

A little tour of assembly methods

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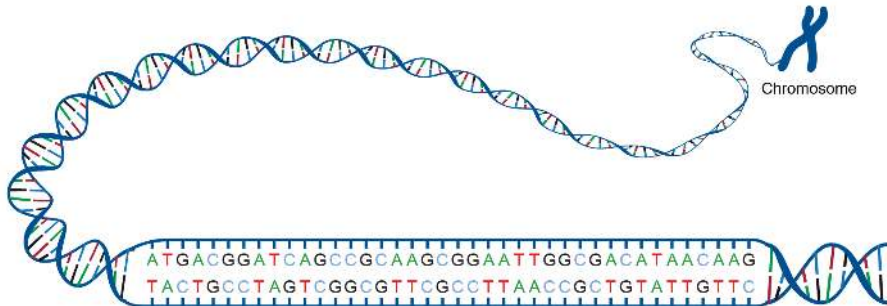
● Content of this course

- How to reconstruct a genome with sequencing data?
- What are the main challenges?
- Which solutions have been proposed?
- Easter egg Warning: 2 assembler names hidden in the slides



genome size: ~ 32 gigabases

- Accessing a genome



From www.genome.gov/genetics-glossary/acgt

- Reads are words from the genome

- Reads are **shuffled** words from the genome

- Genome assembly task

- Using read overlaps

- Genome sequencing: coverage

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- Genome sequencing: coverage

NB: 30-100X are often required for assembly projects

- How to assemble

- Assembly idea number 1: assemble the longest overlaps

- Your time to shine!

- The Greedy solution

- The actual solution

- What happened?

- Do we expect many repeats?

Input interpretation:

$$\left(\frac{4^{31} - 1}{4^{31}} \right)^{1/2} (5 \times 10^6 (5 \times 10^6 - 1))$$

Decimal approximation:

0.999997289498784302383172055421363836712023171938932024106...

From en.wikipedia.org/wiki/Birthday_problem

- The burden of assembly: genomic repeats

- 15: 44,994
- 21: 1,169
- 31: 559
- 41: 323
- 51: 225
- 61: 192

Genomic repeats are NOT random events

• Greedy assemblers

- Simple and efficient scheme
- Rely on **local** best choice (greedy)
- May create errors because of local choices when there are repeats



- History from the last century

- Graph representation

- Assembly **idea number 2: consider all overlaps**

- Greedy solution

- One piece solution

- Multiple repeats

- First solution

- Second solution

- Parsimonious solution: do not assemble

Repeats lead to the fragmentation of the assembly

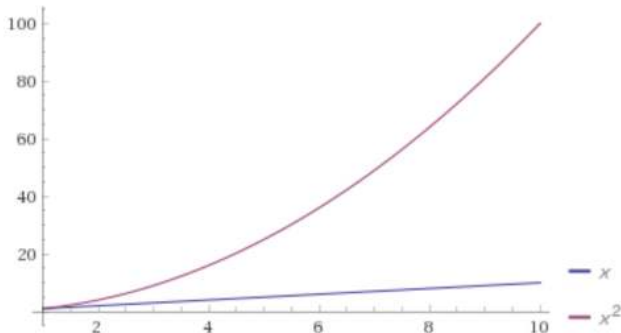
- Missing information also fragments the assembly

- Assembly **concession number 1: output fragments**
In the real world, assemblers often provide pieces of genomes rather than complete ones

- Overlap graph prerequisite : all overlaps

- Overlap graph burden: number of reads

$n(n-1)/2 = \mathcal{O}(n^2)$ possible overlaps for n reads



Linear: 2X data 2X time

Quadratic: 2X data 4X time

- Overlap graph burden: number of reads

$n(n-1)/2 = \mathcal{O}(n^2)$ possible overlaps for n reads

# Reads	# Overlaps
1000	499,500
10,000	50 million
100,000	5 billion
1 million	500 billion
10 million	50 trillion...

Most overlaps are too small to be considered...

The overlap computation is not linear

Talking about CPU years on large genomes...

● Overlap graphs in a nutshell

- Graphs of overlaps between the reads
- Can provide a global solution for assembly
- Can be difficult in real cases because it requires a lot of computation (overlaps)

S. cerevisiae, *D. melanogaster*, human could be assembled using overlap graphs approaches (Celera (Myers et al. 2000), SGA (Simpson & Durbin 2011), ...)

- Fast forward

- Assembly **idea number 3: Focus on genome words**

- Genome words / read words

- Reconstitute larger genomic words

extract all k -mers ($k = 7$):

- The de Bruijn graph

- de Bruijn graph assembly

- de Bruijn graph time!

Hint: Use 7-mers

• Solution

- de Bruijn graph versus overlap graph

- de Bruijn graphs abstract redundancy

- de Bruijn graphs only rely on $k - 1$ overlaps

- de Bruijn graphs limitation

Fixed overlaps

- de Bruijn graphs limitation

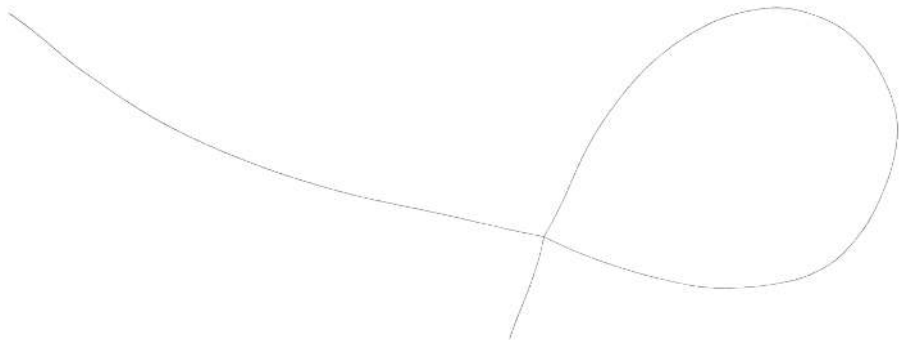
Repeats...

each k -mer appears only once in a de Bruijn graph

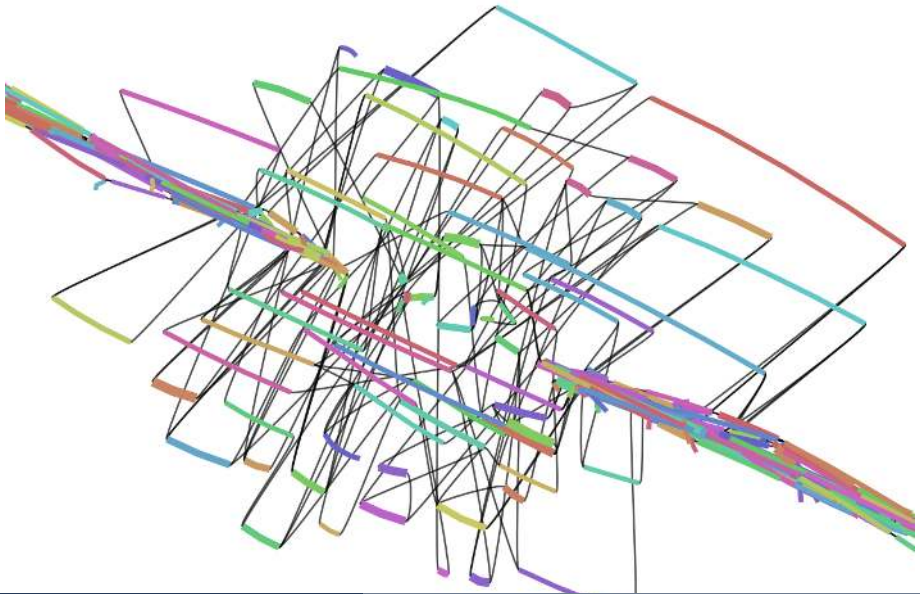
- de Bruijn graph limitation

- On the representation of de Bruijn graphs

- de Bruijn graph on a real dataset



- de Bruijn graph on a real dataset ZOOMED IN

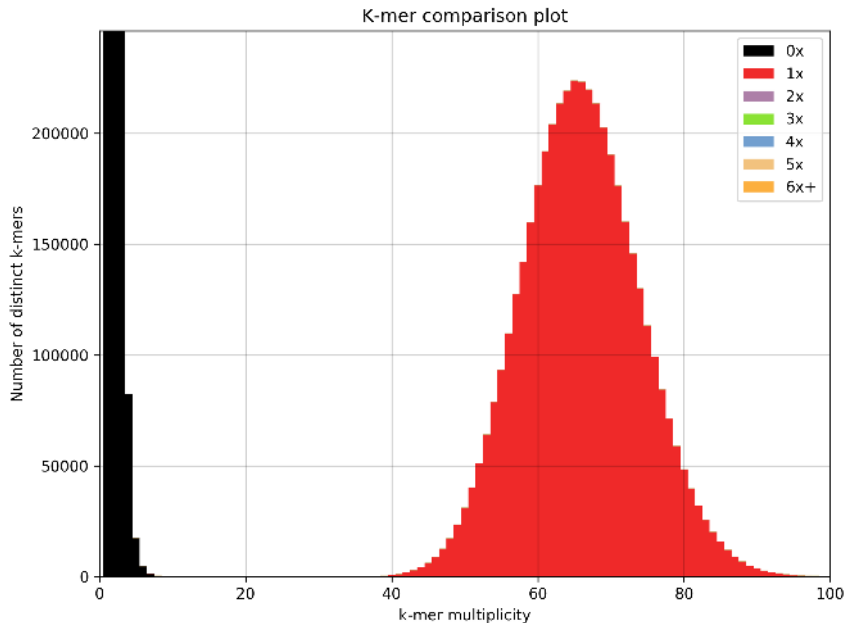


- Sequencing errors

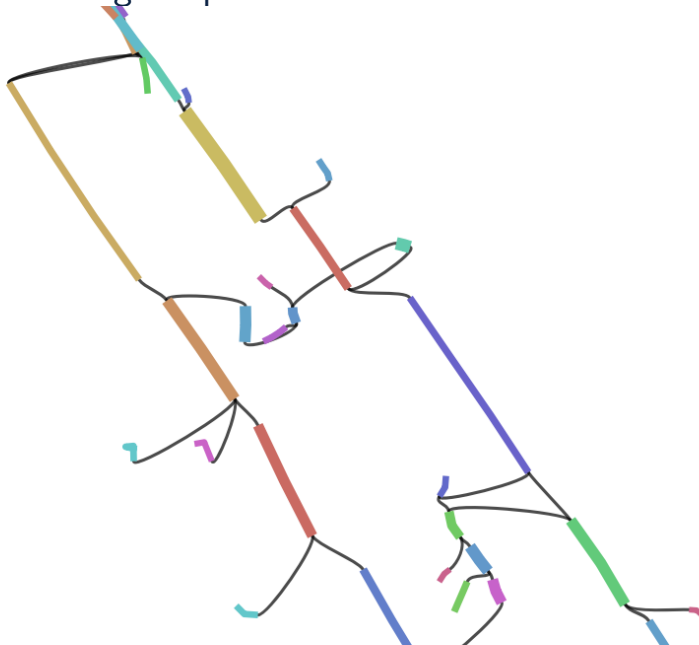
- Erroneous k -mers vs genomic k -mers

Erroneous k -mers are seen less than genomic ones

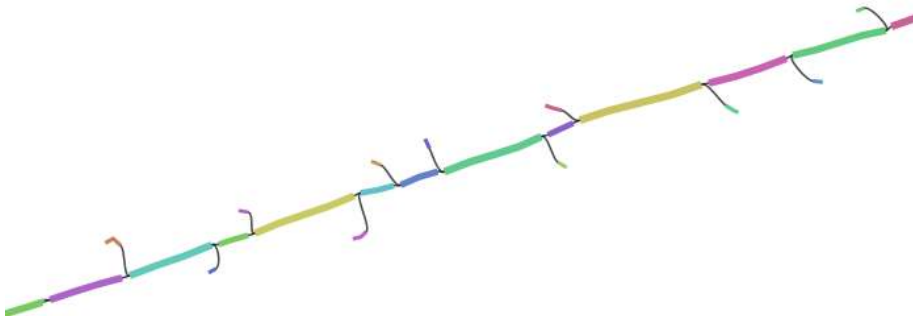
• K -mer histogram



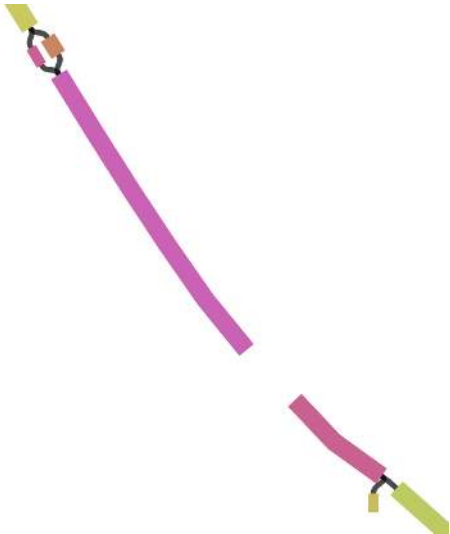
- Removing unique k -mers



- Removing k -mers seen less than 3 times



- Removing k -mers seen less than 4 times

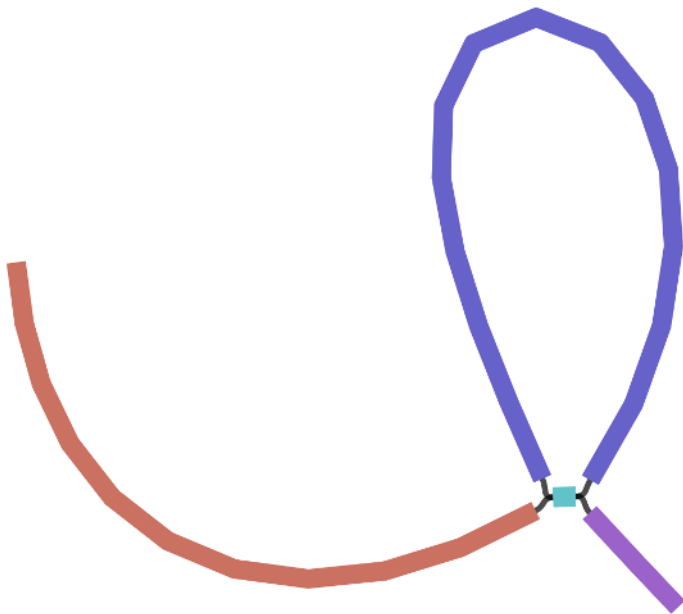


- Errors in de Bruijn graphs

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- Errors in de Bruijn graphs

- (Almost assembled phage !)



- de Bruijn graphs in a nutshell

- Graph of words of size k , $k-1$ overlaps
- Collapses identical k -mers
- Very successful, have replaced the overlap graphs with high throughput sequencing data
- Still outputs fragments of the genome



White spruce, 20 gigabases

- Multiple k assembly

Most de Bruijn graph assemblers can now perform several assemblies with different k -mer sizes to produce an improved "super" assembly

Build DBG with $k=5$ and $k=7$ from those reads

AAAATCGATCTC

TCTCATCGAATT

- Multiple k assembly

Most de Bruijn graph assemblers can now perform several assemblies with different k -mer sizes to produce an improved "super" assembly

- Multiple k assembly

Most de Bruijn graph assemblers can now perform several assemblies with different k -mer sizes to produce an improved "super" assembly

- de Bruijn graph on an eukaryota

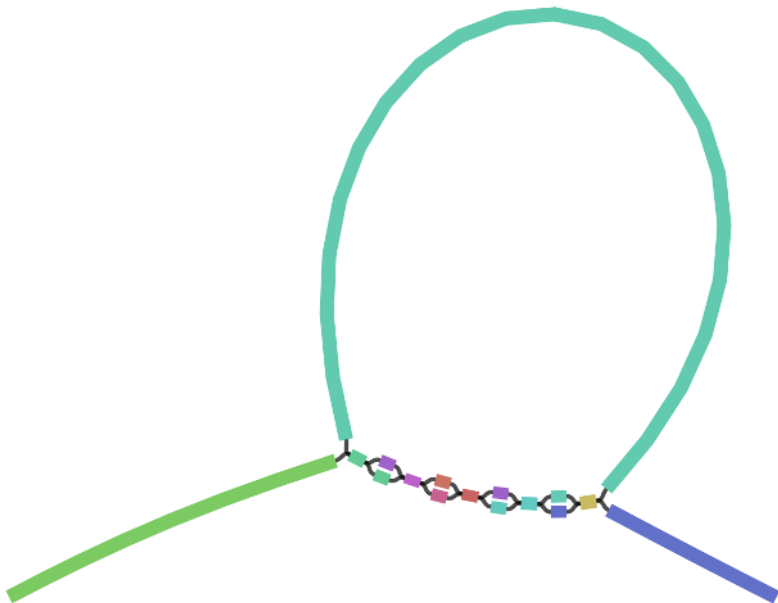


- Two or more genomes per individual

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- Assembly **concession number 2**: collapse variability

- Paralog genes/repeats



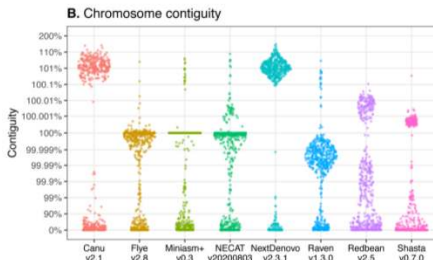
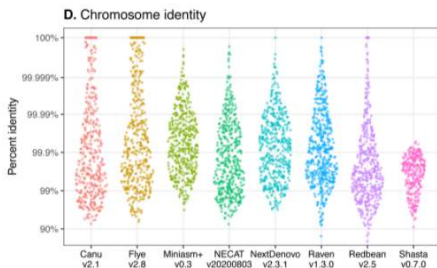
- Paralog genes/repeats in graph

- An assembler is a set of heuristics

- Nodes coverage
- Graph local/global topology
- Reads that can be mapped on nodes
- Estimated coverage/genome size
- ...

● An assembly is a model

- 1 Assemblies contain errors
- 2 Different tools can produce very similar assemblies
- 3 A single tool can produce very different assemblies with small changes of parameters(!)



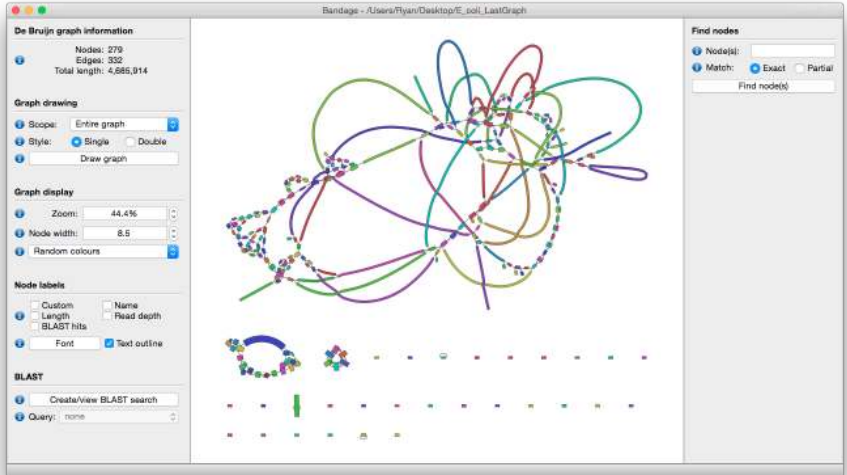
From github.com/rrwick/Long-read-assembler-comparison

- What do we do post-assembly?

- ① Assess its quality
- ② Improve it
- ③ Use it!

• Visualize assembly

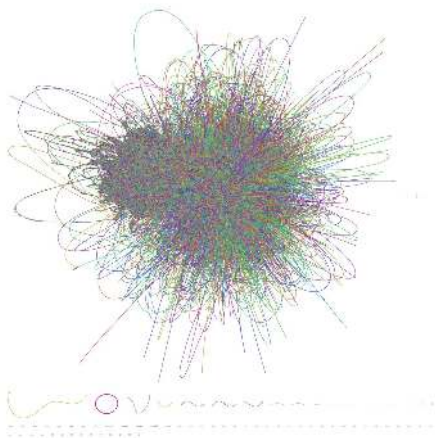
Bandage tool can visualize assembly graphs (GFA)



From [rwick.github.io/Bandage](https://github.com/rwick/Bandage)

- Visualize assembly

Bandage tool can visualize assembly graphs (GFA)



- Assembly continuity

N50

N50 can be described as a weighted median statistic such that 50% of the entire assembly is contained in contigs or scaffolds equal to or larger than this value.

Example: 1 Mbp genome

50%



- Assembly continuity

N50

N50 can be described as a weighted median statistic such that 50% of the entire assembly is contained in contigs or scaffolds equal to or larger than this value.

N75

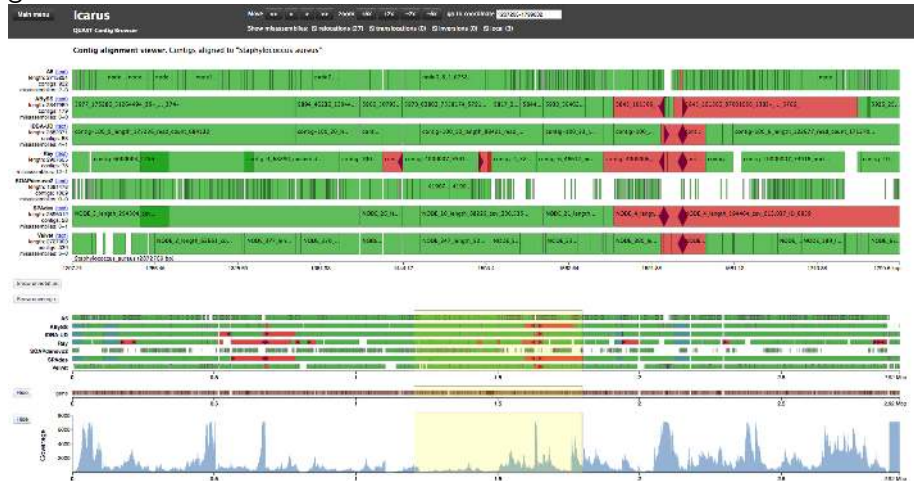
N75 is the same statistic for 75% of the assembly

NG50

Similar to the N50 but only takes into account contigs/scaffolds that can be **aligned** on the reference genome and consider 50% of the **genome size** instead of the assembly size

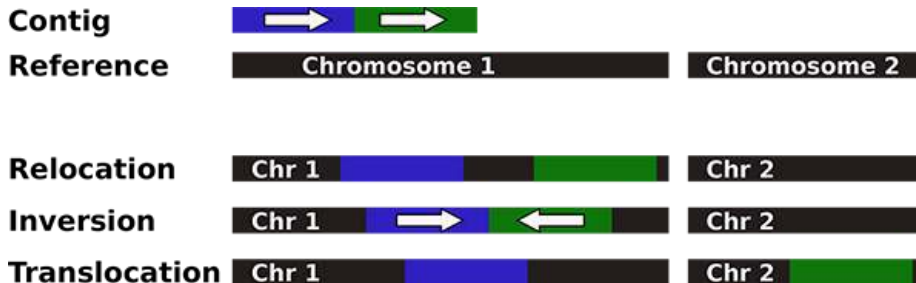
● Evaluate assembly

Contigs can be mapped and compared to a reference/closely related genome



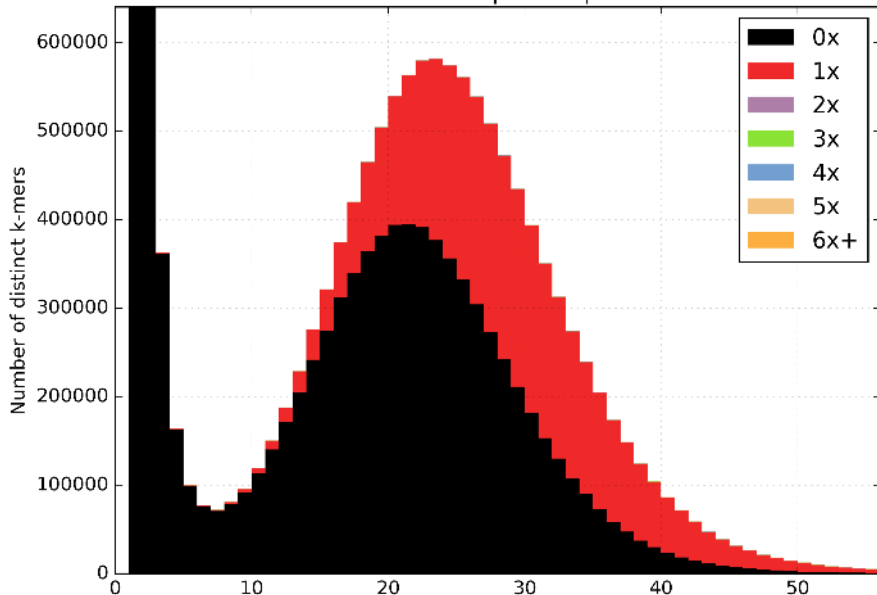
From quast.bioinf.spbau.ru/manual.html

● Misassemblies

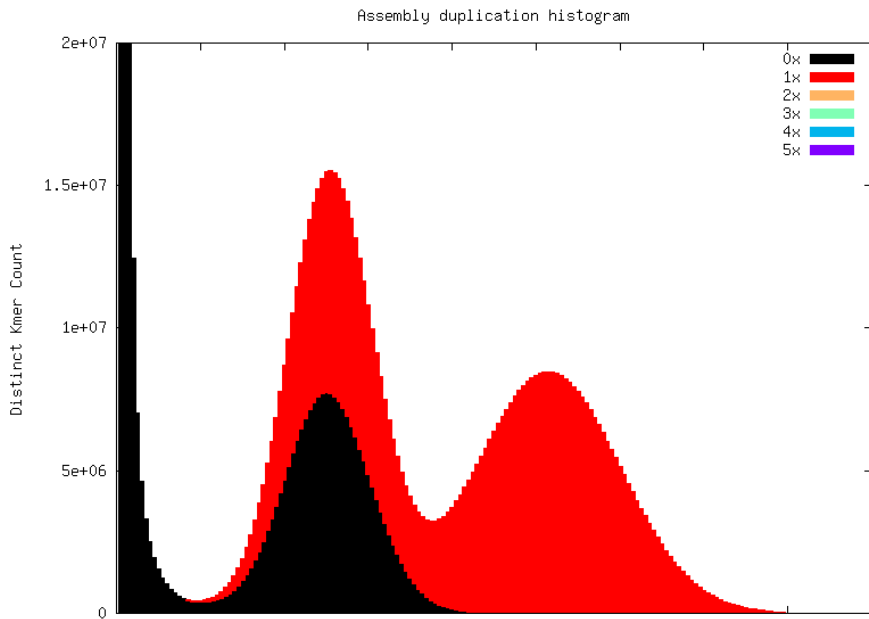


● K -mer spectrum visualization with KAT

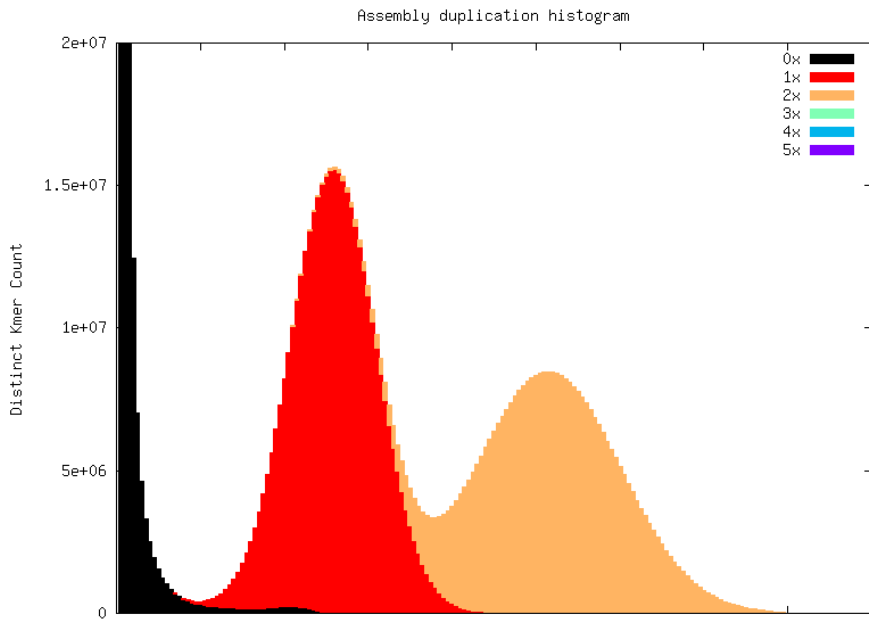
K-mer comparison plot



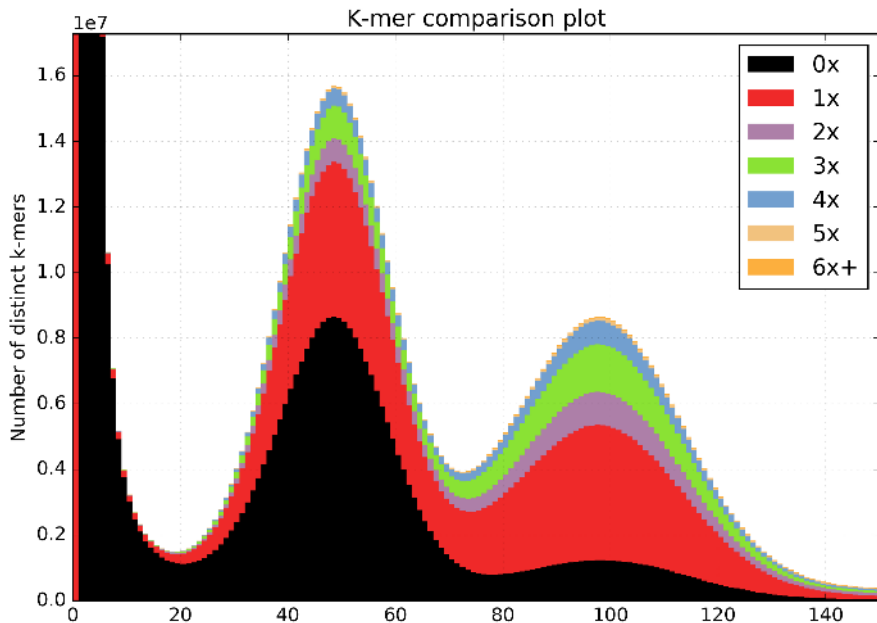
● *K*-mer spectrum visualization with KAT



● *K*-mer spectrum visualization with KAT

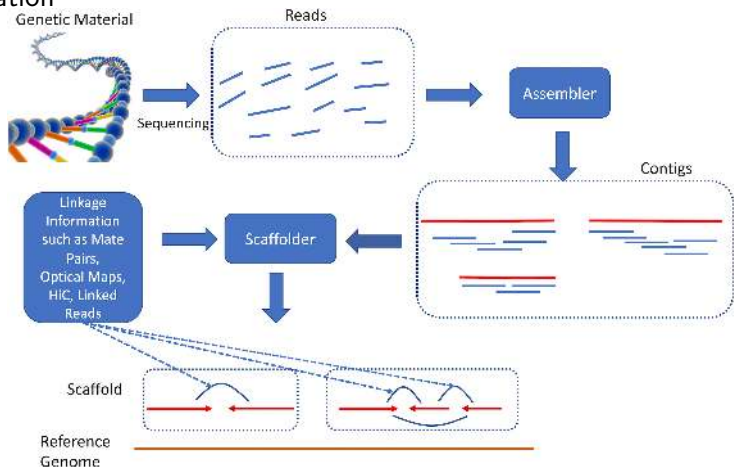


● K -mer spectrum visualization with KAT



● Scaffolding

Softwares can improve the assembly continuity by using other kinds of information



From "Modern technologies and algorithms for scaffolding assembled genomes" Plos Computational Biology

● The end

... of the theoretical part



- Intermission

• Sanger



- Medium reads $\approx 1000bp$
- Very low error rate $\approx 0.01\%$
- low throughput (up to billion of reads per run)
- Costly (\$500/Mb)

No longer used for assembly

• Second generation sequencing



NextSeq Series 



HiSeq 4000 System



HiSeq X Series[†]



NovaSeq 6000
System

- Short reads $\approx 150bp$
- Low error rate $< 1\%$
- High throughput (up to billion of reads per run)
- Cheap (\$0.50/Mb)
- GC bias

Mainly assembled using de Bruijn graphs

• State-of-the-art

- SPAdes [Bankevich 2012]
- Megahit [Li 2015]
- IDBA [Peng 2012]



- SGA [Simpson 2012]
- Discover denovo [Weisenfeld 2014]
- Abyss [Simpson 2009]

- Third generation sequencing



- Long reads $\approx 10 - 100\text{kbp}$
- High error rate $\approx 10\%$
- High throughput (up to millions of reads per run)

• Nanopore VS Pacbio

- Portable
- Ultra long reads (100kbases, some reads reach the megabase level)
- Mostly deletions

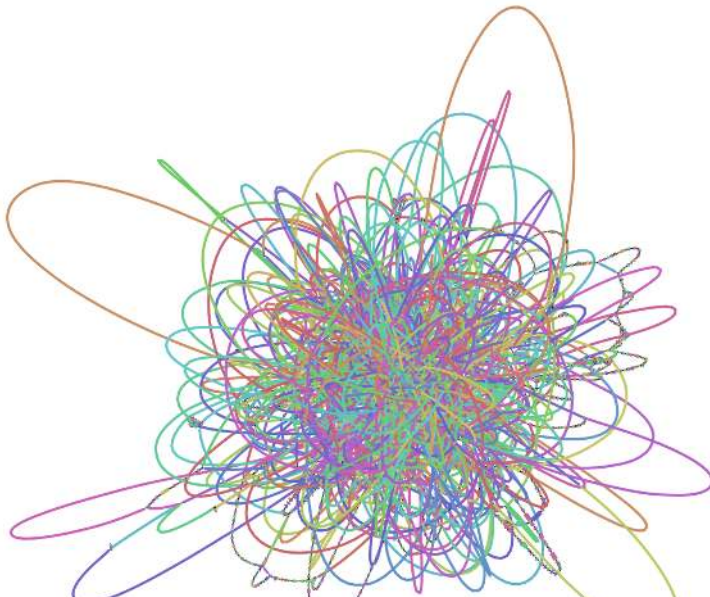
- More mature
- HiFi reads (99.9% identity)
- Mostly insertions

- Repeats spanning

- Repeats spanning

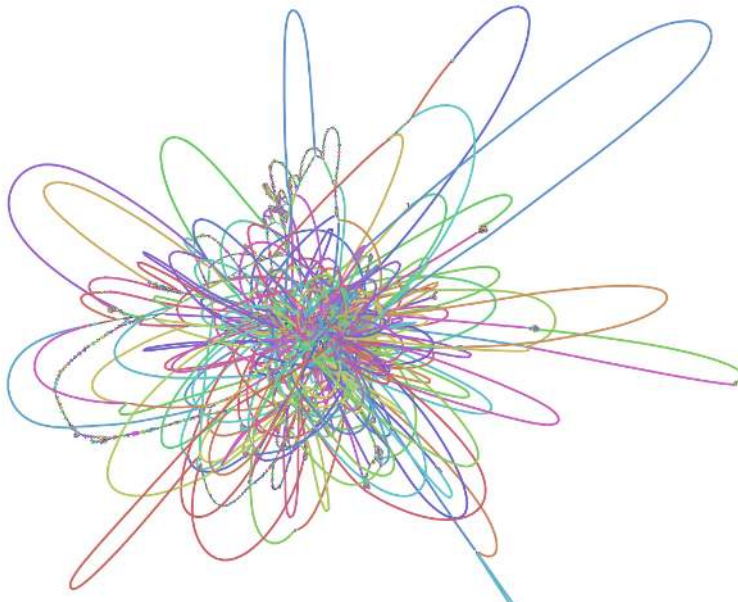
- Read length matter

Read size=21



- Read length matter

Read size=31



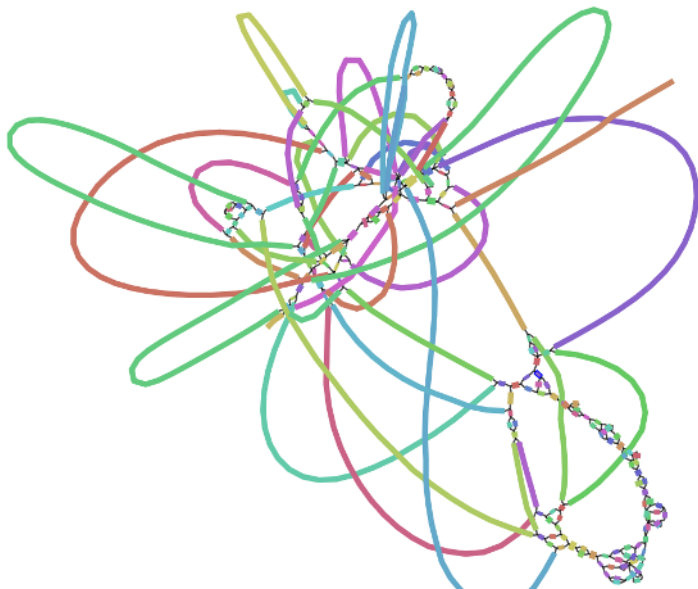
- Read length matter

Read size=63



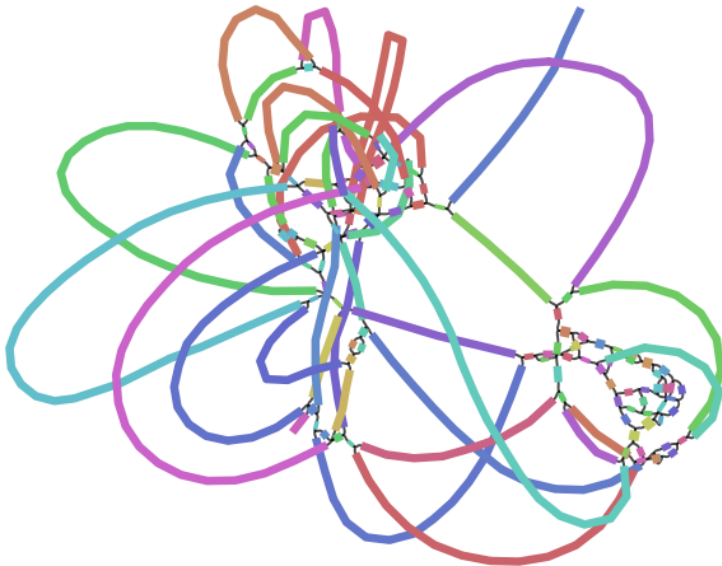
● Read length matter

Read size=255



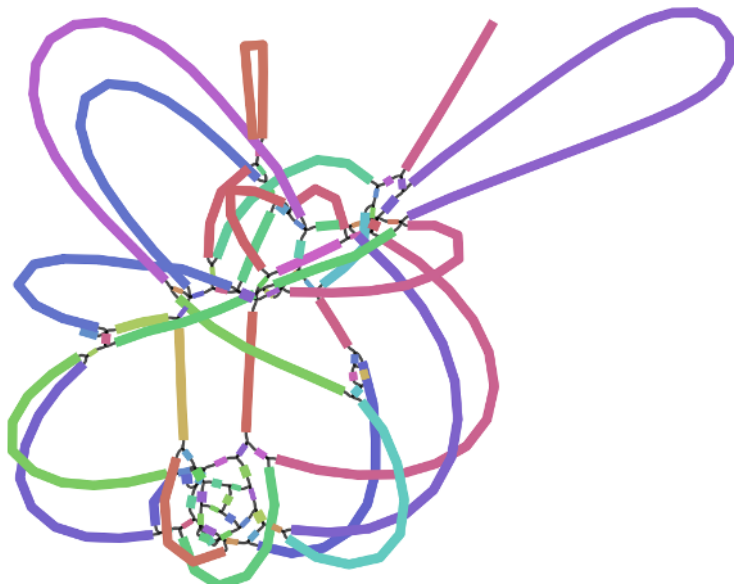
● Read length matter

Read size=500



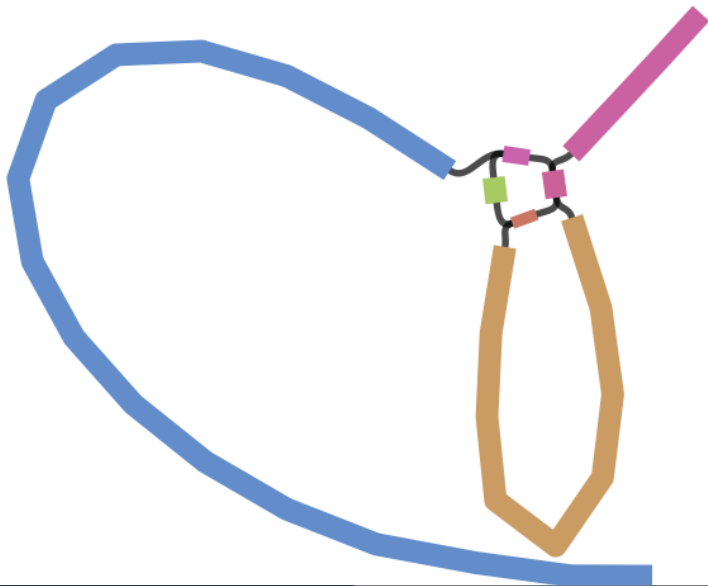
● Read length matter

Read size=1000

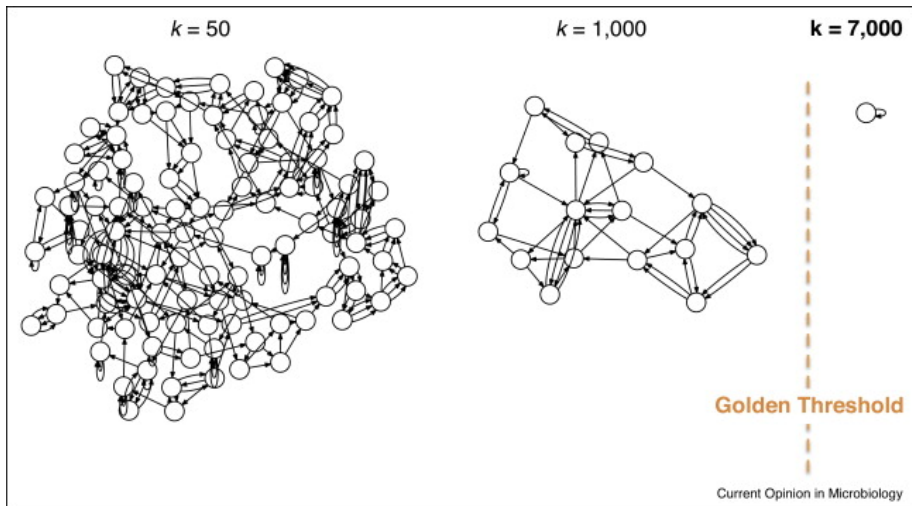


● Read length matter

Read size=2000



- Great hope for assembly



From "One chromosome, one contig: complete microbial genomes from long-read sequencing and assembly" Current Opinion in Microbiology 2015

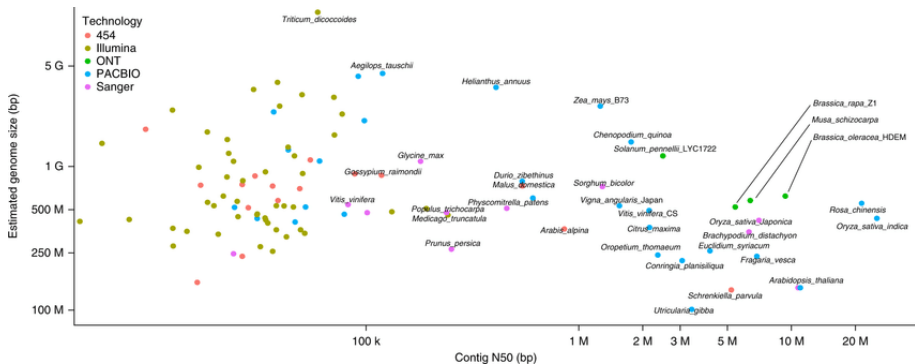
- Long reads killed the assembly star



Laura Landweber @LandweberLab · Jan 2

Our newest version of Oxytricha's somatic genome is out (rdcu.be/bZNfC) and has 18,617 distinct chromosomes. That's 2000 more than we previously published in doi.org/10.1371/journal.pgen.1004000. PacBio captured most chromosomes in single reads: Genome sequence, No assembly required

Great hope for assembly



From "Chromosome-scale assemblies of plant genomes using nanopore long reads and optical maps" Nature Plants 2018

● Which assembly strategy is best suited?

- Long reads $\approx 10kbp$
- High error rate $\approx 10\%$
- High throughput (up to millions of reads per run)

Based on long reads properties, which assembly solution would you choose and why?

Vote!

- Greedy
- Overlap graph
- de Bruijn graph

- Long reads for assembly: de Bruijn graph?

- Long reads for assembly: overlap graph?

Supposed to be super expensive!

• Longer reads, better overlaps

- Less reads for the same coverage
- Larger overlaps

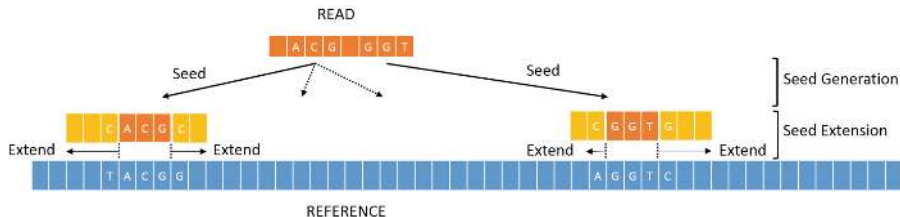
5Mb bacteria example with 100X coverage

- 5 million 100bp reads
- 99 bp average overlap

- 50,000 10kbp reads
- 9,900 bp average overlap

- 5,000 100kbp reads
- 99,000 bp average overlap

- Are large overlaps hard to compute?



Aligning very long and highly erroneous regions is expected to be expensive, as alignment is quadratic $\approx \mathcal{O}(n^2)$!

- "Anchor chaining" in overlap graph

- For long reads: typically Minimap's [Li 2016] job
- "Anchor chaining": find common chains of anchors (k -mers) in the same order in 2 sequences (can be **linear** in practice in most cases)

- Long reads for assembly: overlap graphs

- Sequencing errors

Insertion and deletion made calling almost impossible

- Using coverage to remove noise: Consensus

- Exercise: Perform a consensus

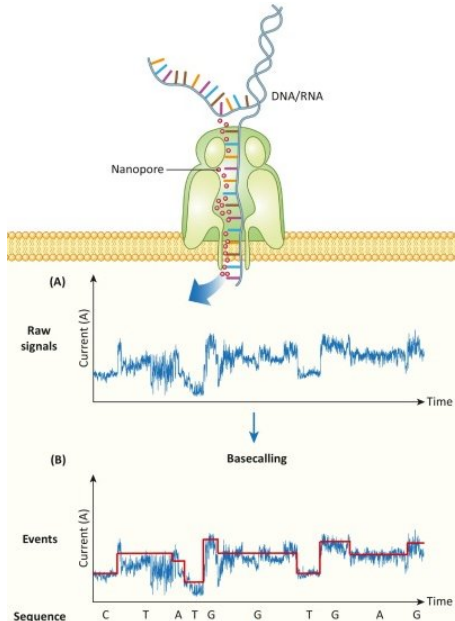
- Exercise: Perform a consensus - solution

- Consensus during assembly

- Consensus after assembly: polishing

- Consensus after assembly: polishing

- Homopolymers are hard to read



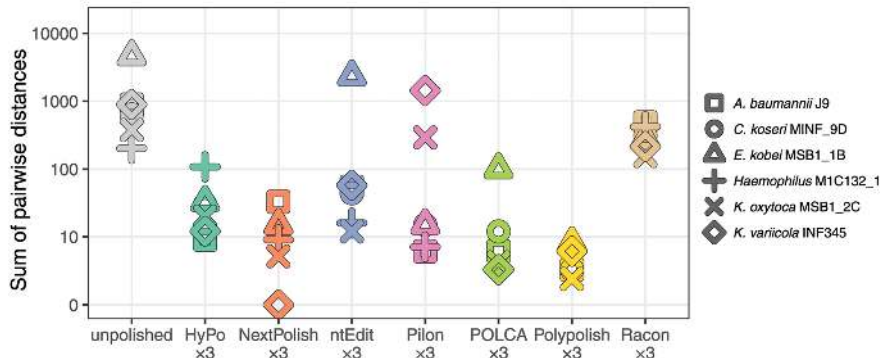
- Polishing using accurate reads

• Systematics errors

Polishing with Illumina data can improve the final error rate

A. Single-tool short-read polishing

ALE change:	0	110696	113366	87707	113056	113061	115623	82446
total distance:	7635	212	74	2519	1775	128	28	1867

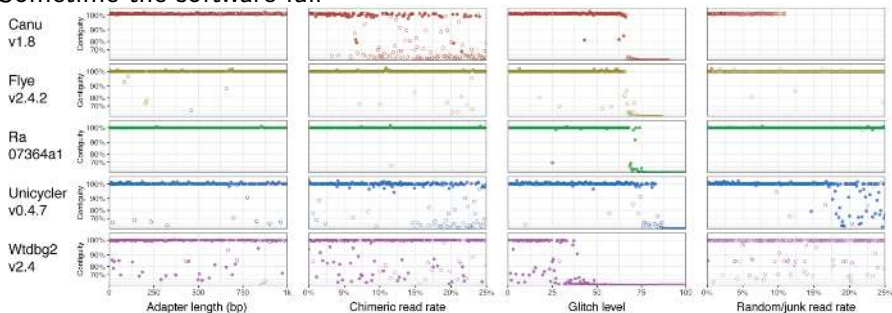


From Polypolish: Short-read polishing of long-read bacterial genome assemblies

● Long reads for assembly: assembly solved?

Assembly is not solved yet

Sometime the software fail



From github.com/rrwick/Long-read-assembler-comparison

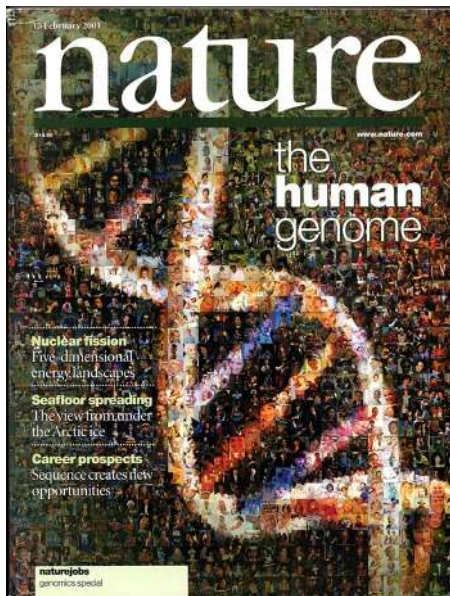
- Long reads for assembly: assembly solved?

Assembly is not solved yet

Sometimes the data cannot solve the problem

- Very large repeated region
- Low local coverage
- Chimeric/noisy reads

● 20 years later



● Telomere-to-Telomere consortium

Has produced in 2021 a complete human genome with one contig per chromosomes !

- 30x PacBio HiFi
- 120x coverage of Oxford Nanopore (ultra long reads)
- 70x PacBio CLR
- 10X Genomics, BioNano DLS and Arima Genomics HiC
- 100 authors from 50 labs

- Long reads assemblers

- Flye (Repeat graph) [Kolmogorov et al 2019]
- Raven (OLC) [Vaser et al 2021]
- NECAT/MECAT(OLC) [Xiao et al 2017]

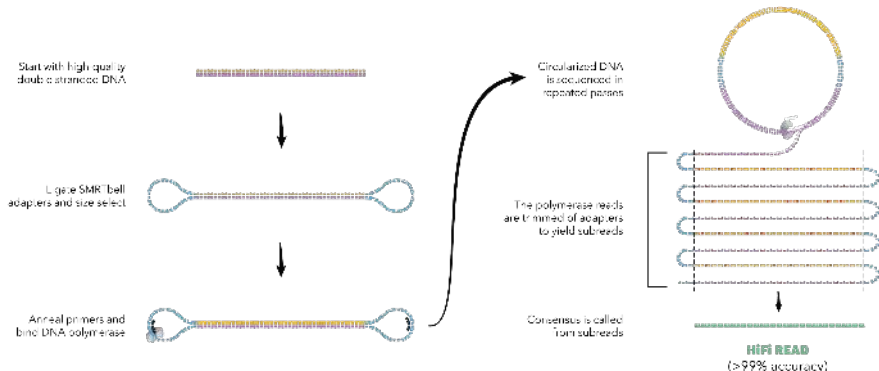
- Canu (Greedy) [Koren et al 2017]
- Shasta (OLC) [Saffin et al 2020]
- Redbean (fuzzy de Bruijn graph) [Ruan 2019]
- ...

- Long read assembly summary

- Overlap graphs with quick overlap computation
- Long reads can span repeats and improve assemblies
- Methods to polish contigs



● Consensus during sequencing



Stands for "High Fidelity"

Very low error rates $\approx 0.1\%$ 0.01%

Almost only homopolymer errors remain

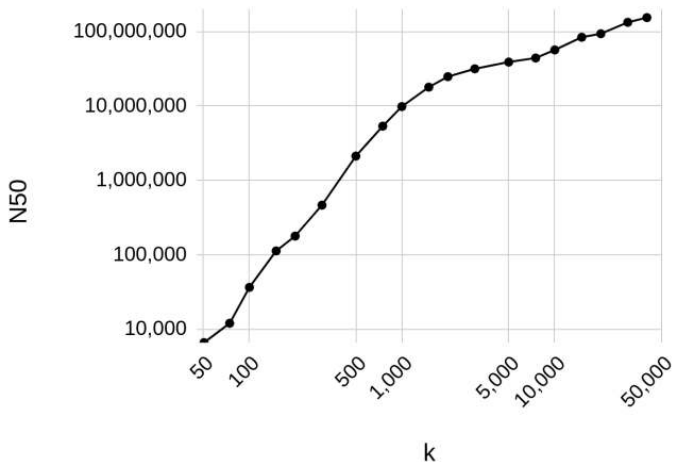
● HiFi Assembly

With almost error-less long reads we have several promising improvements ahead:

- Use de Bruijn graph (more efficient data structures)
- Assemble large genomes very fast
- Perform diploid assembly

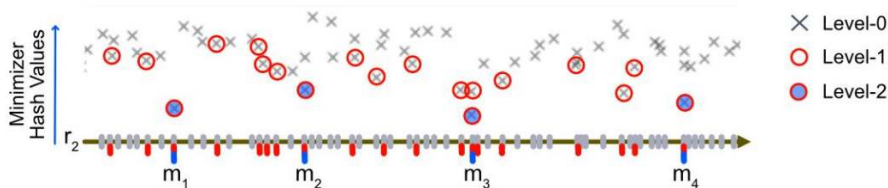
• de Bruijn graph Assembly

Using $K=500$ and $K=5000$ de Bruijn graphs to assemble

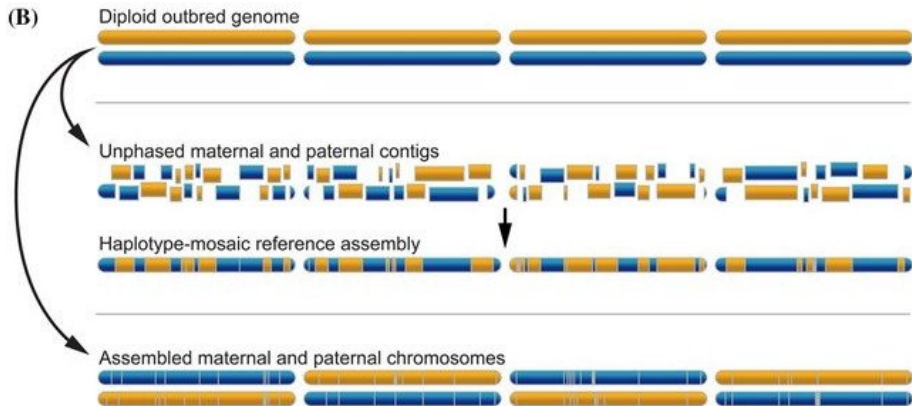


- Very fast genome assembly

Human genome assembled within 2 hours (Peregrine assembler) and 10 minutes (RMBG assembler)

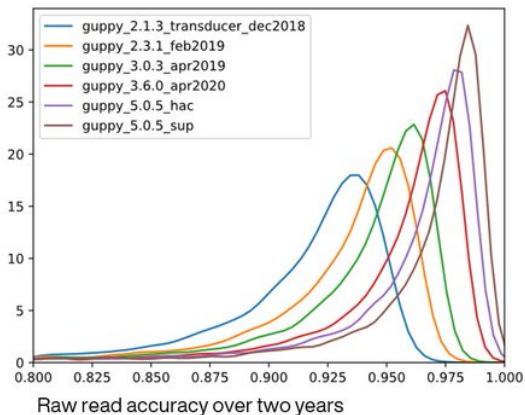


• Diploid assembly



● Ongoing progress

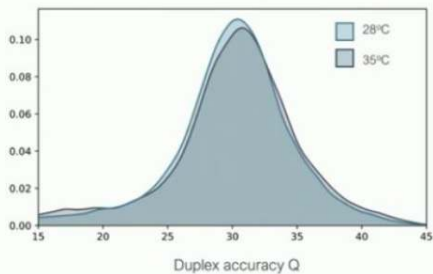
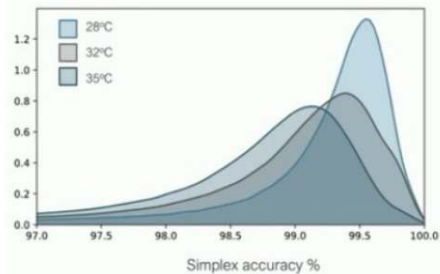
Errors in Nanopore sequencing data are rapidly diminishing



Q20 chemistry achieved modal accuracy $> 99\%$

- High fidelity nanopore incoming?

Nanopore duplex reads could deliver long and precise reads in the future



- The end



- Take home messages

- Short reads: de Bruijn graphs / Long reads: Overlap graph
- Repeats are the core issue
- Output fragments of genomes (**contigs**)
- Several parameters and heuristics used in practice

● On going work

- Reconstruct haplotypes
- Scaling on large genomes
- Robustness to noisy data
- Repetitive regions

- The end



PopGenGoogling
@popgengoogleing



i trust you to figure out your own genome

[Traduire le Tweet](#)

3:01 AM · 14 déc. 2019 · [Twitter for iPhone](#)

37 Retweets **267** J'aime
