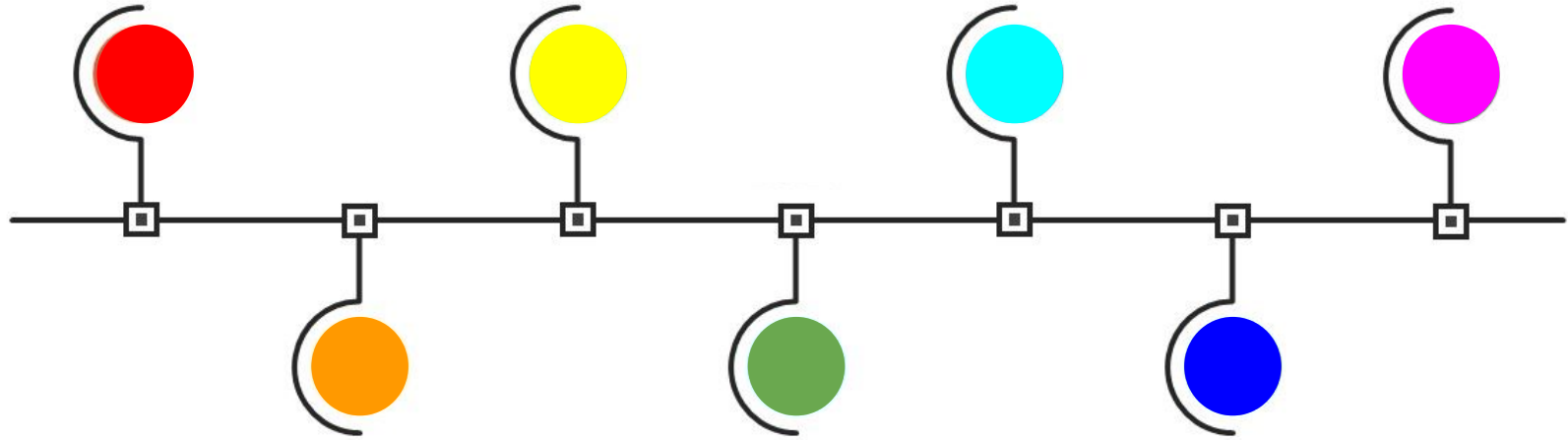
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**04 PHYLOGENOMIC  
SUBSAMPLING**

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**Why?** To avoid an accumulation of nonphylogenetic signals as a product of heterogeneities in evolutionary processes, reduce computing time and improve model fit.

This step can be used to *explore phylogenetic conflicts*, *test specific hypotheses* of relationships, measure the impact of *different sources of bias*, and allow for a *better modeling* of evolutionary processes.

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**How?** By checking the properties of genes or sites and selecting the ones that minimize bias.

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Which properties?

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## Information content

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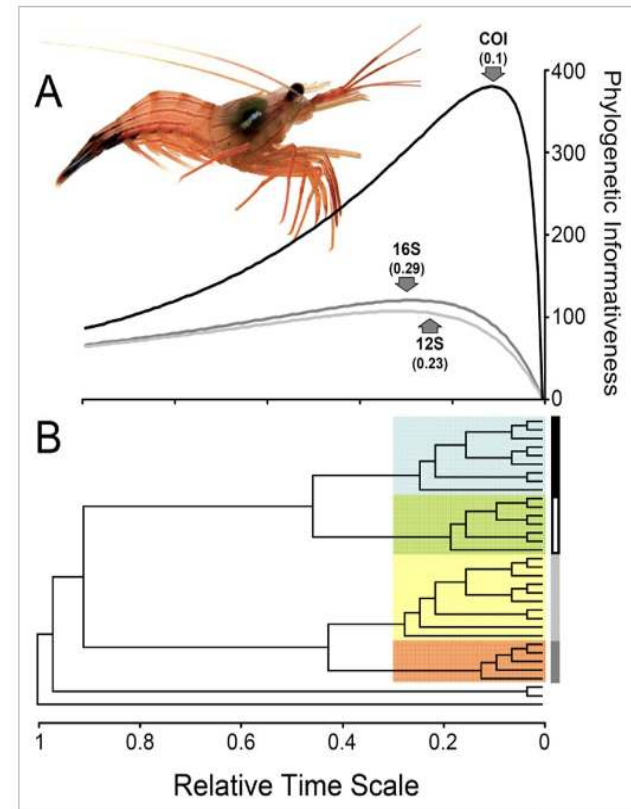
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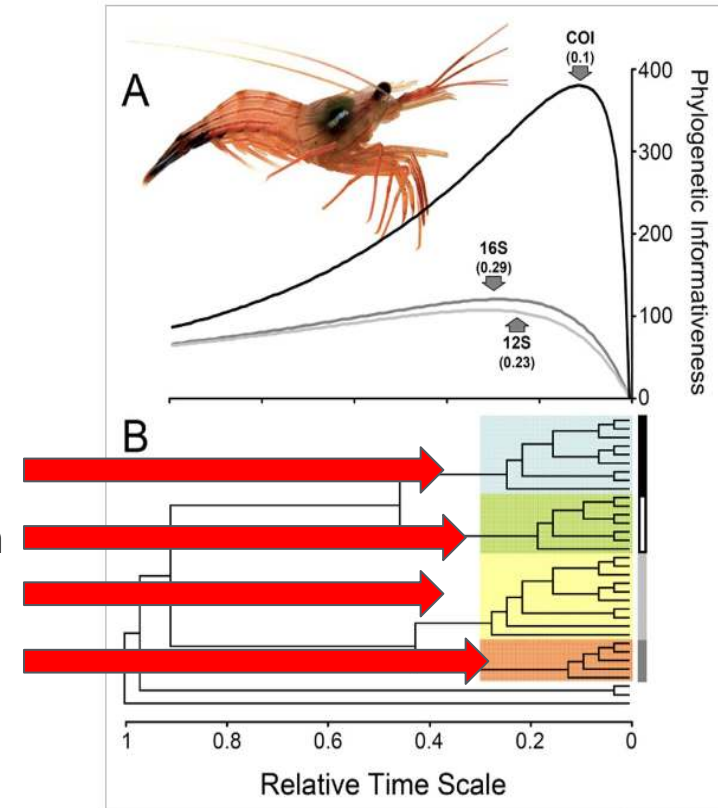
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## Phylogenetic signal

Good information  
to infer these  
nodes



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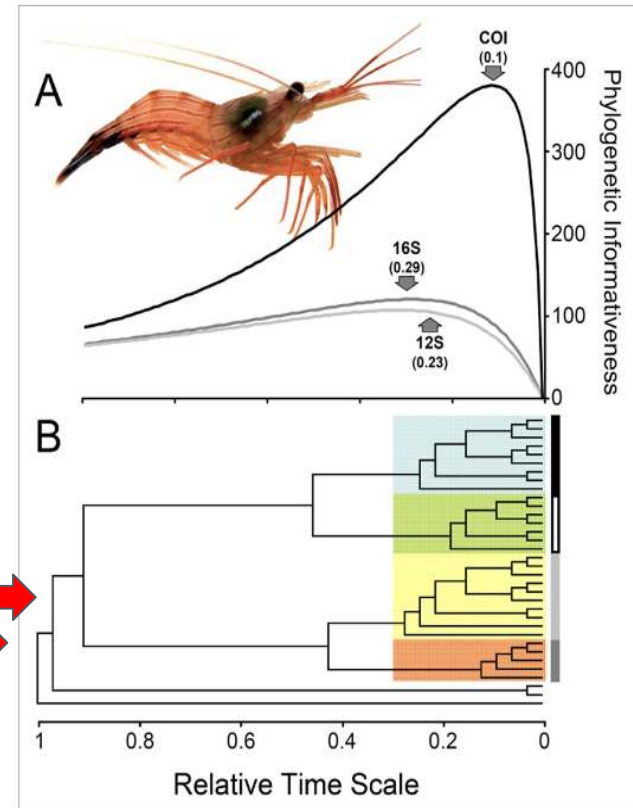
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Not enough  
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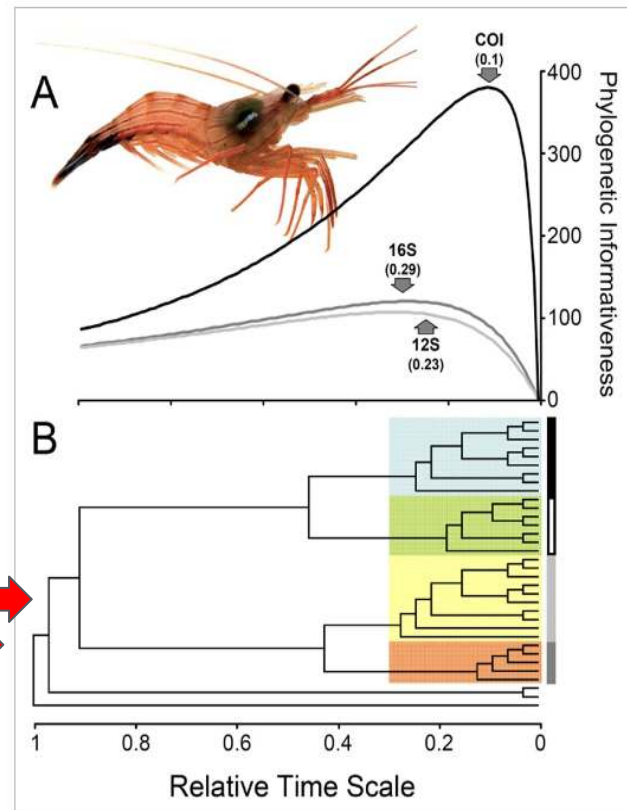
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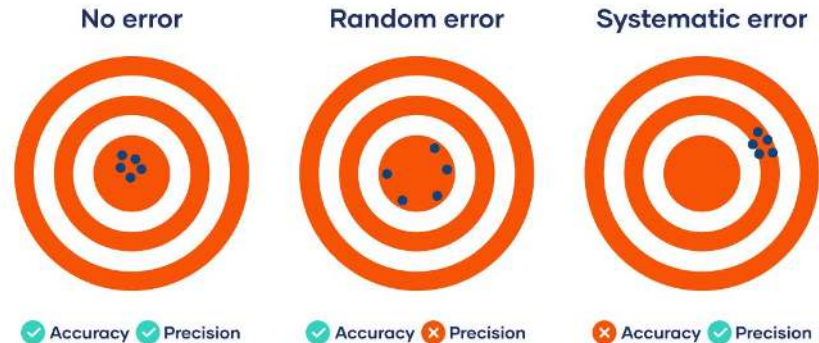
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**Systematic error:** when a calculated value deviates from the true value in a consistent way.

### Random vs. systematic error



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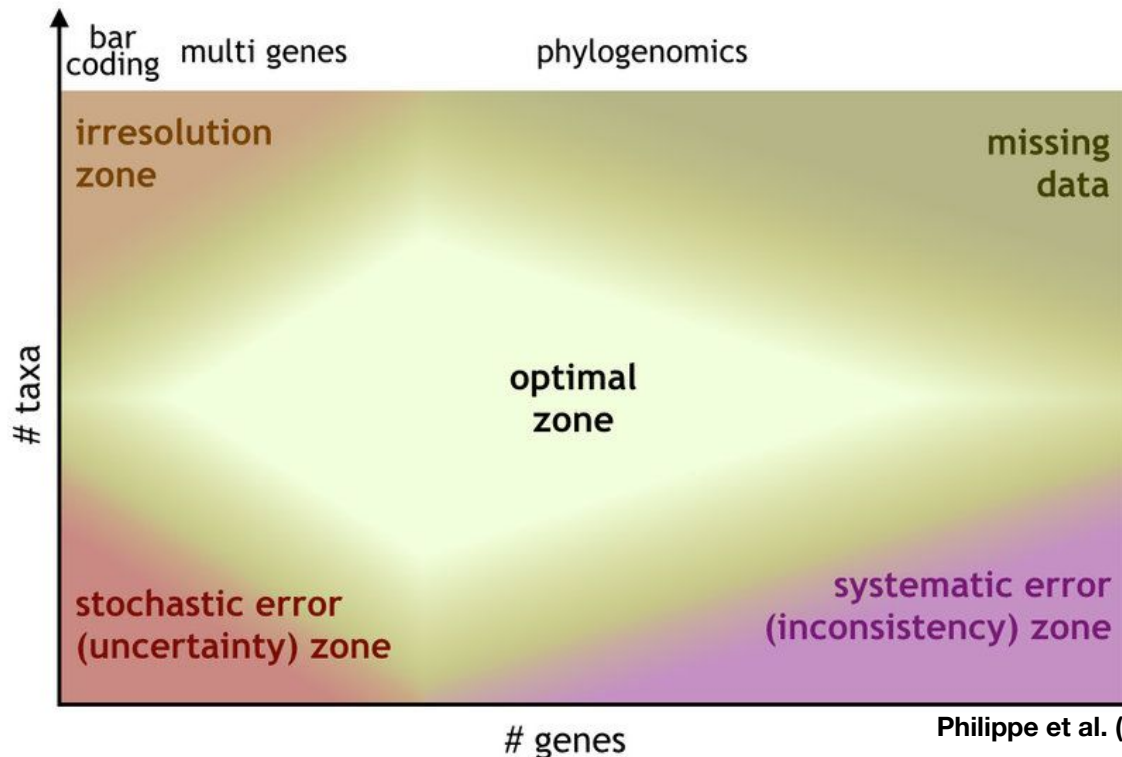
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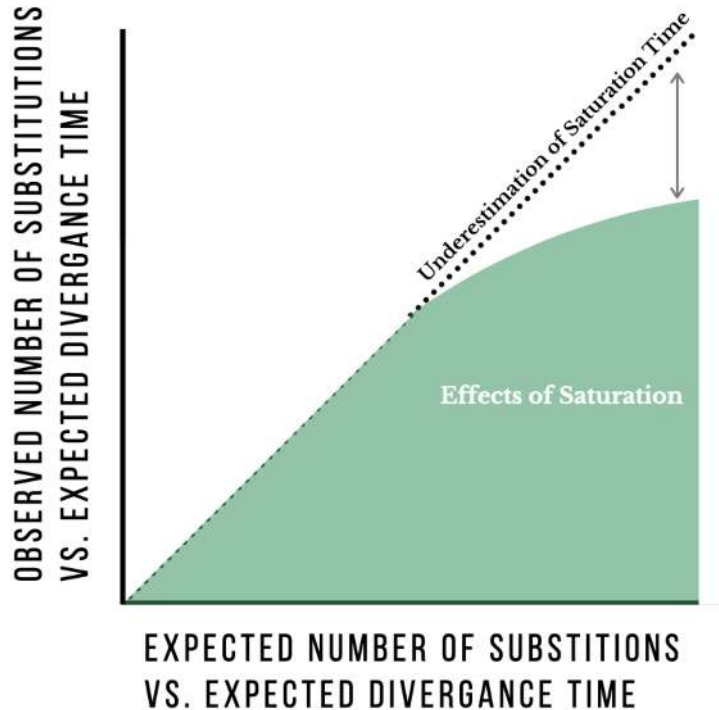
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|-----------|--------|--------|-----------------|-----|
|           | Site 1 | Site 2 | Site 3...Site n |     |
| Species A | Leu    | Met    | Lys             | Hys |
| Species B | Leu    | Leu    | Asn             | Pro |
| Species C | Leu    | Met    | Lys             | Pro |
| Species D | Leu    | Ile    | Leu             | Leu |

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# 04 PHYLOGENOMIC SUBSAMPLING

## Which properties?

### Information content

- > length of alignment
- > missing data
- > level of occupancy

### Phylogenetic signal

- > average support
- > Robinson-Foulds distance

### Systematic error

- > root-to-tip distance (ie, the degree of deviation from a strict clock-like behavior)
- > average pair-wise patristic distance between terminals (indicative of susceptibility to long-branch attraction)
- > level of saturation
- > compositional heterogeneity

Jacob and Marina today

Antonis Rokas and Jacob on 29th Jan



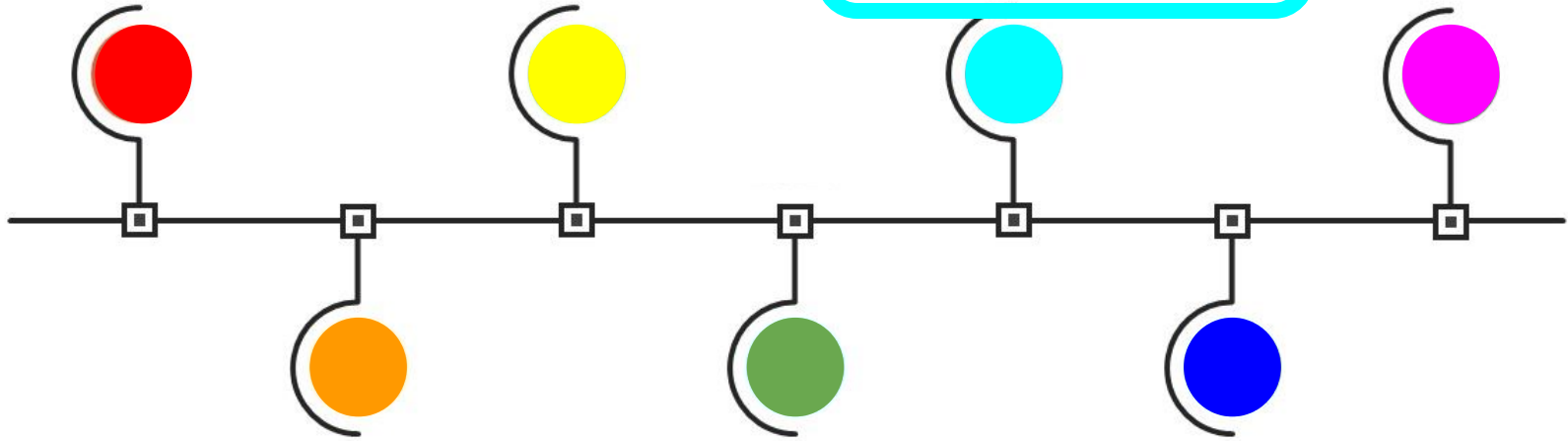
**01 DATA**

**03 ALIGNMENT  
& TRIMMING**

**05 SUPERMATRIX  
VS INDIVIDUAL  
GENES**

**02 ORTHOLOGY  
INFERENCE**

**04 PHYLOGENOMIC  
SUBSAMPLING**



## 05 SUPERMATRIX VS INDIV. GENE TREES

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~~Gene tree  $\approx$  Species phylogeny~~

**Gene tree  $\neq$  Species phylogeny**

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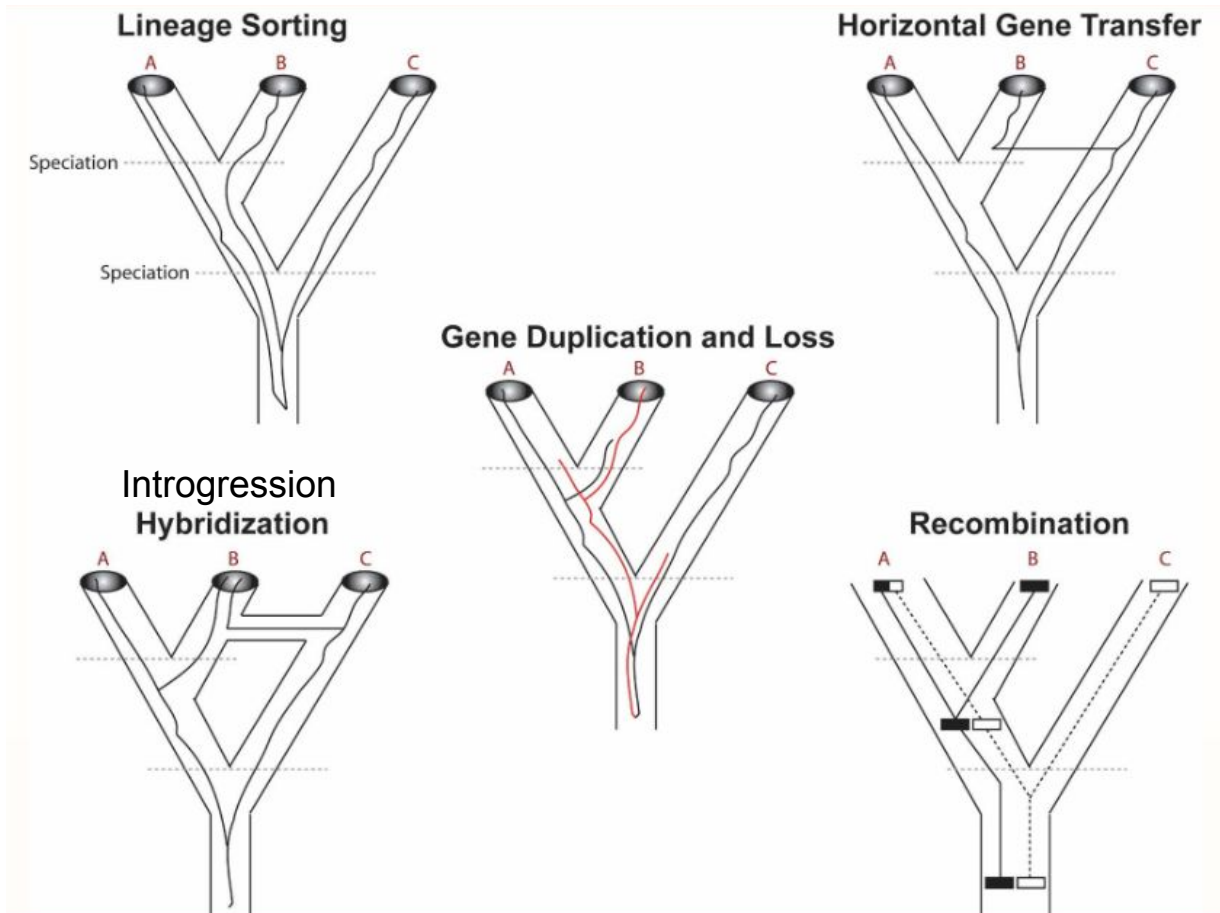
### Analytical factors

They lead to failure in accurately inferring a gene tree; these can be either due to **stochastic error** (e.g., insufficient sequence length or taxon samples) or due to **systematic error** (e.g., observed data far depart from model assumptions)

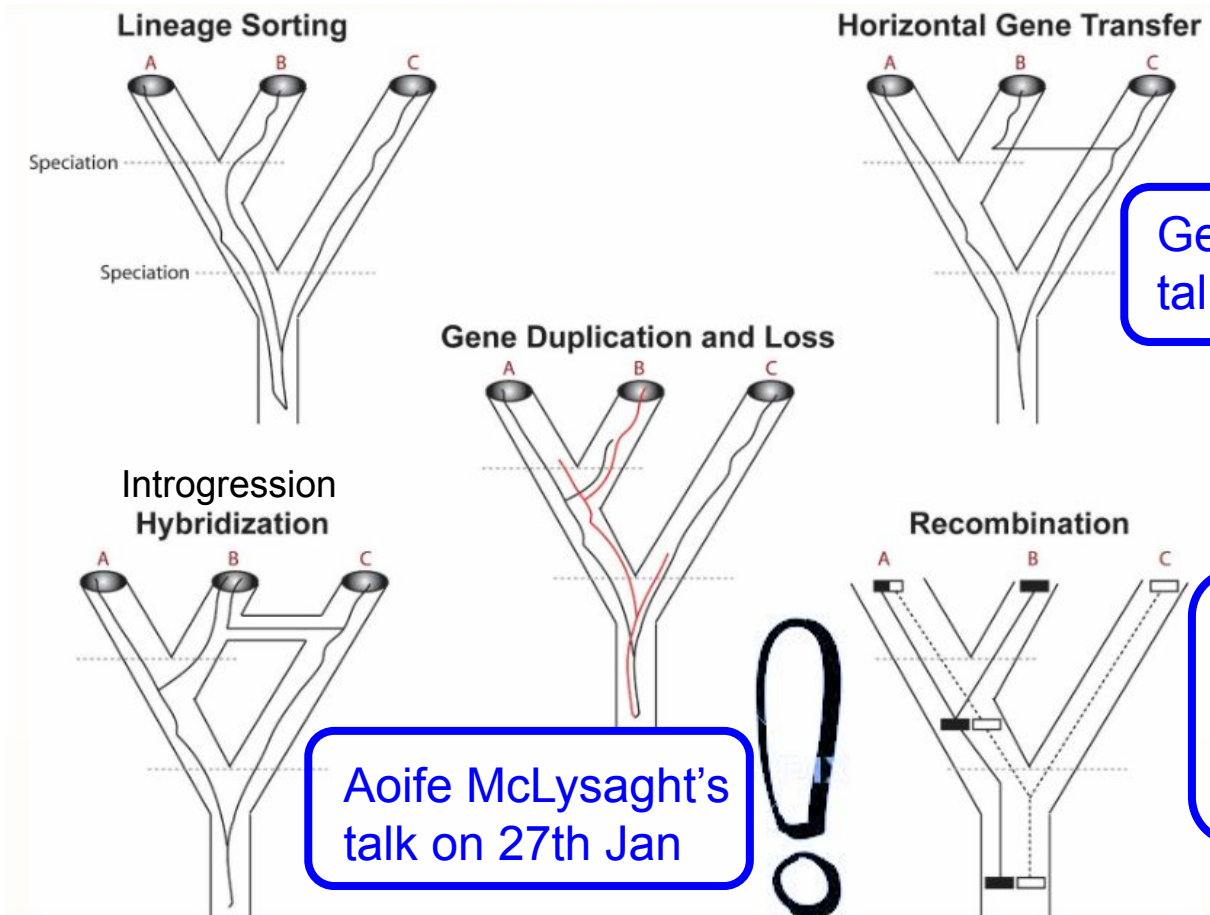
### Biological factors

They lead to gene trees that are topologically distinct from each other and from the species tree. Known factors include **stochastic lineage sorting**, **hidden paralogy**, **horizontal gene transfer**, **recombination** and **natural selection**

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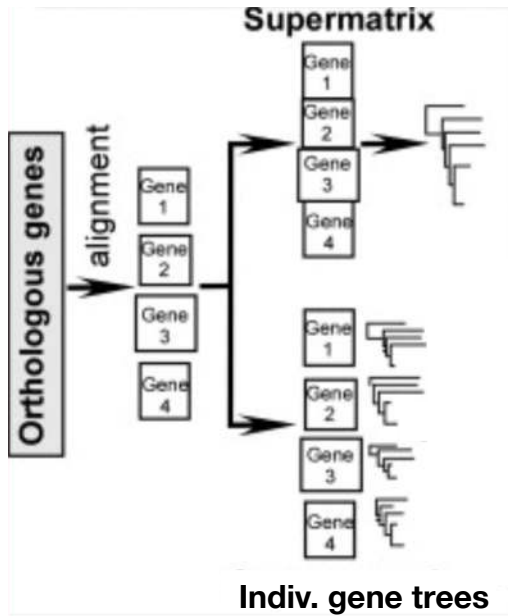


Gergely Szöllősi's  
talk on 1st Feb

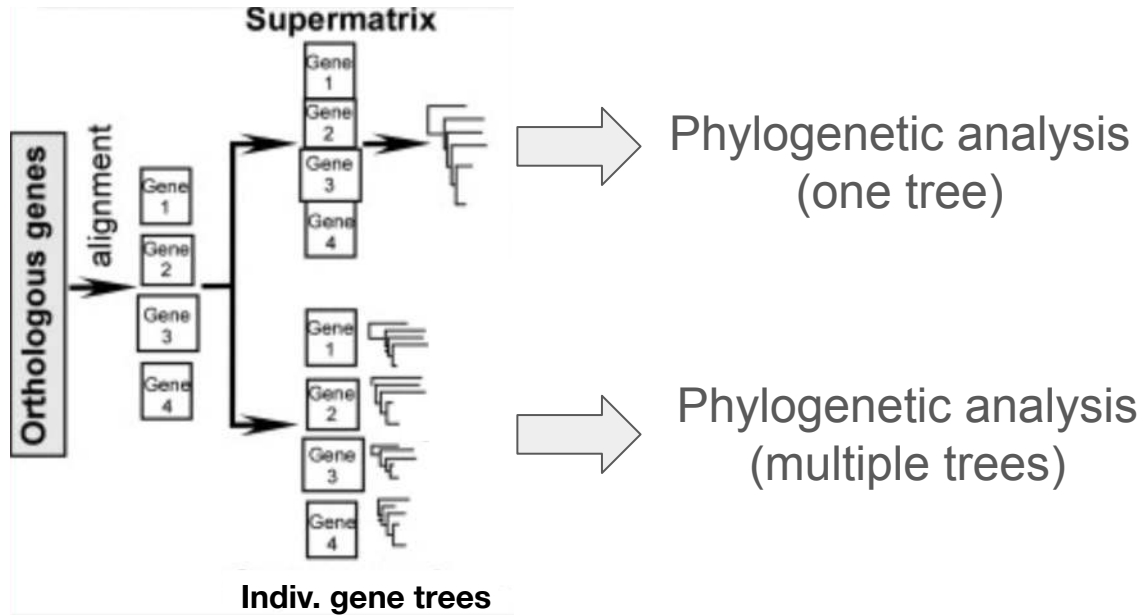
Aoife McLysaght's  
talk on 27th Jan

Toni  
Gabaldón's talk  
on 30th Jan

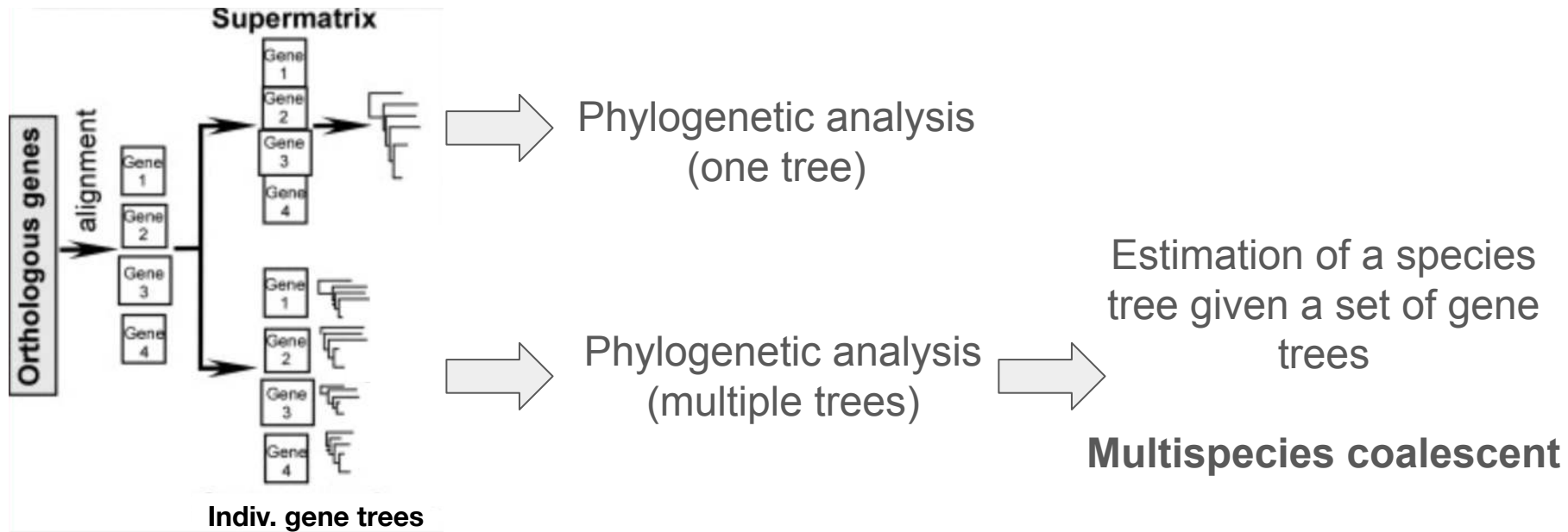
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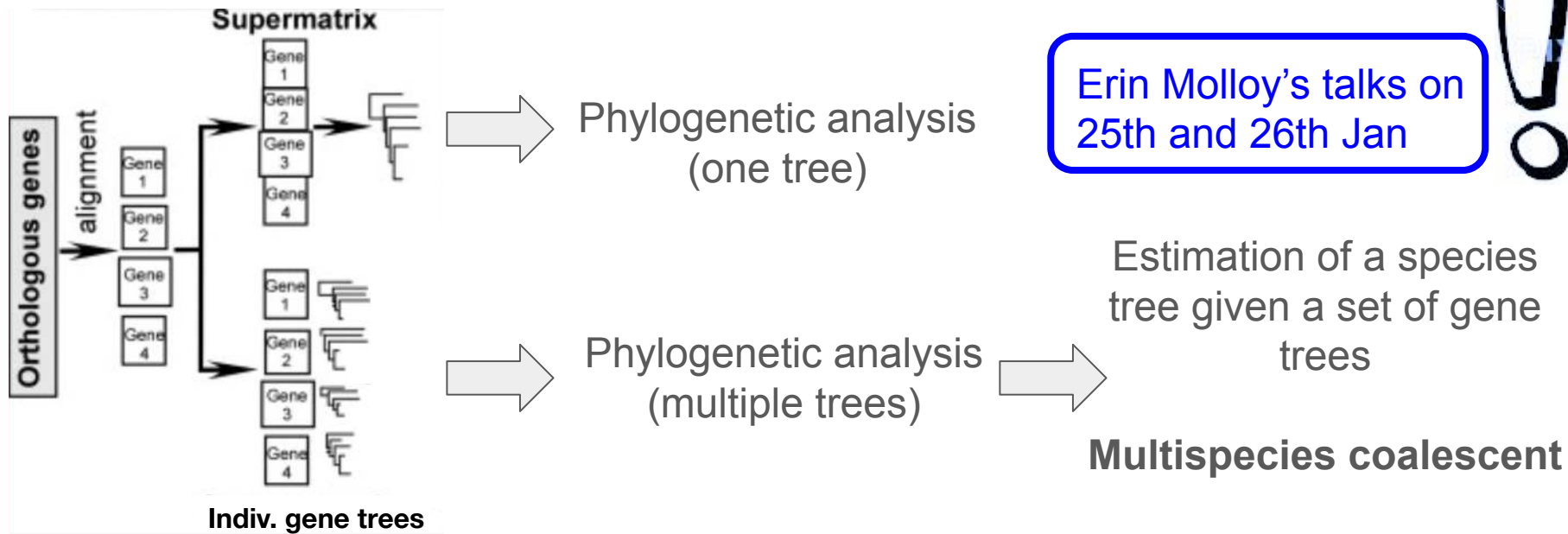
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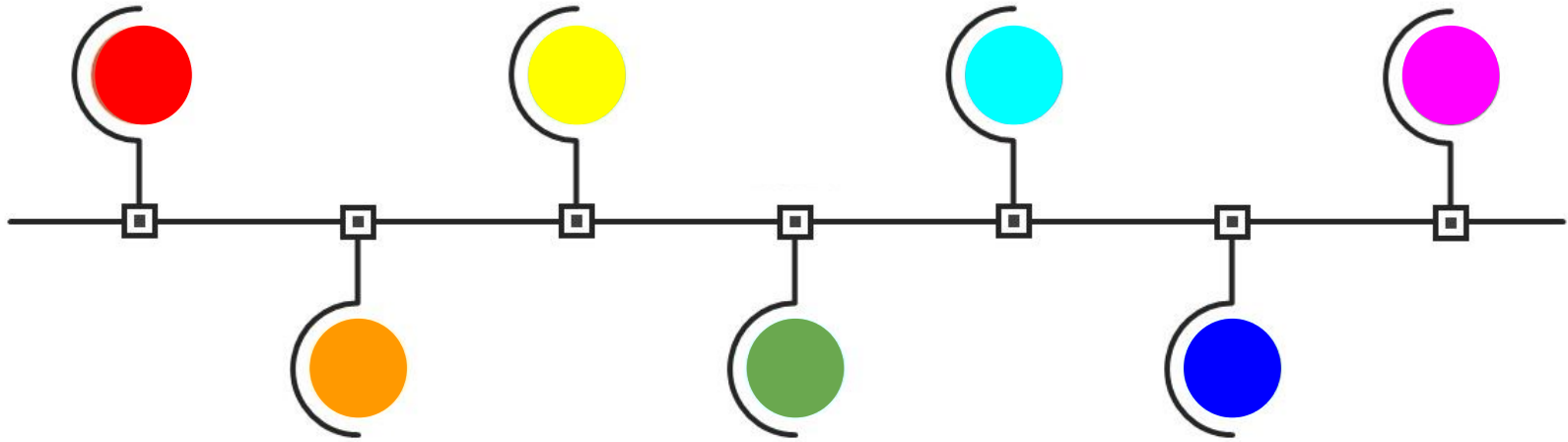
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# 06 MODEL SELECTION & PHYLOGENETIC INFERENCE

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**DATA**

# 06 MODEL SELECTION & PHYLOGENETIC INFERENCE



**DATA + MODEL OF EVOLUTION**

# 06 MODEL SELECTION & PHYLOGENETIC INFERENCE



**DATA + MODEL OF EVOLUTION  
+ METHOD**

# 06 MODEL SELECTION & PHYLOGENETIC INFERENCE



**DATA + MODEL OF EVOLUTION**

**+ METHOD**

**+ A WAY TO ASSESS HOW GOOD YOUR HYPOTHESIS IS**

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**DATA** + **MODEL OF EVOLUTION**

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**DATA** + **MODEL OF EVOLUTION** (= substitution model)

A model that describes changes in sequences over evolutionary time and transforms the number of changes in an evolutionary distance

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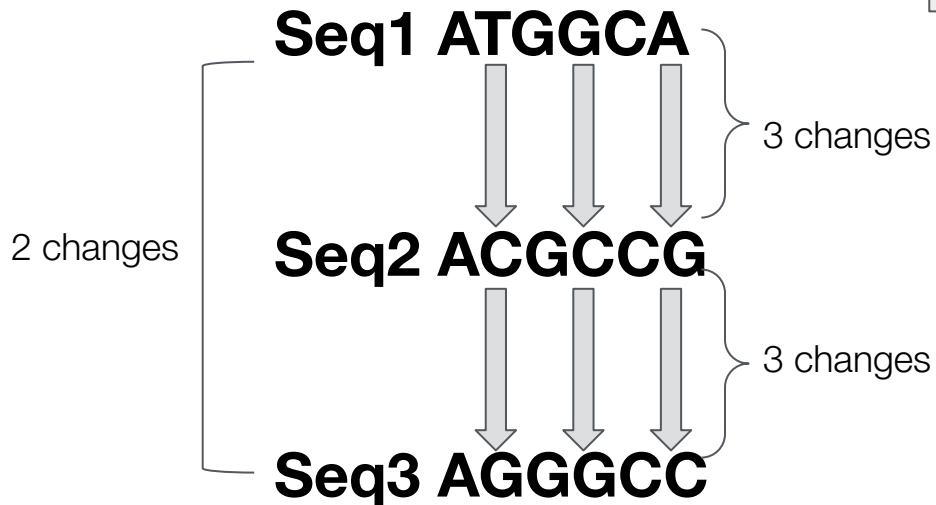
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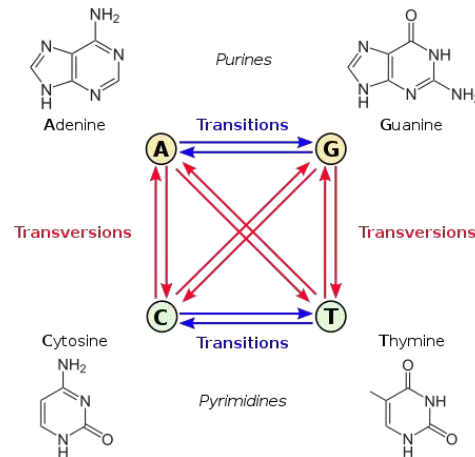
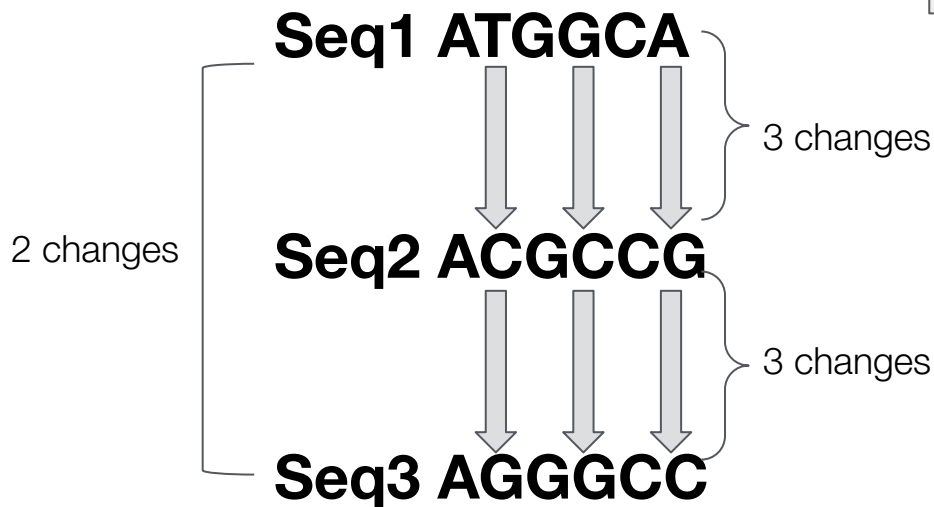
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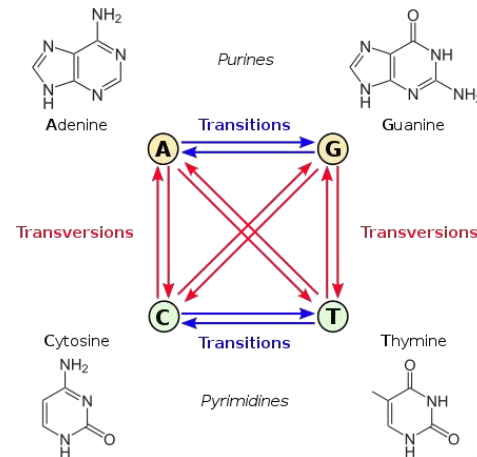
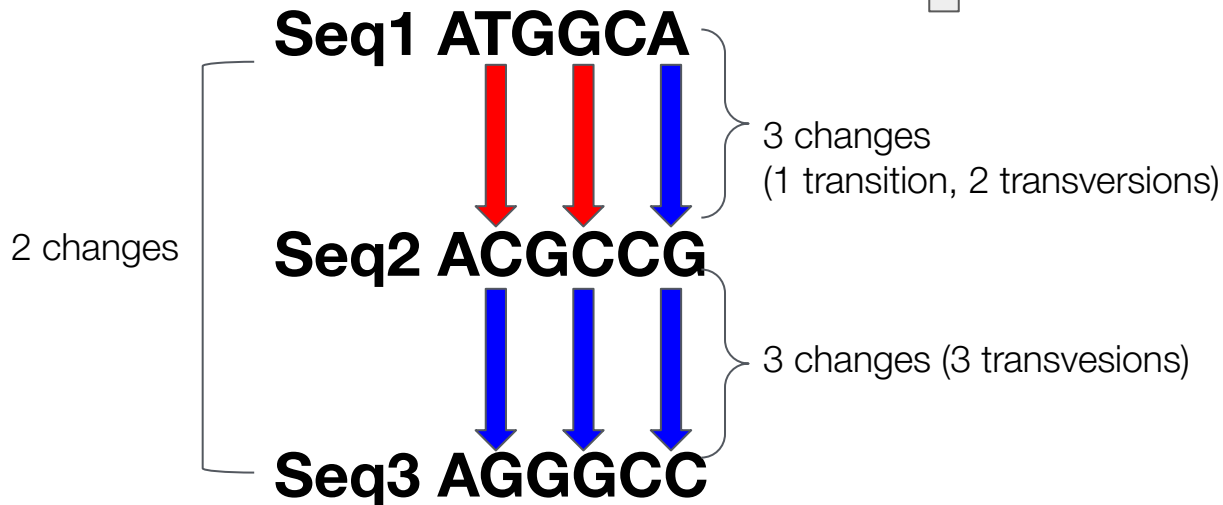
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**Seq2 ACGCCG**

**Seq3 AGGGCC**

3 changes

3 changes

2 changes

Complexity



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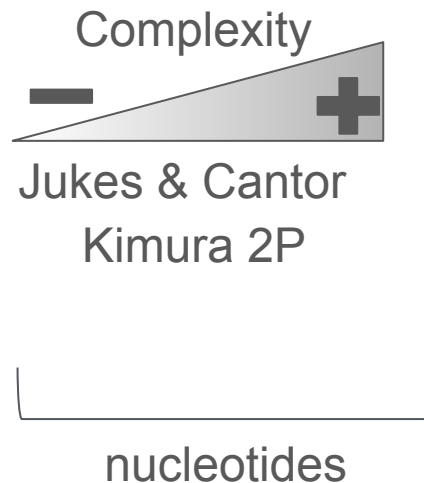
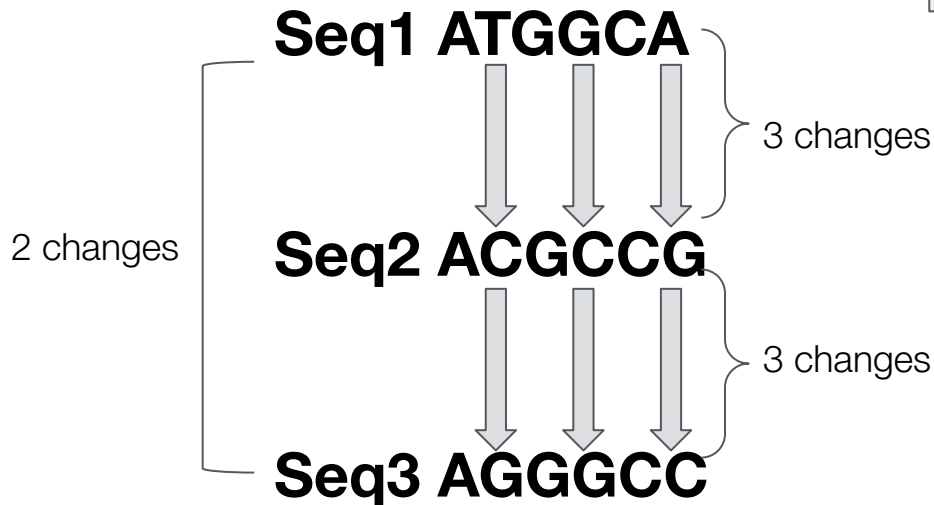
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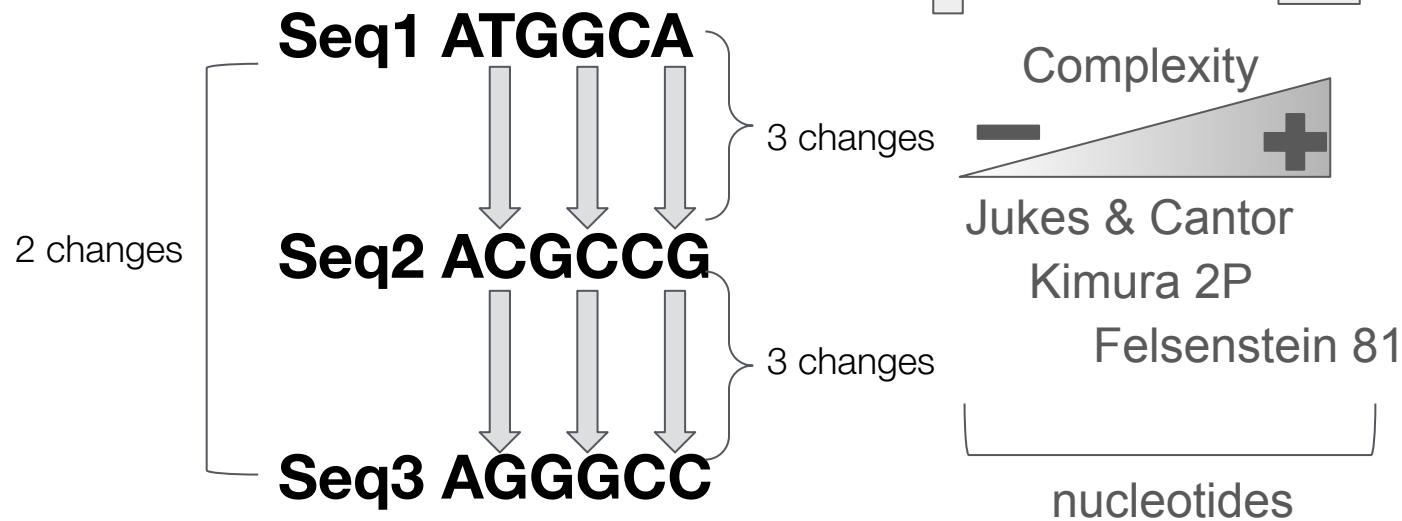
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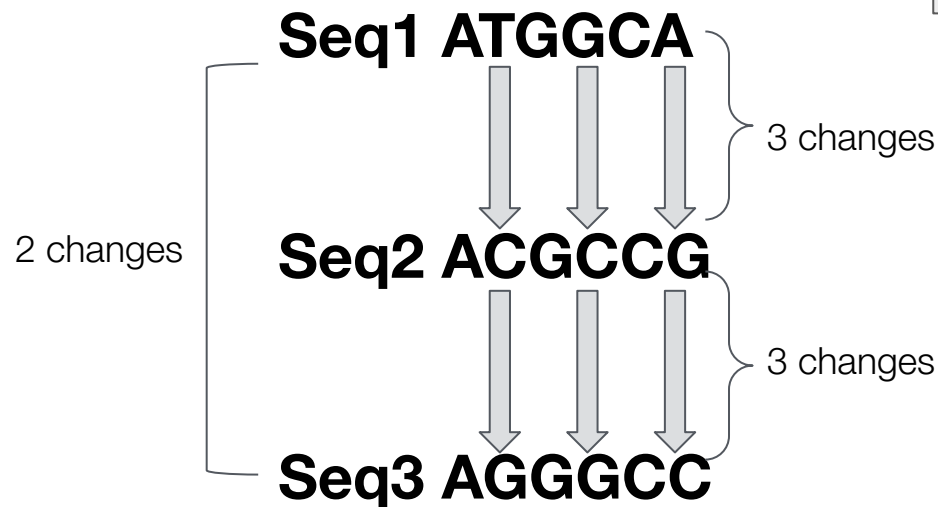
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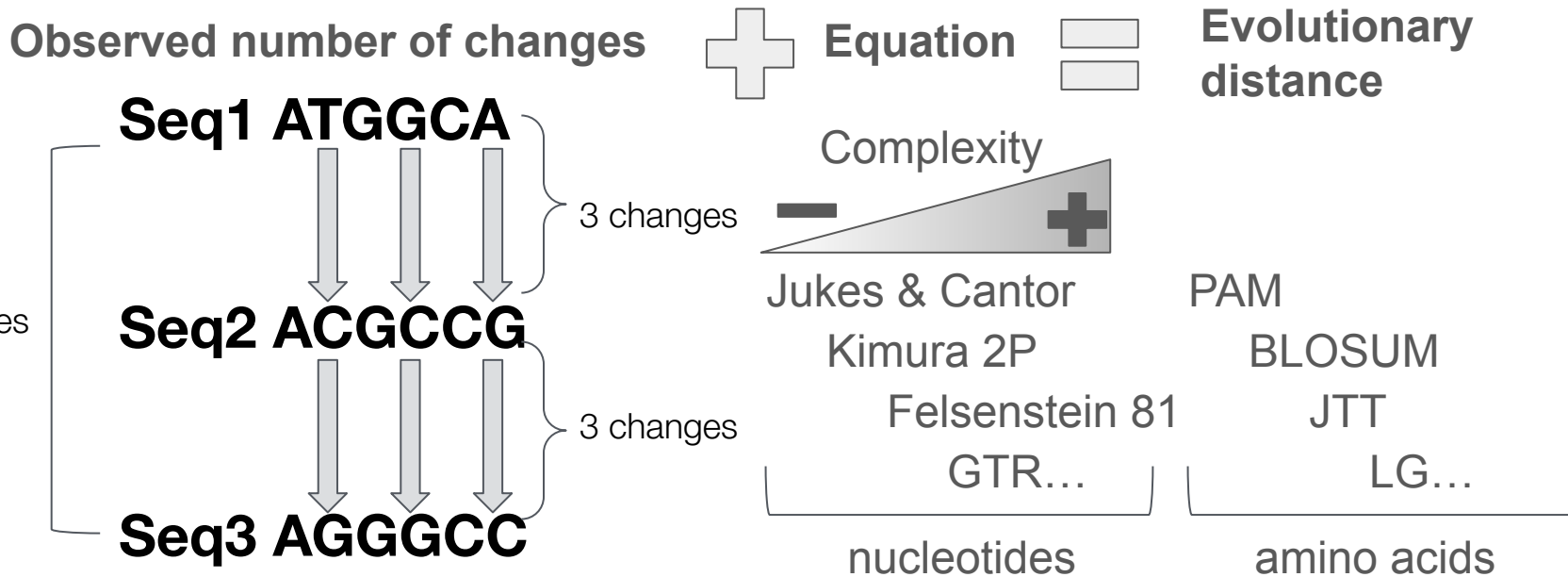
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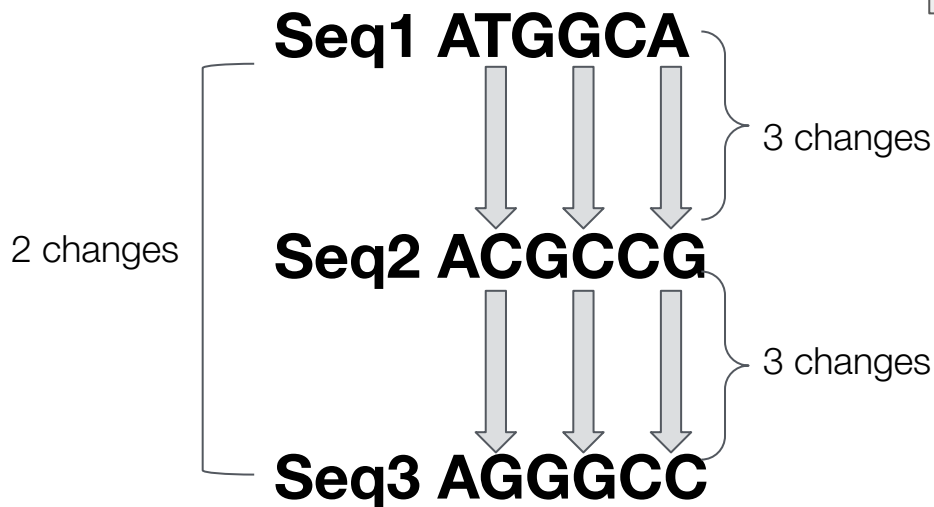
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All models are wrong,  
but some are useful.

George Box, British statistician (1919 – 2013)

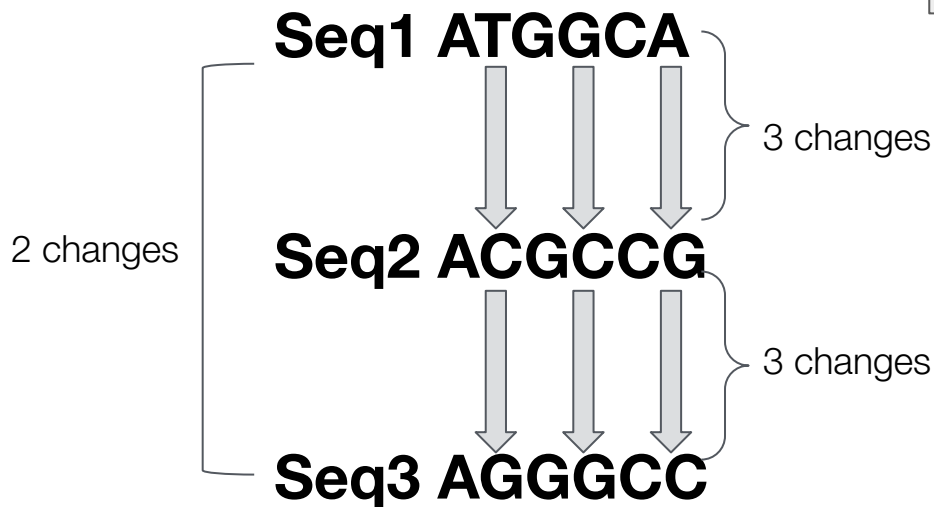
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More about models:

Olivier Gascuel's talk on 25th Jan



# 06 MODEL SELECTION & PHYLOGENETIC INFERENCE



**DATA** + **MODEL OF EVOLUTION**  
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DATA + MODEL OF EVOLUTION  
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Two main methods:

**Maximum Likelihood (ML)** and **Bayesian Inference (BI)**

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*'What is the probability that this model ( $T$ ) is correct, given the data ( $D$ ) that we have observed?'*

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**BI seeks  $P(T|D)$ , while ML maximizes  $P(D|T)$**

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**DATA + MODEL OF EVOLUTION**

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**+ A WAY TO ASSESS HOW GOOD YOUR HYPOTHESIS IS**

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Traditional metrics:

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- [Felsenstein's bootstrap proportion](#) (FBP)
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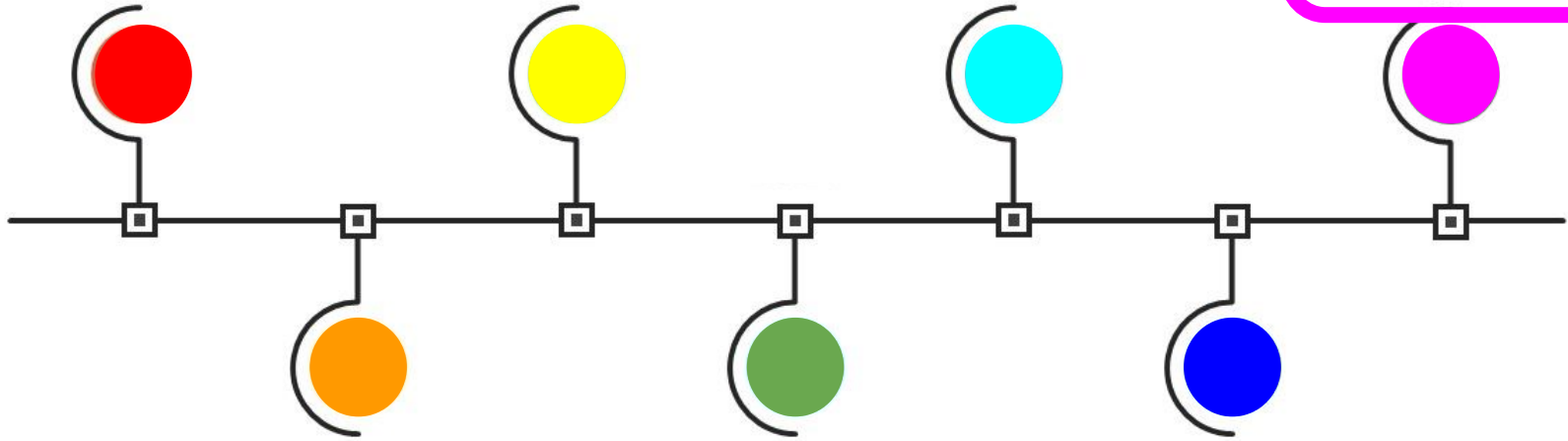
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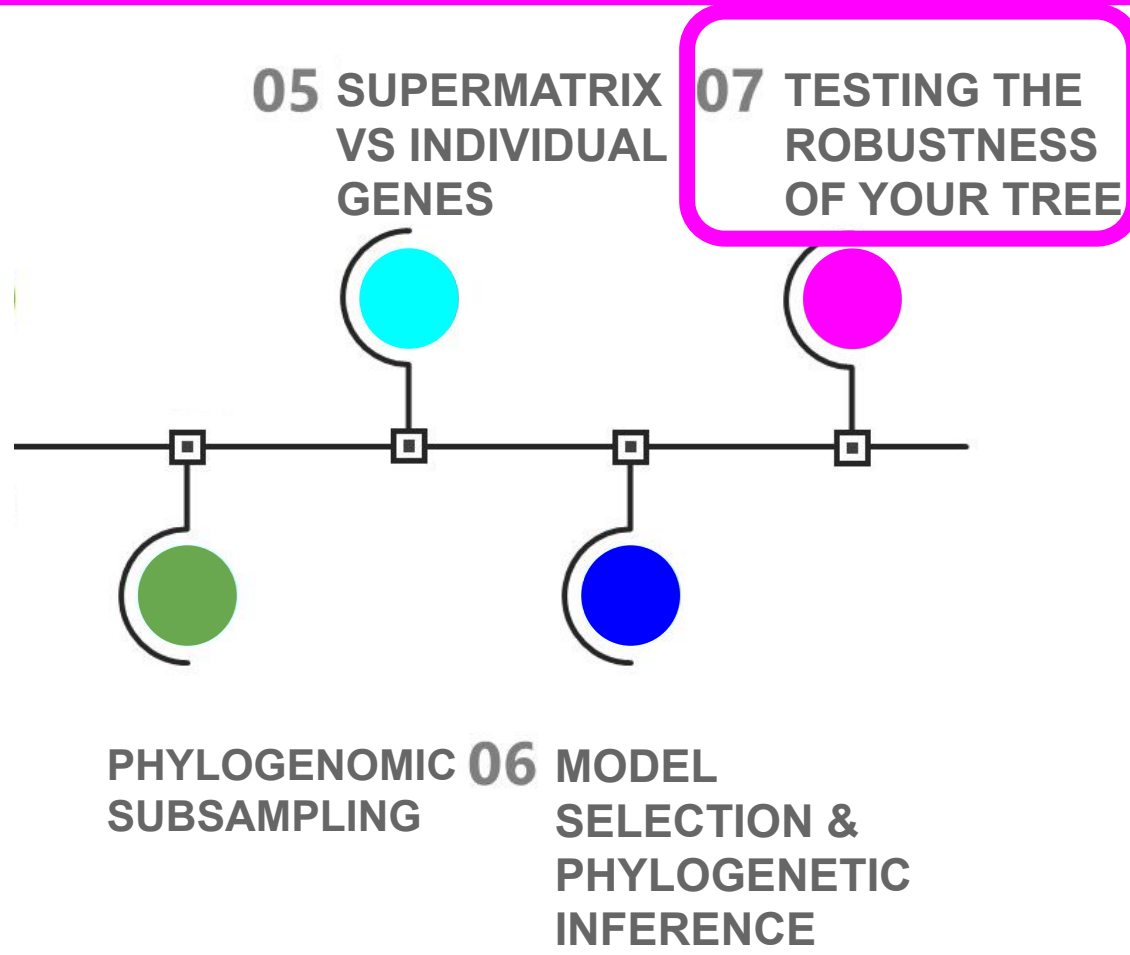
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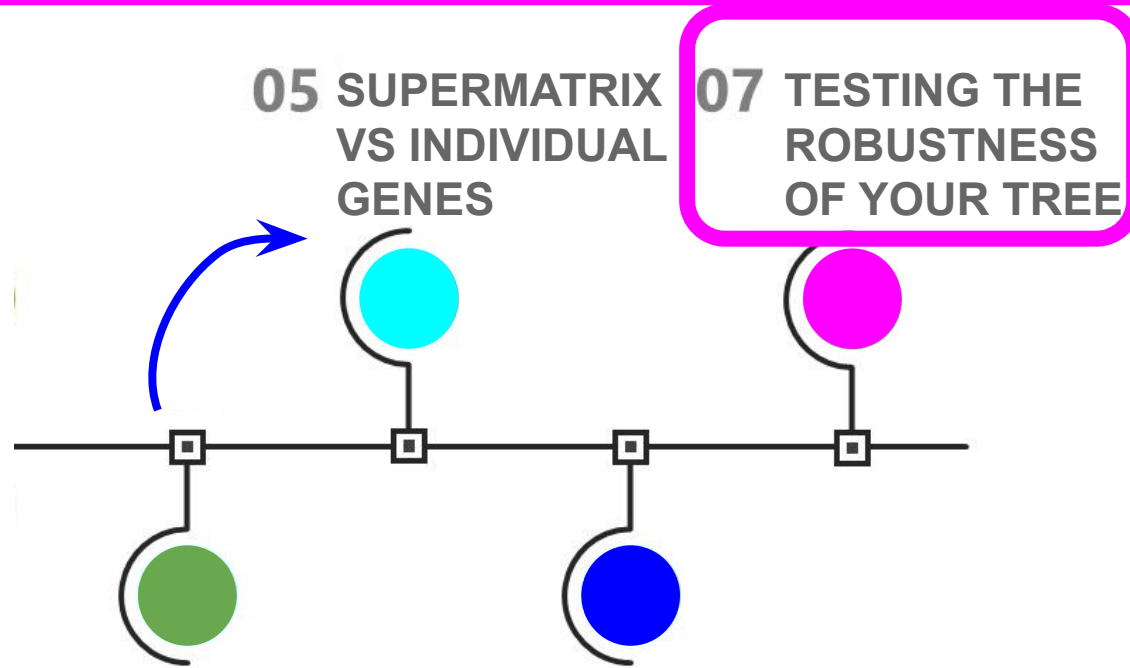


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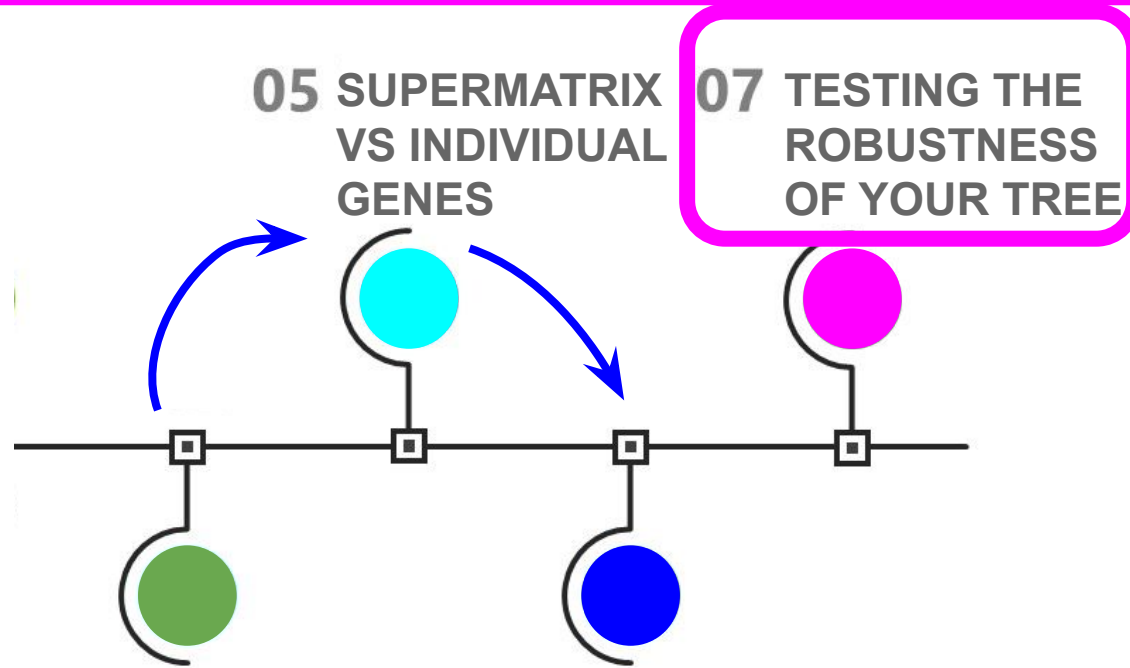
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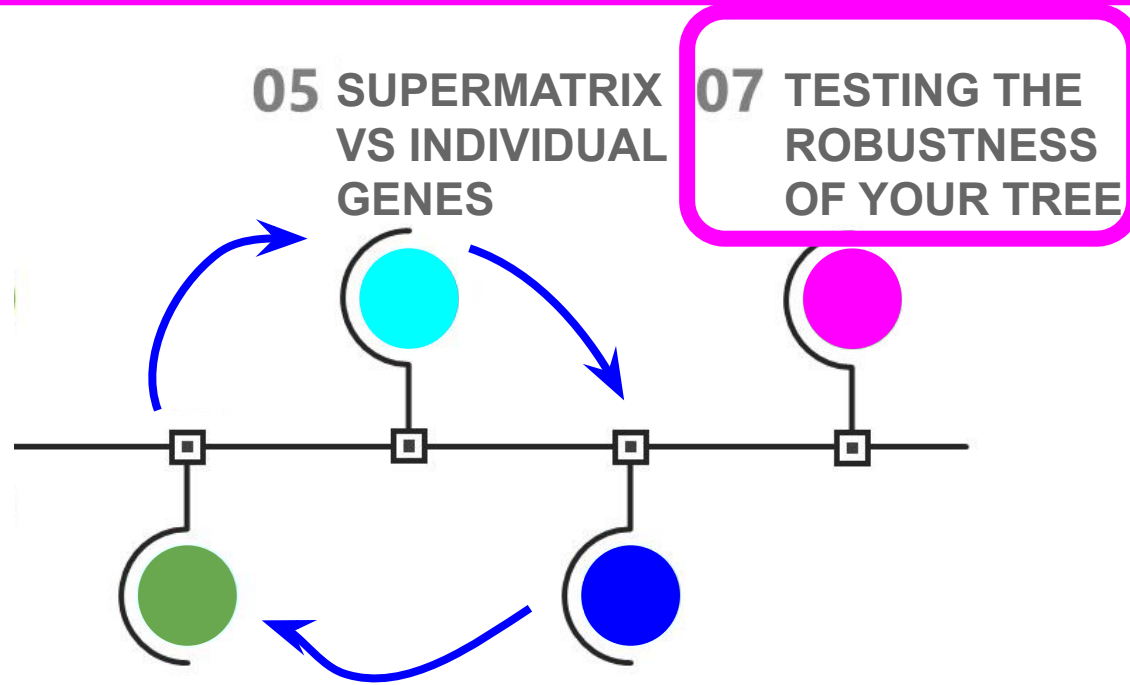
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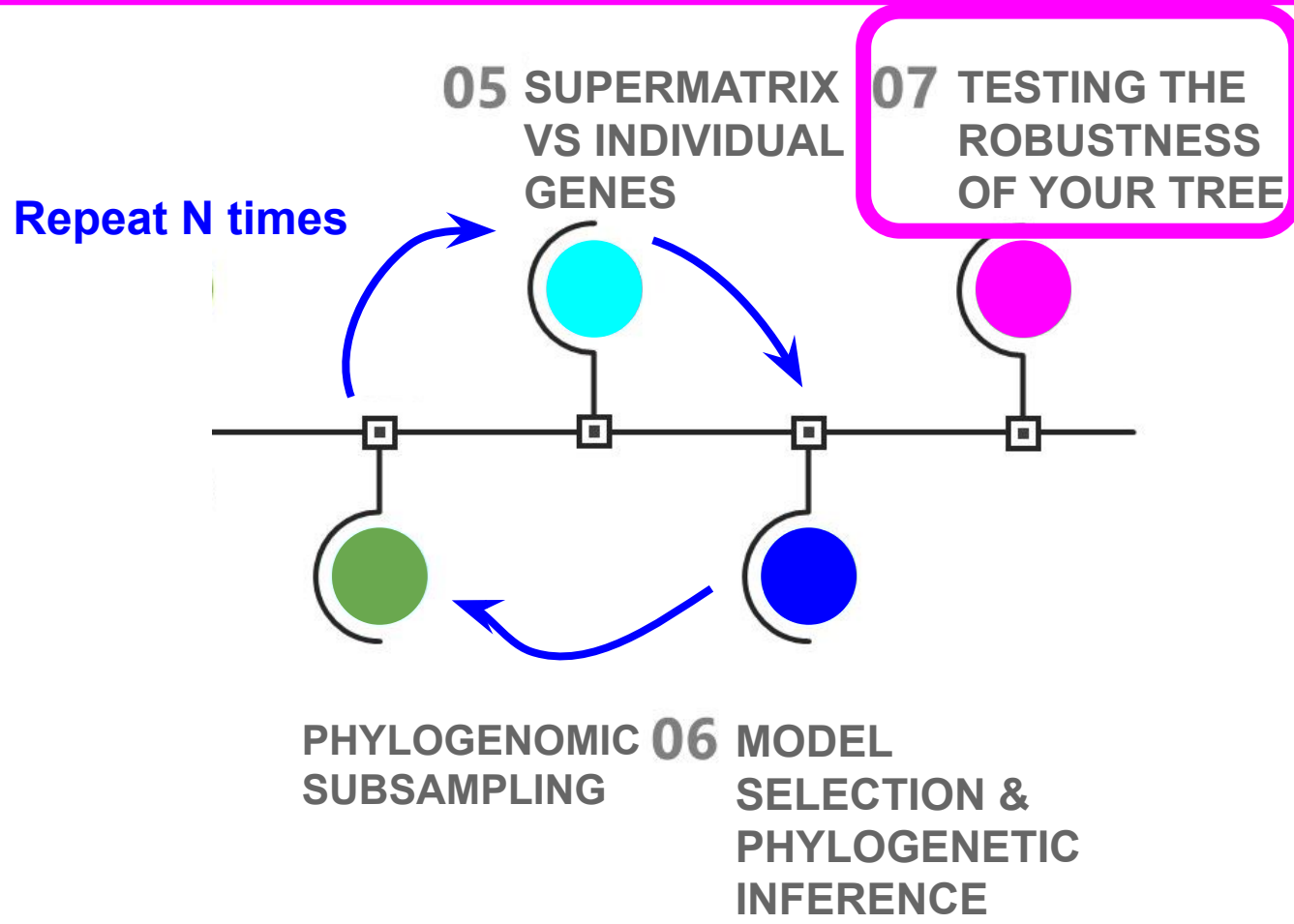


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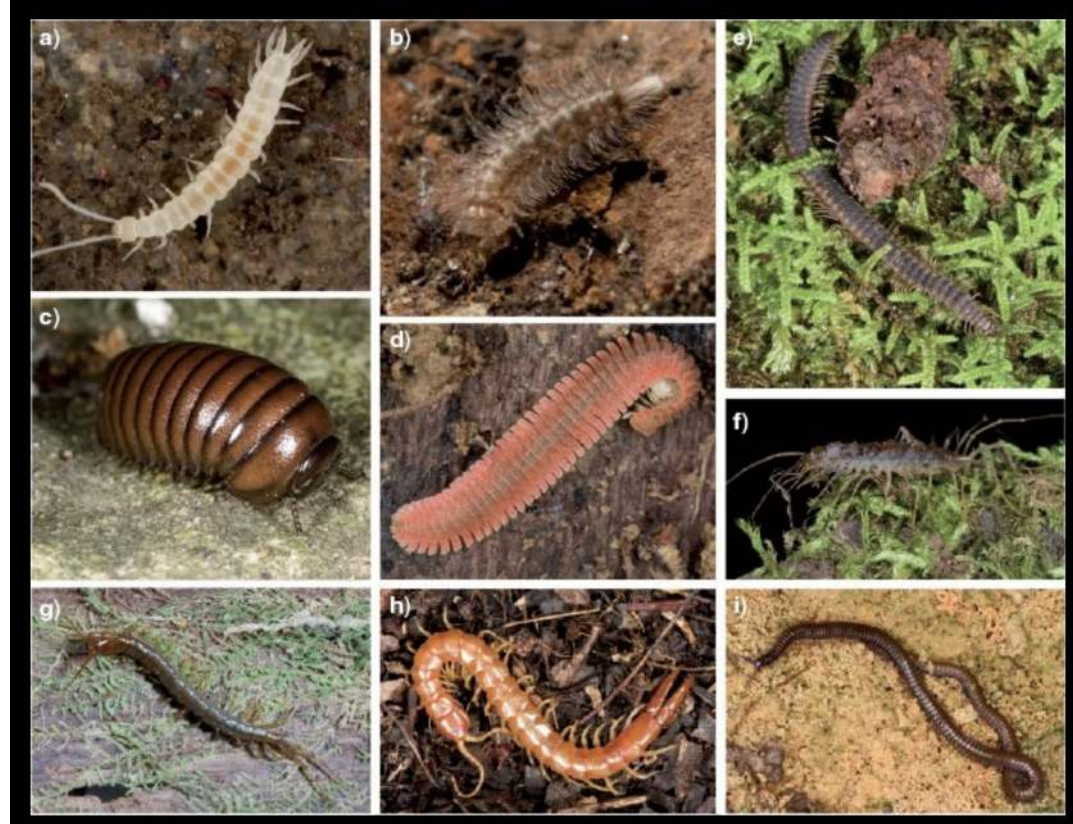


PHYLOGENOMIC 06 MODEL  
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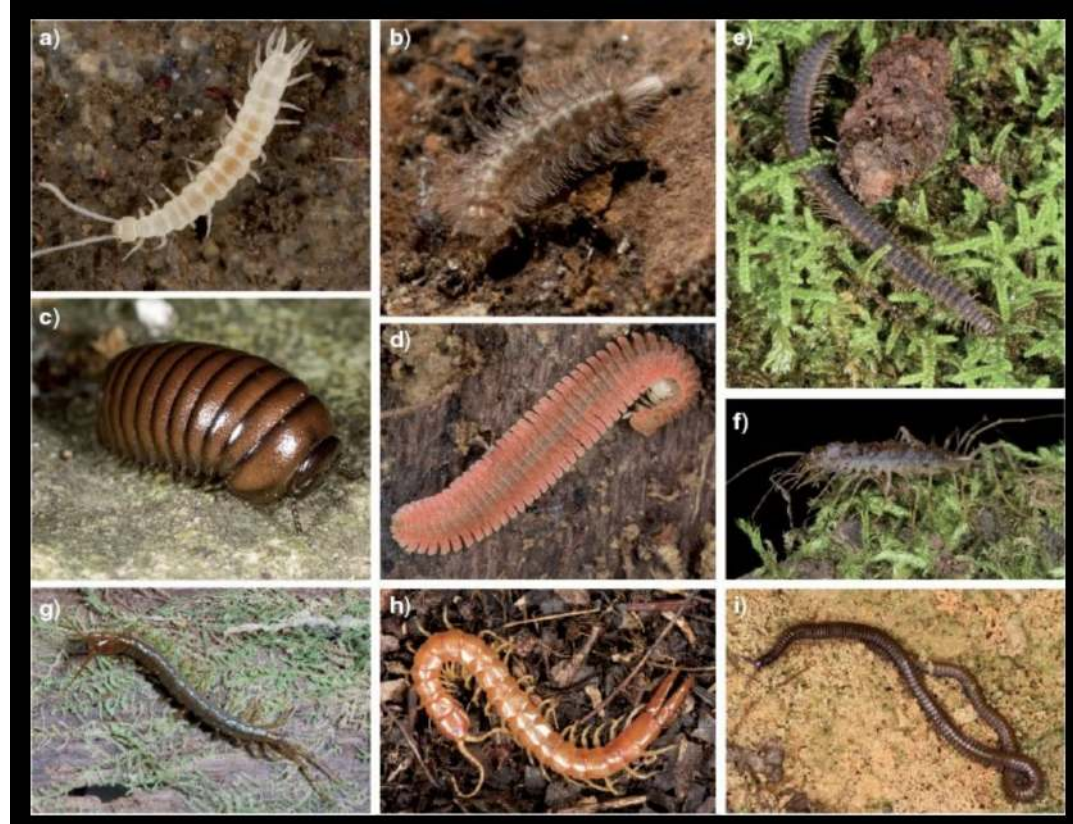
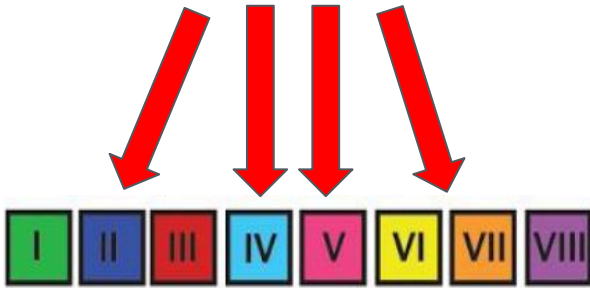


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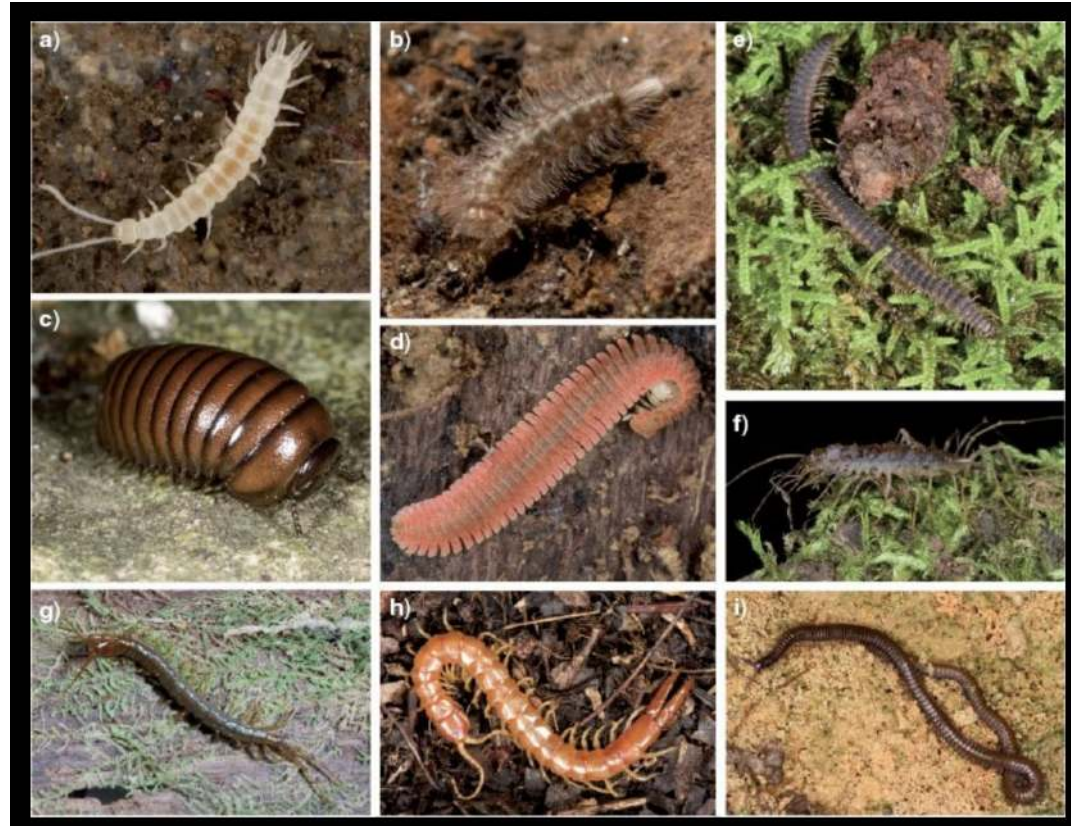
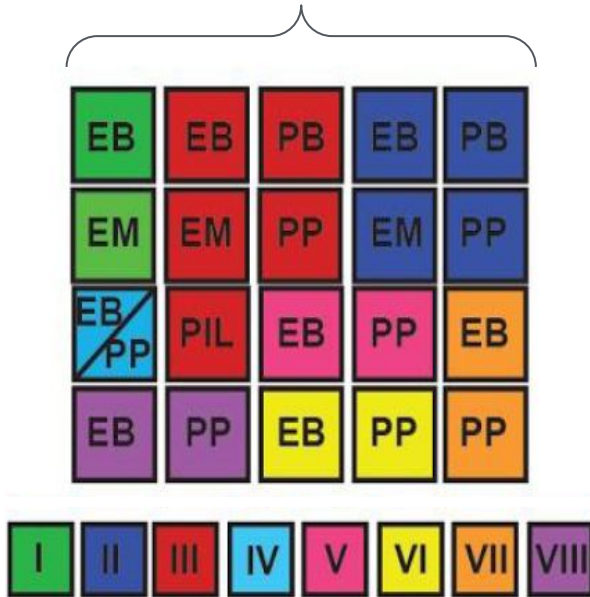
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These are **matrices/subsets**  
of individual gene trees



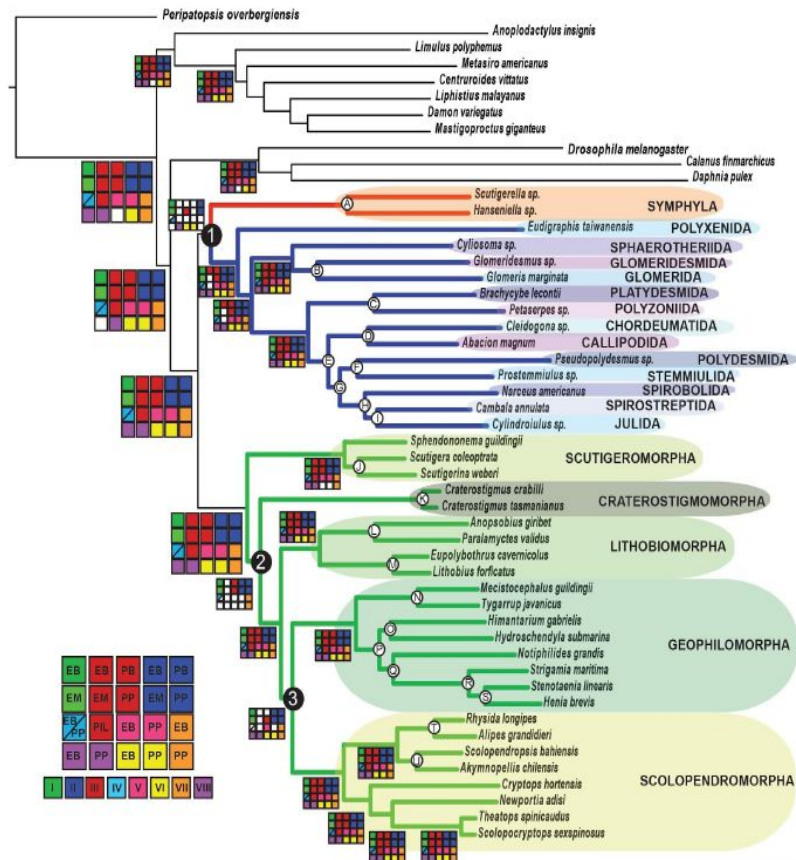
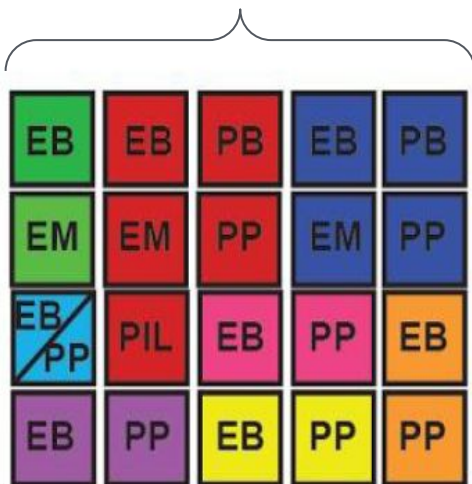
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# AND YOU, HOW IS **YOUR** PROJECT?

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